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Characterization and Comparison of Alkali Extracted Starches from Selected Cereals and Tubers

ABSTRACT

Aims: Starch is used in the food industry to impart functional properties and to modify food texture and consistency. In this study starches isolated from six sources using an alkali extraction method were evaluated for physicochemical properties.

Study design: Complete randomized design.

Place and Duration of Study: Department of Food Science and Technology, Faculty of Livestock, Fisheries, & Nutrition, Wayamba University of Sri Lanka, Makandura, Gonawila (NWP), Sri Lanka between April 2017 and August 2017.

Methodology: Starches were isolated from white rice, foxtail millet, proso millet, cassava, sweet potato and all-purpose wheat flour. Proximate composition, mineral content, physical properties and microscopic characteristics were determined and they were compared.

Results: The level of starch extracted was within the range of 27.5-64.1% on a dry basis. Extracted starch contains about 99% carbohydrate in DW and less than 1% non-carbohydrate fraction (protein, fat, fiber and ash and minerals). The amylose content followed the order: proso millet > wheat > foxtail millet > rice > sweet potato > cassava. There was considerable variation in swelling factor, solubility, gelatinization temperature among all starches. Both swelling power and solubility had a positive relationship with temperature and the swelling power (at 90°C) followed the order: corn > cassava > foxtail millet > wheat > proso millet > rice > sweet potato. The gelatinization temperature of starches ranged from 62°C to 76°C.

Conclusion: The study would be helpful to better understand the chemical, physical and microscopic characteristics of these starches and the application of novel starches obtained from non-conventional sources which are foxtail millet, proso millet and sweet potato as a thickening agent and a substitute to other common starches in food.

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Keywords: Foxtail millet; Microscopic characteristics; Physicochemical properties; Proso millet; Starch isolation

1. INTRODUCTION

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Starch is the second-largest biomass next to cellulose produced on earth. It is the major form of carbohydrate reserve in higher plants. It exists in a granular form and the starch granule is semi-crystalline and insoluble in water. Thus, a large amount of energy can be stored in a relatively small volume [1]. Today, the main sources of starch extraction are

27 seeds, roots and tubers, primarily from maize, wheat, cassava, rice and potato [2]. Starch
28 can easily be extracted with high purity, resulting in a white, tasteless and odorless powder.
29 These good organoleptic properties make it an interesting resource for manifold applications,
30 not only in human food and animal feed but also as feedstock for non-food industrial
31 applications such as pulp and paper, adhesives and bioethanol [3]. Starch is a valuable
32 ingredient in the food industry, it serves not only as a nutrient source for food but also as a
33 thickener, a binding agent, a texturizer, a filler and a film-forming agent in the food industry
34 [4, 5, 6].

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36 Starches from different plant sources vary in their chemical and physical characteristics as
37 well as their gelatinization properties [7]. A selection of starch varieties for different food
38 products depends on starch functional properties such as gelatinization properties, solubility,
39 viscosity, gel stability and retrogradation rate. According to Li [1], these functional properties
40 are determined by the chemical structures of starch. For industrial purposes the main source
41 of starch is maize. However, there are other potential sources of starches such as wheat,
42 cassava, and rice and they possess the potential characteristics required for the industrial
43 uses [8, 9]. Being a tropical country, Sri Lanka has underutilized cereals and yams with a
44 higher content of starches and may be possessing good physico-chemical properties for
45 industrial applications. Further, extraction of starches from these underutilized cereals may
46 improve the value-adding options for underutilized crops and enhance our understanding
47 and knowledge about physicochemical and functional properties of new starchy materials as
48 well as their ability to replace conventional starch in the food industry [5]. Thus the
49 application of non-conventional starch sources of cereals, roots and tubers in the food
50 industry can be broadened which in turn may reduce the dependency on corn, wheat and
51 cassava as the main sources of starch. In recent years, substantial efforts have been made
52 to obtain starches from non-conventional sources and to study their functional, rheological
53 and physicochemical properties. Foxtail millet and proso millet are some of the underutilized
54 cereals grown in Sri Lanka. In the present study, the physicochemical properties of starches
55 isolated from proso millet (*Panicum miliaceum*) and foxtail millet (*Setaria italica*) were
56 studied and compared with other major starch sources such as wheat flour (*Triticum* spp.),
57 rice (*Oryza sativa*), cassava (*Manihot esculenta*) and sweet potato (*Ipomoea batatas*)

