

**MOLECULAR DOCKING IN DRUG DISCOVERY: A REVIEW ON ANTI-SNAKE VENOM DEVELOPMENT**

**Adewunmi Rofiat Funmilola**

Department of Biochemistry, University of Maiduguri, Borno state, Nigeria.

**Gidado Abubakar**

Department of Biochemistry, University of Maiduguri, Borno state, Nigeria.

**Zanna Hassan**

Department of Biochemistry, University of Maiduguri, Borno state, Nigeria.

**Abstract**

Snakebite is a frequent accident faced by rural community's dwellers, and it has remained a neglected public health problem in many countries. Snake venom is a complex mixture of proteins, and they participate to envenomation through a diverse array of bioactivities, such as bleeding, inflammation, and pain, cytotoxic, cardiotoxic or neurotoxic effects. The only approved and accepted treatment for snakebite envenoming is the use of antivenoms produced by the purification of IgG immunoglobulins immunized against specific snake venom. However, various technological approaches are being pursued by different research groups, including the use of small-molecule inhibitors, antibody-based bio-therapeutics and peptide-based aptamer against enzymatic toxins and non-enzymatic toxins in snake venom. Modern bioinformatics tools have been recently developed to mine snake venoms, helping focus experimental research on the most potentially interesting toxins. Some computational techniques predict toxin molecular targets, and the binding mode to these targets. This review presents molecular docking studies of potential targeted key enzymes in snake venom.

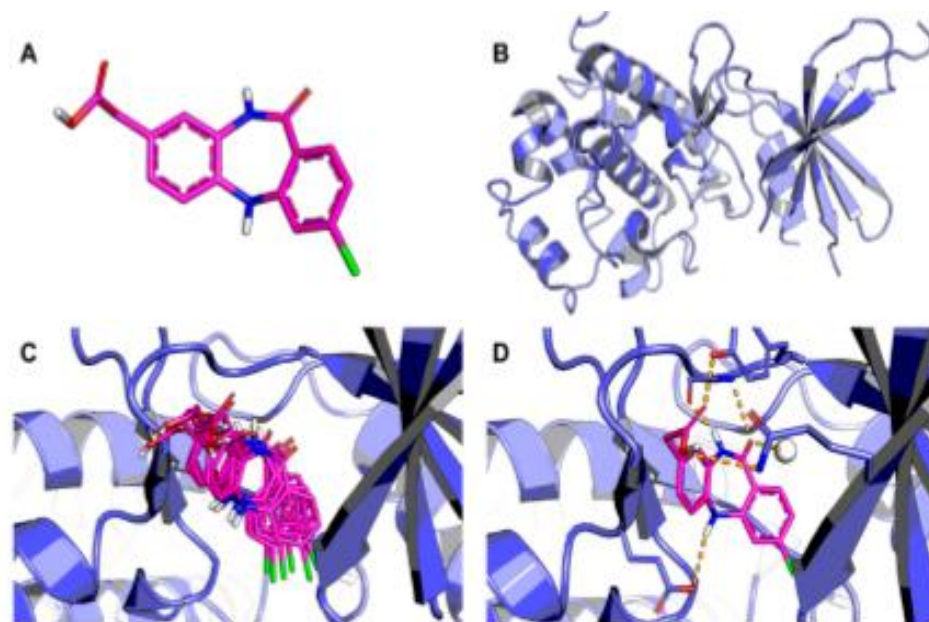
**Keywords: Molecular docking, Drug discovery, Snake venom, bioinformatics, envenomation**

## Introduction

Snakebite is a frequent accident faced by villagers, and has remained a neglected public health problem in many countries even though it is difficult to be precise about the actual number of cases (Reid, 1972). In sub-Saharan Africa alone, snakebite is estimated to cause between 435,000 to 580,000 envenomation and 20,000 to 32,000 deaths every year (Gutierrez *et al.*, 2006). Amputation and disability, tetanus, gangrene, cortical necrosis of the kidneys etc., are among the medical manifestations of snakebite envenomation (Gutiérrez *et al.*, 2017). The five families of poisonous snakes are Colubridae, Elapidae, Hydrophidae, Viperidae and Ataspidae. Snake venom is a complex mixture of different enzymes, which are proteins (Theakston and Reid, 1983). These enzymes determine the toxicity of the snake venom as to whether it is haemotoxic or neurotoxic but the venoms cannot be classified as being exclusively haemotoxic or neurotoxic. Prompt administration of antivenom is the cornerstone of effective snakebite management, although supportive care is crucial too, including assisted ventilation in case of neurotoxic envenomation (Opadijo and Omotosho, 1996). The path to drug discovery is a long, expensive, challenging and arduous task, hence, computer aided drug design and discovery (CADD) is a rapidly growing area that has seen many successes in the last few years (Gutiérrez *et al.*, 2011). Computational drug discovery can help in identifying potent drug molecules and targets via bioinformatics tools, such as molecular docking. They can also be used to evaluate the target structures for possible binding/active sites, generate active drug molecules, check for their dynamic and kinetic properties (Kitchen *et al.*, 2004). Docking is frequently used to predict the binding orientation of small molecule drug candidates to their protein targets in order to predict the affinity and activity of the small molecule. Hence docking plays an important role in the rational drug design (Gore and Desai, 2014).

## Molecular docking

Molecular docking is the process that involves placing molecules in appropriate configurations to interact with a receptor (Ballante, 2018). Molecular docking is one of the most frequently used methods in structural based drug design (SBDD) because of its ability to predict, with a substantial degree of accuracy, the conformation of small-molecule ligands within the Molecules appropriate target binding site (Ballante and Marshall, 2016).



*Fig 1: Outline of the molecular docking process. (A) Three-dimensional structure of the ligand; (B) Three-dimensional structure of the receptor; (C) The ligand is docked into the binding cavity of the receptor and the putative conformations are explored; (D) The most likely binding confirmation and the corresponding intermolecular interactions are identified (Leonardo et al., 2015).*

### **Molecular Docking models**

Over the years, biochemists have developed numerous models to capture the key elements of the molecular recognition process. The models are:

- I. The Lock and Key Theory: As far back as 1890, Emil Fischer proposed a model called the "lock-and-key model, where a substrate fits into the active site of a macromolecule, just like a key fits into a lock. Biological 'locks' have unique stereochemical features that are necessary to their function(Monika et al., 2010).
  
- II. The Induced-Fit Theory: In 1958, Daniel Koshland introduced the "induced-fit theory". The basic idea is that in the recognition process, both ligand and target mutually adapt to each other through small conformational changes until an optimal fit is achieved(Avinashet al., 2013)

## **Application of Molecular Docking in Drug Development**

Drug discovery can be described as the process of identifying chemical entities that have the potential to become therapeutic agents (Keskin *et al.*, 2007). The process of drug discovery is time consuming, tedious and expensive. Therefore, Molecular docking has become one of the most important modeling tools in modern drug discovery because is a very convenient and cheap means to study protein-ligand interactions, which can be used to develop more potent, selective and efficient drug candidates. The application of molecular docking in drug discovery process include the following:

- i. Hit identifications: Docking in combination with scoring function can be used to evaluate large databases for finding out potent drug candidate, which can target the molecule of interest.
- ii. Side effect prediction: Docking-based tools have predicted the efficacy of potential therapeutic compounds and have also helped in predicting the range of unintended and undesired interactions between the specific compound and the human proteome. It also plays a prominent role in the initial prediction of drug's binding properties to nucleic acid. This information establishes the correlation between drug's molecular structure and its cytotoxicity.
- iii. Molecular docking can be used to establish mechanisms of action a potent drug candidate against a molecular target.