58 59 **2. MATERIAL AND METHODS**

60 61 **2.1. Materials**

62 Dehulled seeds of foxtail millet (*Setaria italica*; variety ISC 480), proso millet (*Panicum*
63 *miliaceum*; variety AC 254) and dehulled and polished rice (*Oryza sativa*; Bg 357) were
64 supplied by Field Crops Research and Development Institute (FCRDI), Mahailuppallama, Sri
65 Lanka. Commercial all-purpose wheat flour (Brand name- Prima), Cornflour (Brand name-
66 Motha) were purchased from Cargills Food City, Dankotuwa, Sri Lanka and the fresh roots of
67 cassava (*Manihot esculenta*; Kirikawadi) and sweet potato (*Ipomoea batatas*; Wariyapola-
68 red) were obtained from the local market at Makandura area, Sri Lanka. All the chemicals
69 used for the study were of analytical grade.

70 71 **2.2. Flour preparation**

72 Flour extraction from selected sources was conducted following an established procedure
73 mentioned in Alves [10]. The dehulled grains were cleaned by removing solid and other
74 contaminants. For tubers, peeling, washing and slicing (~5mm) were done. About 100g of
75 cereals/sliced tubers were dried at 40°C for about 30 hours in an oven (Model no:
76 MEMMERT NLE 500) until they reached a constant weight. Subsequently, the dried
77 grains/sliced tubers were milled into flour by using a laboratory scale grinder and sifted

78 through a 300µm sieve. The flours were then packed into a sealed air tide polyethylene
79 container and stored at -18 °C until used for further studies.

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81 **2.3. Isolation of starch**

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83 The starch isolation was performed according to the method described by Correia and
84 Beirão-da-Costa [11], with slight modifications. Briefly, the flour (120g) was soaked in two
85 volumes of 0.25% NaOH at 2-5°C for 24 h. The suspension was homogenized and screened
86 through a muslin cloth and then 180µm sieve. The precipitate was screened successively in
87 63µm sieve. The mixture was centrifuged in a laboratory centrifuge (Model no: D-78532
88 Tuttlingen, Germany) at 800× g (4520 rpm) for 15 min. The mucilaginous layer was scraped
89 away and the precipitate was then suspended in water. This last step was repeated twice.
90 Isolated starch was dried for 48 hours at 45°C in the electrical drying oven. Then isolated
91 starch was ground into a fine powder using a laboratory-scale grinder and sifted through a
92 300µm sieve. The isolated starches were weighed and determined the yield of starch on the
93 dry weight basis. The starch was then packed into a sealed air tide polyethylene container
94 and stored in a laboratory freezer.

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96 **2.4. Microscopic characteristics of starches**

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98 The granular shape of starches was examined following the method as described by Snyder
99 [12]. A small drop of water was placed on one side of a standard microscope slide. About 5
100 mg of starch sample was transferred onto the water using a dissecting needle. The starch
101 was mixed thoroughly to disperse starch. A cover slip was placed over the suspension taking
102 care to avoid entrapment of air bubbles. Excess water was wicked off with a small piece of
103 tissue paper held at the edge of the cover slip to obtain a thin film. The granular shape was
104 examined under a polarized light microscope at ×10×40 magnification. The micrographs
105 were used to compare the morphology of the starch granules.

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107 **2.5. Determination of proximate composition**

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109 Moisture, protein, fat and ash contents in starches were determined by the method using the
110 methods described in AOAC [13]. Crude fiber contents were determined by Weedy method
111 using Fibertec™ M6 Fibre Analysis System. Total carbohydrate content was determined by
112 subtracting the sum of the values of crude protein, crude fat and ash content (% dry weight
113 basis) of the sample from 100 [14].

114

115 **2.6. Amylose content determination**

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117 The total amylose content of starches was determined using the spectrophotometric method
118 described by Hoover and Ratnayake [14].