## **Venom Molecular Targets**

Numerous snake venom proteins have been identified as druggable or potentially druggable targets, including phospholipases A<sub>2</sub> (PLA<sub>2</sub>s), metalloproteinases (SVMP), serine proteases (SVSP), kallikrein-like serine proteases, 5' nucleotidase, ATPase, alkaline phosphomonoesterase and acetylcholine esterase, and many of these have been characterized crystallographically. These protein crystal structures serve as structural models for in-silico screening using molecular docking techniques. Often, there are different structures, usually with different co-crystallized ligands, that can provide slightly different and complementary binding sites for docking studies (Kitchen *et al.*, 2004).

## Phospholipase A<sub>2</sub>

PLA<sub>2</sub> are a group of esterolytic enzymes present in snake venoms that typically catalyze the breakdown of glycerophospholipids, the main component of biological membranes, into lysophospholipids and a fatty acid (Harrison *et al.*, 2009). PLA<sub>2</sub>s are found in the venoms of Viperidae, Elapidae, and Colubridae snakes. The lengths of PLA<sub>2</sub>s vary from 119–134 amino acids, and they share a common scaffold of four main helices with seven intrachain disulfide bonds (Dubovskii *et al.*, 2014). The snake venom PLA<sub>2</sub>s are split into two groups, group I PLA<sub>2</sub>s are found predominantly in elapid and some colubrid snakes, while group II are found only within Viperidae (Kini 2003). Group I are generally  $\beta$ -neurotoxins which act pre-synaptically, sometimes binding to voltage gated potassium channels (Kini 2003). After binding, neurotoxic PLA<sub>2</sub>s can sometimes hydrolyze nerve terminal phospholipids causing permanent neurotoxicity (Harrison *et al.*, 2009). This has the effect of causing paralysis, while group II PLA<sub>2</sub>s tend to act cytotoxically, predominantly as myotoxins, causing myonecrosis via the disruption of the plasma membrane (Dubovskii *et al.*, 2014). Indeed, both natural and synthetic PLA<sub>2</sub> inhibitors are able to attenuate the morbidity and mortality of snake bite envenomation (Boldrin *et al.*, 2017).

## Computational studies on phospholipase A<sub>2</sub> inhibitors

There are many reports on computational molecular modeling methods used for the development of PLA<sub>2</sub> inhibitors that contribute to the attenuation of snake venom toxicity (Manjo, 2018). These applications use the x-ray crystallographic, 3D structural information generated in the last few decades, and methods such as molecular dynamics (MD) simulations and docking. Structurally, snake venom PLA<sub>2</sub>s is divided into classes I and II, this is based on their amino acid sequence and disulfide bonding pattern (Qiu and Liu, 2014). However, they have a conserved structure which contains an N-terminal  $\alpha$ -helix (H1), a Ca<sup>2+</sup>-binding loop, two antiparallel  $\alpha$ -helices (H2 and H3), a two-stranded antiparallel sheet ( $\beta$ -wing), and a long C-terminal loop. In general, folding is stabilized by seven disulfide bonds (with different pattern in classes I and II). Nargotra *et al.* (2011) evaluated a library of natural products and synthetic molecules through docking studies on *D. russelii* PLA<sub>2</sub> to identify possible inhibitors. Their study led to in silico identification of several molecules as PLA<sub>2</sub> inhibitors, with most of them belonging to phenolic and substituted benzaldehydic compounds (Mnoj *et al.*, 2014). The same authors proposed the docking poses inside PLA<sub>2</sub> of *D. russelii* for synthetic phenolic compounds effective against

snake venom. They found that phenolic compounds having hydroxyl and methoxyl groups in their benzene ring showed maximum inhibitory potency. The majority of molecular modeling applications in literature for studying PLA<sub>2</sub>s are oriented to rational design of novel inhibitors for the treatment of different Viperidae snakebites. In another work, Zhang *et al.* (2013) docked structural elements of the persimmon tannin PT40 (a highly galloylated condensed tannin with an unusual flavonol terminal unit) inside Chinese cobra (*Naja atra*) PLA<sub>2</sub> binding site, to understand the inhibitory mechanism of this natural product. They found that the residues Trp18, Try27, Gly29, His47, and Tyr63 are involved in the interactions. In 2014, Pereañez *et al.* studied the mode of action of morelloflavone with PLA<sub>2</sub> of *Crotalus durissus*, using docking. Authors found that morelloflavone occupies part of the substrate binding cleft of *C. durissus* PLA<sub>2</sub>, forming hydrogen bonds with the residues Gly33, Asp49, Gly53, and Thr68 of the enzyme, and  $\pi$ - $\pi$  stacking with the residue Tyr52. The same authors used docking to investigate the interactions between *C. durissus* PLA<sub>2</sub> and bile acids, such as cholic acid and ursodeoxycholic acid. Authors found that bile acids interact with the binding active site of PLA<sub>2</sub> through different interactions, cholic acid showed hydrogen bonds with His48, whereas, ursodeoxycholic acid showed hydrogen bonds with Asp49 and Tyr28. Also, Chavan and Deobagkar (2014) applied molecular docking simulation techniques to propose the putative interactions of LT10 peptide (small synthetic peptide derived from N-terminal of the lethal toxin neutralizing factor) with *Naja naja* PLA<sub>2</sub>. Molecular docking was performed to analyze the stability of the complex obtained by docking method.

Villar and collaborators (2008) demonstrated that synthetic inhibitor derivatives from nitrostyrene that contain typical nitro groups at the ortho-, meta-, and para- positions on the aromatic ring were more efficient against the enzymatic, edematogenic, and myotoxic activities of PLA<sub>2</sub>s from *B. jararacussu* venom. In a related research, Da Silva and his colleagues (2009), performing molecular modeling studies between Asp49-PLA<sub>2</sub> from *C. adamantus* venom and synthetic derivatives polyhydroxy phenolic compounds, concluded that some conformations of these groups might positively influence enzymatic activity inhibition.

Finally, in 2017, Mohanapriya *et al.* investigated phospholipase A<sub>2</sub> inhibitory activity of some medicinal plants against *Naja naja* venom using auto Dock. They observed that the molecular docking analysis of plant compounds against PLA<sub>2</sub> molecule shows the effective binding site in

the C-terminal end (Ile104, Ala101),  $\alpha$ - helix 3 (Val 47, Phe 46) and  $\alpha$ - helix 4 (Asp122, Pro121). Therefore, they concluded that *Aerva lanata* could be an effective treatment in treating snake bite.

### **Metalloproteinases (SVMPs)**

Snake Venom Metalloproteinases (SVMPs) are zinc-dependent proteinases ranging from 20 to 110 kDa in size and are categorized into P-I, P-II, and P-III classes according to their structural domains (Gutiérrez *et al.*, 2016). Studies have found that the SVMP is most abundant components in snake venom particularly viper species. Previous research has shown that SVMP induces hemorrhage by directly affecting the capillary blood vessels by clearing key bonds of the basement membrane components in a highly selective fashion, and thus affecting the interaction between the basement membrane and the endothelium (Laustsen *et al.*, 2017). Previous research has revealed that metalloproteinase is a mediator for edema, local tissue damage, inflammation, and hemorrhage (Markland and Swenson, 2013)

### **Computational studies on SVMPs inhibitors**

In 2015, Sathishkumar *et al.* provides in depth analysis on model the SVMP protein and also the identification of potent lead compounds (BD17344, BD905, BD 904, BD 25279, BD16837, BD13364, BD5228, BD17338, BD7951, BD26458, BD20708, BD16010, BD4991, BD5001, and BD15993) against SVMP. They model 3D structure of the SVMP using different softwares and the best model was selected based on the stereochemical properties. The best lead compounds for SVMP were screened using different databases viz. Binding, Maybridge, Hitfinder, and TOSLab databases. They identified fifteen potent hits through the Glide score and Glide energy. It was conclusively shown that all the compounds are quite stable in the active site of SVMP and thus, the isolated compounds might show promising activity against SVMP when compare to screening compounds.