119

120 **2.7. Amylopectin content determination**

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122 Amylose and amylopectin contents were expressed relative to the total starch content.

123

$$\text{Amylopectin content (\%)} = \% \text{ Total starch content} - \% \text{ amylose content}$$

124 **2.8. Determination of mineral content**

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126 The analysis of phosphorous was done using the method described by Varvel and Peterson,
127 [15] and the absorbance was measured using a UV-Visible spectrophotometer (Model no:
128 SP- 3000 Plus). Calcium, Magnesium, Zinc, Copper, Iron, Manganese and Aluminum
129 concentrations in starch samples were analyzed by using iCE 3000 series Thermo Scientific
130 Atomic Absorption Spectrometer.

131

132 **2.9. pH of starch**

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134 The method reported by Benesi et al. (2004) as cited in Nand [16] was used for pH
135 determination. Approximately five grams of starch sample was added to 20 ml of distilled
136 water in a beaker. The contents were stirred for 5 min. Starch was allowed to settle and the
137 pH of the water phase was measured using a calibrated pH meter.

138

139 **2.10. Particle Size Analysis**

140

141 The geometric mean diameter and particle size distribution of the flour samples were
142 determined by the sieving method as described in Patva [17]. The particle size of isolated
143 starches was determined by sieving method. Each empty sieve was accurately weighed.
144 The sieves were set as such, sieve with the pour size that largest diameter was on the top
145 and placed on sieve shaker. Accurately 100 g of starch was weighed and put into the top
146 sieve. The lid was placed on top and tightens clamps. Then the shaker was started to sieve
147 and shaken for 10 minutes and weighed at 5-minute intervals thereafter. Sieving was
148 completed when the weight on the smallest sieve containing starch changes less than 0.2%
149 of total sample weight for 5 minutes. Then the starch left on each sieve and amount of starch
150 sieved from final sieve were accurately weighed. Calculate the percentage of the weight of
151 each sieve. Then the geometric mean diameter of analyzed starches was determined using
152 the following equation.

$$\text{Geometric mean diameter of the particles by mass(mm)} = \log^{-1} \frac{[\sum (W_i \log d_i)]}{\sum W_i}$$

153 Where, W_i is the mass on the i^{th} sieve (g), n is the number of sieves, and
154 d_i is the nominal sieve aperture size of the i^{th} sieve (mm)

155

156 **2.11. Gelatinization Temperature**

157

158 The gelatinization temperature of starch was determined by the method described by Linus
159 [18]. A 20 ml of 0.29% W/V suspension of the sample in water in a 25 mL beaker was
160 warmed in a water bath at 40°C. The solution was thoroughly mixed and prepares a smear
161 from it. The smear was observed under the mid-power of the microscope. The temperature
162 was then gradually raised while mixing continuously and measuring temperature. Then after
163 every ~2°C a sample was withdrawn and observed under a light microscope until the starch
164 is fully gelatinized. The temperature when starch granule begins to lose their structure was
165 recorded as gelatinization temperature.

166

167 **2.12. Swelling power and solubility**

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169 Swelling power (SP) and solubility index (SOL) were determined using a modification of the
170 method of Leach et al. (1959) as cited in [19].

171

172 **2.13. Statistical analysis**

173

174 Significant differences between the results were calculated by analysis of the Least
175 Significant Difference (LSD) with the help of SAS software. Differences at $P < 0.05$ were

176 considered to be significant. Results were expressed as mean \pm SD (Standard Deviation).
177 Values were the average of triplicate experiments.

178

179 3. RESULTS AND DISCUSSION

180

181 Table 1 shows the percentage yield of flour and starches obtained from studied starch
182 sources. Flour extraction from cereals and tubers lies in the range 38.88–83.40% at 12%
183 moisture level. Compared to tubers, cereals have given the higher amount of flour due to
184 higher moisture content in tubers than cereals. Sweet potato yielded a relatively higher
185 amount of flour than cassava. The yield of flour of all starches was significantly different
186 ($P=0.05$) to each other. In this study commercially available corn flour and wheat flour were
187 used. The starch yield depends on the starch content of the plant source, variety of the plant,
188 starch granular size, presence of soluble and resistant starch and extraction method. The
189 yield of starch of all sources ranged from 27.49 to 64.10 % on a dry basis and visually the
190 resultant dried starches had an attractive appearance. Higher starch amount yielded from
191 corn flour (70.57 %) and it was significantly higher ($P=0.05$) than others. Proso millet, foxtail
192 millet and sweet potato yielded starches from their flours about 52.82%, 27.49% and 40.77%
193 respectively. However, the yield of both cassava and proso millet were not were not statically
194 significant different ($P=0.05$). According to Mistry [20], alkali concentration significantly
195 affected the starch yield and the lower starch yield at lower alkali concentration can be
196 observed due to lower solubility and dispersibility of glutelin proteins present in the flour type.
197 Furthermore, high levels of hydration and low density of the damaged starch granules
198 increased the viscosity of the slurry of flour and lead to difficulties in the screening
199 operations and therefore lower level of starch yield can be observed [20]. However, in the
200 present study, the aim was to evaluate a method of starch extraction which is relatively
201 simple and might readily be adapted to larger scale preparation conditions.

202

203 **Table 1: Percentage of extracted flour and starch extracted from sources**

Selected starch sources	% flour extracted	Starch Extractability (%)
Wheat	Nd	54.43 \pm 1.88 ^c
Rice	83.40 \pm 1.10	64.10 \pm 2.04 ^b
Proso millet	74.60 \pm 1.32	52.82 \pm 2.35 ^d
Foxtail millet	78.42 \pm 1.24	27.49 \pm 2.33 ^f
Cassava	38.88 \pm 1.41	54.47 \pm 0.69 ^{c,d}
Sweet potato	46.52 \pm 0.92	40.77 \pm 5.43 ^e
Corn	Nd	70.57 \pm 1.66 ^a

204 *Results are the mean of triplicate analyses and are expressed as mean \pm SD.*

205 *The same letter (a,b) indicate they are not significantly different. Values followed by the same letter in*
206 *each column are not significantly different ($p < 0.05$) by LSD test.*

207

208 3.1. Determination of microscopic characteristics of starches

209

210 The optical micrographs of the starches are given in Figures 1 (A, B, C, D, E, H). All of the
211 starch granules seem to consist of both simple and compound granules and packed in a
212 slightly different matrix. The native starches of studied sources consisted of a mixed
213 population of large, medium and small granules. Wheat starch granule (A) shapes include
214 symmetrical spheres and asymmetrical spheres. The rice starch granules (B) were

215 ellipsoidal to irregular or cubical and similar observations were also reported in [21]. Proso
216 millet starch granules (C) were spherical as well as polygonal in shape. The foxtail millet
217 starch granules (D) had irregular shapes, which varied from oval, round to polygonal in
218 shape and granules are larger than proso millet starch granules. The cassava starch
219 granules (E) appeared as spherical and similar observations were seen in [16]. The shapes
220 of the sweet potato starch granules (F) varied from polygonal to round/bell shapes and this is
221 in agreement with previous reports on sweet potato starch granules as mention in [22].
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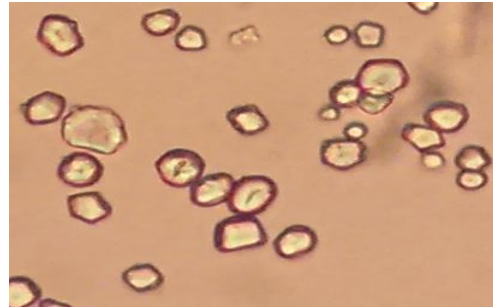
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Wheat Starch (A)



Rice starch (B)