Senkatachalaiah *et al.* 2014 reported the inhibitory effect of compound 5d, an apigenin based molecule against SVMPs both *in silico* and *in vivo*. The researchers found that the molecular docking of compound 5d and bothropasin demonstrated the direct interaction of hydroxyl group of compound with Glu146 present in hydrophobic pocket of active site and does not chelate  $Zn^{2+}$ . Hence, it is concluded that compound 5d could be a potent agent in viper bite management.

In 2009, Pithayanukulet *et al.* studied the ethanolic extract from seed kernels of Thai mango (Anacardiaceae) and its major phenolic principle (pentagalloylglucopyranose) inhibitory effects on the caseinolytic and fibrinogenolytic activities of Malayan pit viper and Thai cobra venoms in *in vitro* tests. Molecular docking studies revealed that the binding orientations of the phenolic principles were in the binding pockets of snake venom metalloproteinases (SVMPs). The phenolic principles could form hydrogen bonds with the three histidine residues in the conserved zinc-binding motif and could chelate the  $Zn^{2+}$  atom of the SVMPs, which could potentially result in inhibition of the venom enzymatic activities and thereby inhibit tissue necrosis.

Lina *et al.*, 2018 investigated the interactions between triterpenes (Ursolic acid, Oleanolic acid, Madecassic acid,  $\beta$ -boswellic acid, Betulin and Betulinic acid) and snake venom metalloproteinase using molecular docking simulation. The simulations revealed the atomic interactions that underlie binding between the triterpenic acids, most notably the electrostatic interaction between carboxylate groups of the compounds. The **researcher's findings** suggested that the occlusion of the S10 sub-site is essential for inhibition of proteolytic activity of metalloproteinases. They **also** found out that pentacyclic triterpenes having a carboxylate group at their C-17 position (betulinic, madecassic, oleanolic and ursolic acids) inhibit metalloproteinase proteolytic activity in experiment and exhibit favorable binding free energies and occlusion of the S10 subsite in simulation.

Conclusively, Muthusamy *et al.*, 2015 analyzed the role of  $Zn^{2+}$  and  $Ca^{2+}$  ions in the protein metalloproteinase activity using a compound clerodane diterpenoid by molecular docking. They performed molecular dynamics simulations up to 50ns using Desmond to understand the role of ligand in the active site of SVMP protein by various combinations. It was discovered that in the absence of both  $Zn^{2+}$  and  $Ca^{2+}$  ions from the protein, molecular docking simulation were fluctuating, particularly absence of  $Zn^{2+}$  ion with the protein. Based on the results, they concluded that the  $Zn^{2+}$  and  $Ca^{2+}$  ions have a vital role in the active site of SVMP protein.

### **Serine proteinases (SVSPs)**



SVSPs are found in venoms of the snake families Viperidae, Elapidae, and Colubridae (Fernandez *et al.*, 2011). Venom Serine Proteinases (SVSPs) belong to the S1 family of serine proteinases and display molecular masses ranging from 26 to 67 kDa with two distinct structural domains (Serrano, 2016). It catalyzes the cleavage of covalent peptide bonds in proteins and play key roles in diverse biological processes ranging from digestion to the control and regulation of blood coagulation, the immune system and inflammation (Matsui *et al.*, 2000). The anticoagulant SVSPs activate protein C via a thrombomodulin-independent mechanism. The most studied SVSP enzyme is from *Agkistrodon contortrix* venom, commercially referred to as Protac, which specifically converts protein C to activated protein C by hydrolyzing the Arg169-Leu170 bond, functioning independently of plasmatic factors (Panfoli *et al.*, 2010)

In 2018, Subhamay and Iman studied molecular interaction between serine protease and Hesperetin (one of the major flavonoid glycoside naturally present in several citrus fruits such as lemons and oranges) to investigate the likelihood of inhibition. The researchers obtained the amino acid sequence of thrombin-like serine protease from sharpnosed pit viper snake venom from the online database system of National Centre for Biotechnology Information. Employing AutoDock as molecular docking analysis software, they discovered the ideal ligand binding pose of hesperetin which is in close proximity to catalytic residues of snake venom serine protease, i.e., serine, histidine, and aspartic acid. Therefore, the generated *in silico* results by the researchers suggest that the novel structure hesperetin - flavanone might act as a potent inhibitor of thrombin-like snake venom proteases, and unlocks the possibilities for designing drugs of the inhibitors of snake venom serine proteases.