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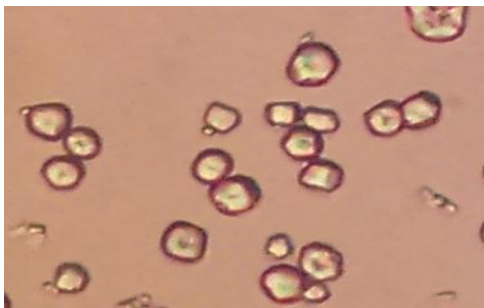
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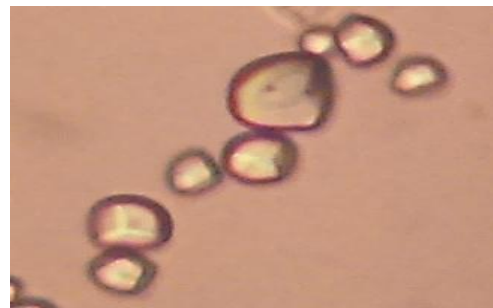
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Proso millet starch (C)



Foxtail millet starch (D)

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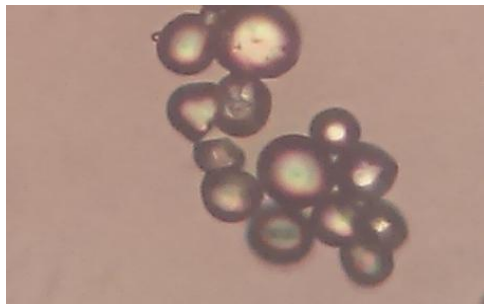
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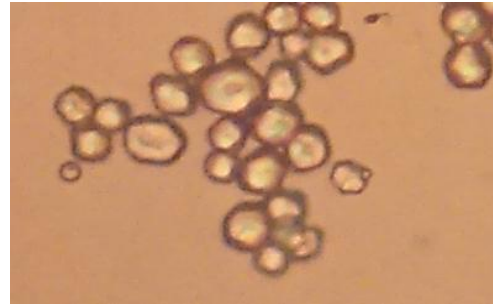
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Cassava starch (E)



Sweet potato starch (F)

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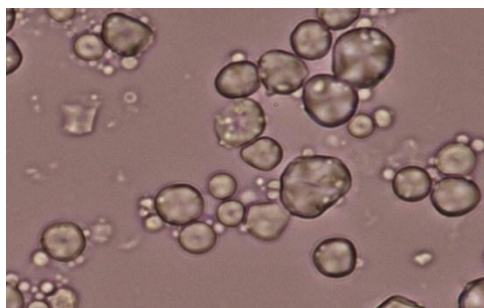
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Corn starch (G)

Figure 2: Optical micrographs of starch granules from selected sources at x400 magnification.

249 **3.2. Chemical characteristics of starches**

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251 The proximate composition of the different starches is shown in Table 2. The moisture
 252 content of studied starches was varied among sources and this could be attributed to the
 253 variation in the extent of drying of the starches. Moisture content falls within the moisture
 254 level (<20%) recommended for commercial starches as mentioned in Soni [23]. It was
 255 observed that the moisture content of starches is below 13%, which is recommended for
 256 safe storage in most starch producing countries [24]. Table 2 shows tuber starches
 257 contained a higher percentage of moisture compared to cereal starches. The protein
 258 contents in starch were very low and most of the proteins in the sources have removed
 259 during the extraction process. The protein content of the starches was ranged from 0.43 to
 260 0.71% on a dry basis. Lower protein contents were also reported in starches extracted from
 261 root tubers of purple, yellow and white sweet potatoes [25]. The highest value was observed
 262 from rice whereas sweet potato had the lowest. There was no statistical difference ($P=0.05$) in
 263 the protein content of both cassava and corn starches. The protein content of starches
 264 followed the order: wheat > foxtail millet > rice > proso millet > cassava > sweet potato.
 265 Studied tubers showed significantly lower protein level compared with that of cereals. Low
 266 protein content may be an indication of the absence of endosperm protein which could affect
 267 the purity and crystallinity of the starch and as a result could adversely affect the
 268 physicochemical properties of the starches. The fat and fiber content of the starches was
 269 very low and it is in the range of 0.10-0.14%.