In a similar research, Roney *et al.*, 2018 investigated the inhibitory activity of citrus bioflavonoid, hesperetin on two thrombin-like snake venom serine proteases isolated from *Crotalus simus*. They performed the computational molecular docking studies to assess the possibility of serine protease thrombin-like being inhibited by flavonoids. Considering seven possible binding sites, their results suggest that hesperetin could form a hydrogen bond with Arg60 and His57 with the hydroxyl group on the carbon-5 from its A ring and oxygen atom 1 in the ring C could form hydrogen bonds with the Gly193 and Lys192. In addition, the methoxy group of the B ring could form hydrogen bond with the Gly216 and Asp217. They concluded

that flavonone is an interesting inhibitor for snake venom serine proteases, and could open up a possibility for drug design of snake venom serine protease inhibitors.

## **Conclusion**

Snake venoms are amongst the most fascinating animal venoms regarding their complexity and evolution. The generation and visualization of venom enzyme-inhibitor binding data from molecular docking simulation provide opportunity to explain structural features of enzyme inhibition mechanism. In this regard, molecular docking could be used as a pre-screen to identify compounds that are more likely to have different activity against a potential venom molecular target, which could lead to effective treatment of snake bite envenomation.

Ethic: NA

Consent: NA

## **Reference**

Gutierrez, J.M., Calvete, J.J., and Habib, A.G., et al. (2017), Snakebite envenoming. *Nature Reviews Disease Primers*. 3: 17063.

Gutiérrez, J.M., Theakston, R.D.G., and Warrell, D.A. (2006). Confronting the neglected problem of snake bite envenoming: The need for a global partnership, *PLoS Med*. 3:150.

WHO (2010), Guidelines for the prevention and clinical management of snakebite in Africa, Brazzaville: World Health Organization

Theakston, RDG., Warrell, D.A. (2000). Crisis in snake antivenom supply for Africa. *The Lancet* 356 (9247): 2104.

Nargotra, A., Sharma, S., Alam, M.I., Ahmed, Z., Bhagat, A., Taneja, S.C., Qazi, G.N., and Koul, S. (2011) In Silico identification of Viper Phospholipase2 Inhibitors: Validation By In Vitro, In Vivo Studies, *J. Mol.* 17: 3063–3073

Mohanapriya, M., Nandhini, A. R., Praveen, K. P., Yoganandhini, G., and Gowri-Shankar, B. A. (2017). Anti-phospholipase activity of medicinal plants against *Naja naja* venom, *Int. Res. J. Pharm.* 8(10):189-195

Zhang, Y., Zhong, L., Zhou, B., Chen, J.Y., and Li, C.M. (2013). Interaction of characteristic structural elements of persimmon tannin with Chinese cobra pla2. *Toxicon* 74, 34–43.

Serrano, S.M. (2013). The long road of research on snake venom serine proteinases, *Toxicon* 62:19-26.

Villar, F. P. F., Lima, T. D., and Veber, C. L. (2008). “Synthesis and evaluation of nitrostyrene derivative compounds, new snake venom phospholipase A<sub>2</sub> inhibitors,” *Toxicon* 51(8): 1467–1478

Da Silva, S. L., Calgarotto, A. K., Maso, V. (2009). “Molecular modeling and inhibition of phospholipase A<sub>2</sub> by polyhydroxy phenolic compounds,” *European Journal of Medicinal Chemistry* 44(1): 312–321.

Matsui, T., Fujimura, Y. and Titani, K. (2000). Snake venom proteases affecting hemostasis and thrombosis, *Biochim Biophys Acta* 1477:146-56

Panfoli, I., Calzia, D., Ravera, S. and Morelli, A. (2010). Inhibition of hemorrhagic snake venom components: Old and new approaches. *Toxins (Basel)* 2:417-27

Kitchen, D.B., Decornez, H., Furr, J.R., and Bajorath, J. (2004). Docking and scoring in virtual screening for drug discovery: methods and applications, *Nat Rev Drug Discov.*3(11):935–949.

Friesner, R.A. and Banks, J.L., and Murphy, R.B. (2004). Glide: a new approach for rapid, accurate docking and scoring. 1. Method and assessment of docking accuracy, *J Med Chem.* 47(7):1739–1749.

Wodak, S.J. and Janin, J. (1978). Computer analysis of protein-protein interaction, *J Mol Biol.* 124(2):323–342.

Gore, M. and Desai, N.S. (1983). Computer-aided drug designing, *Methods Mol Biol.* 1168:313–321

Theakston, R.D.G. and Reid, H.A. (1983). Development of simple standard assay procedures for the characterization of snake venoms, *Bulletin of the World Health Organization* 61: 949 – 956.

Onika, G., Punam, G. and Sarbjot, S. (2010). An overview on molecular docking, *International Journal of Drug Development & Research* 2(2): 229-231.

Ballante, F. (2018). Protein-Ligand Docking in Drug Design: Performance Assessment and Binding-Pose Selection, *Methods in Molecular Biology* 1824. pp. 67–88.

Manoj, G. (2018). Tyagi "Development of Novel Phospholipase2 Inhibitors Using Molecular and Computational Techniques" *IOSR Journal of Pharmacy and Biological Sciences (IOSR-JPBS)* 13(2): 05-12

Qiu, F., Chen, Y., Liu, L. (2014). Lai. Discovery of Novel Secretory Phospholipase A2 Inhibitors Using Virtual Screen. *Chem.Biol. Drug. Des.* 84: 216-222

Manoj, G., Sumith, K. M. (2016). Lipoprotein Associated Phospholipase A2 Enzyme; Possible New Roles and Inhibition for Therapeutic Intervention. *Int J Res Med Sci.* 2(3):805-809

Ballante, F., and Marshall, G.R. (2016). An Automated Strategy for Binding-Pose Selection and Docking Assessment in Structure-Based Drug Design, *Journal of Chemical Information and Modeling* 56 (1): 54–72.

Avinash, R., Veerabhadra, A. and Rao, A. (2013). A review on molecular docking, Novel tool in drug design and analysis, *Journal of Harmonized Research in Pharmacy* 2(4): 215-218.

Subhamay, P. and Iman, E. (2018). Molecular docking studies of snake venom serine protease of sharp-nosed pit viper with hesperetin Article in *Asian Journal of Pharmaceutical and Clinical Research* 11(6): 457-461.

Chavan, S.G.; Deobagkar, D.D. (2014). In silico molecular interaction analysis of LTNF peptide-LT10 with snake venom enzymes, *Protein Pept. Lett.* 21: 646–656.

Rangel-Santos, A.C., Mota, I. (2000). Effect of heating on the toxic, immunogenic and immunosuppressive activities of *Crotalus durissus terrificus* venom. *Toxicon* 38: 1451–1457

Fernández, J., Alape-Girón, A., Angulo, Y., Sanz, L., Gutiérrez, J.M., Calvete, J.J., Lomonte, B. (2011). Venomic and antivenomic analyses of the Central American coral snake, *Micrurus nigrocinctus* (Elapidae). *J. Proteome Res.* 10: 1816–1827

- Gutiérrez, J.M., Calvete, J.J., Habib, A.G., Harrison, R.A., Williams, D.J., Warrell, D.A. (2017). Snakebite envenoming. *Nat. Rev. Dis. Primer* 3: 17063
- Gutiérrez, J.M., León, G., Lomonte, B., Angulo, Y. (2011). Antivenoms for snakebite envenomings. *Inflamm. Allergy Drug Targets* 10: 369–380
- Pereanez, J.A., Patino, A.C., Nunez, V., Osorio, E. (2014). The biflavonoid morelloflavone inhibits the enzymatic and biological activities of a snake venom phospholipase a2. *Chem. Biol. Interact.* 220: 94–101.
- Pithayanukul, P., Leanpolchareanchai, J., Saparpakorn, P. (2009). Molecular docking studies and anti-snake venom metalloproteinase activity of Thai mango seed kernel extract.
- Pawelek, P., Cheah, J., Coulombe, R., Macheroux, P., Ghisla, S. and Vrieling, A. (2000). The structure of l-amino acid oxidase reveals the substrate trajectory into an enantiomerically conserved active site. *EMBO Journal* 19: 4204–4215.
- Kini, R.M. (2003). Excitement ahead: Structure, function and mechanism of snake venom phospholipase A2 enzymes. *Toxicon* 42: 827–840
- Gutiérrez, J.M., Escalante, T., Rucavado, A., Herrera, C., Fox, J.W. (2016). A Comprehensive View of the Structural and Functional Alterations of Extracellular Matrix by Snake Venom Metalloproteinases (SVMPs): Novel Perspectives on the Pathophysiology of Envenoming. *Toxins* 2016, 8, 304
- Markland, F.S.; Swenson, S. Snake venom metalloproteinases. *Toxicon* 62: 3–1
- Boldrini-França, J., Cologna, C.T., Pucca, M.B., Bordon, K.D.C.F., Amorim, F.G., Anjolette, F.A.P., Cordeiro, F.A., Wiezel, G.A., Cerni, F.A., Pinheiro-Junior, E.L. (2017). Minor snake venom proteins: Structure, function and potential applications. *Biochim. Biophys. Acta BBA-Gen. Subj.* 1861: 824–83.8
- Lomonte, B., Calvete, J.J. (2017). Strategies in ‘snake venomics’ aiming at an integrative view of compositional, functional, and immunological characteristics of venoms. *J. Venom. Anim. Toxins Trop. Dis.* 23.
- Harrison, R.A., Hargreaves, A., Wagstaff, S.C., Faragher, B., Lalloo, D.G. (2009). Snake Envenoming: A Disease of Poverty. *PLoS Negl. Trop. Dis.* 3:56

Laustsen, A.H., Johansen, K.H., Engmark, M., Andersen, M.R. (2017). Recombinant snakebite antivenoms: A cost-competitive solution to a neglected tropical disease PLoS Negl. Trop. Dis. 11:5361

Venkatachalaiah, S.,Mahalingam, S. S.,Sebastian, A.,Mahadevappa, H.,Siddaiah, C., Nayaka,K., Kemparaju, B.,Kesturu, S. Girish, Kanchugarakoppal, S. R. (2019). Novel Apigenin Based Small Molecule that Targets Snake Venom Metalloproteases, Toxin 19:22

Hegde, K., Naseeb, K.M., Syed, A., Deepak, T.K. and Kalangotttil, A. (2014). Evaluation of antivenom activity of ethanolic extract of Buchanania lanzan bark against naja kaouthia snake venom, *Unique journal of pharmaceutical and biological sciences*, 02 (02): 39-42.

Opadijo, O. G. and Omotosho, A.B. (1996). Snake bite in Ilorin: A review of 155 cases, Nigeria MedicalPract. 32: 30-32.

Dubovskii, P.V., Utkin, Y.N. (2014). Cobra Cytotoxins: Structural Organization and Antibacterial Activity. *Acta Naturae* 6: 11–18.

Lina, M. P., Jaime, A. P., Ettayapuram,R. A. S., and Jeffrey,C.(2018). Interactions between Triterpenes and a P-I Type Snake Venom Metalloproteinase: Molecular Simulations and Experiments, TOXIN 8