270

271 **Table 2: Proximate composition of starches (%)**

Components	Moisture	Crude protein	Crude Fat	Crude fiber	Ash	Carbohydrate
Wheat	12.01±0.4 ^b 10.02±0.1 ^d	0.71±0.01 ^a	0.14±0.01 ^a	0.12±0.00 ^a	0.30±0.01 ^a	87.02
Rice	11.92±0.1 ^c	0.62±0.00 ^b	0.13±0.00 ^b	0.12±0.00 ^b	0.25±0.00 ^b	89.12
Proso millet	9.94±0.6 ^c	0.59±0.00 ^c	0.12±0.01 ^c	0.13±0.00 ^c	0.34±0.00 ^c	87.24
Foxtail millet	13.71±0.4 ^a	0.60±0.00 ^d	0.12±0.00 ^d	0.14±0.00 ^d	0.36±0.00 ^d	89.18
Cassava	12.34±0.5 ^b	0.52±0.01 ^e	0.10±0.00 ^e	0.14±0.00 ^e	0.47±0.00 ^e	85.51
Sweet potato	10.03±0.3 ^d	0.43±0.00 ^f	0.11±0.00 ^f	0.16±0.00 ^f	0.43±0.00 ^f	86.95
Corn		0.52±0.01 ^e	0.14±0.00 ^g	0.10±0.01 ^g	0.23±0.00 ^g	89.19

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Average values of three measurements (For n=3 ± SD). All data reported on a dry basis. Values followed by the same letter in each column are not significantly different (p <0.05) by LSD test.

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The total ash content of the starches ranged from 0.25 to 0.47% and the highest value was observed in cassava starch and the lowest from corn. Significantly lower cereal starches contain lower ash content. Higher mineral content was observed in cereal starches than tuber starches studied. Ash is the inorganic residue remaining after the water and organic matter has been removed by heating in the presence of oxidizing agents, which provides a measure of the total amount of minerals within a food. It has been suggested that higher ash contents of starch are a result of the presence of material commonly referred to as "fine fiber" [14]. The term fine fiber designates a highly hydrated material originating in the cell wall structures normally surrounding the starch granules. Higher ash content was indicated the lower yield of starch due to the higher fine fiber content. Generally, starch ashes are mainly composed of phosphorous, sodium, potassium, magnesium and calcium [26, 7]. The ash content is an estimate of the total mineral content of the starch. Even though, the values

287 for all the starches were low. The low ash content is an indication of the good quality of the
 288 starches because high mineral content is sometimes used to retard the growth of certain
 289 microorganisms [14]. The carbohydrate content of each starch is determined by difference on
 290 a dry basis and results of the analysis are given in Table 2. The alkaline steeping method is
 291 considered as one of the best methods of extraction high purity of starch [27]. Another
 292 popular method is the enzymatic method. All studied starches contained carbohydrate within
 293 the range of 85-89% and the highest values were observed in rice, foxtail millet and corn
 294 whereas the lowest carbohydrate content in cassava starch.
 295 The mineral compositions of the studied starches are presented in Table 3. The phosphorus
 296 content of all starches was ranged from 9.41 to 55.13 mg/100g. The highest value was
 297 obtained from rice and the lowest from wheat. Phosphorus content of rice, foxtail millet and
 298 proso millet was higher than root starches. According to [26], phosphorus in cereal exist as
 299 phosphate groups which are bounded to the amylopectin molecules and confer a
 300 polyelectrolyte nature to the chains. This ionic nature allows starch dispersions to develop
 301 high viscosity [26]. In the same way, Bergthaller [28] have proposed that the thickening
 302 capacity of starch is associated with the high content of phosphate ester groups. Root
 303 starches contain very low amounts of phosphorus and mostly in the phosphate monoester
 304 forms [29]. The higher phosphate content of starch is also the responsible parameter of high
 305 viscosity developed by these starch dispersions. Significantly higher calcium content was
 306 observed in starches of wheat, rice and foxtail millet than other studied starches.

307
308

Table 3: Mineral content of starches

Starch source	Mineral Content (mg per 100g)						
	P	Cu	Al	Zn	Ca	Fe	Mn
Wheat	9.41	1.06	9.95	0.28	41.82	6.05	0.28
Rice	55.13	1.25	23.27	1.41	32.24	6.45	0.56
Foxtail millet	28.22	1.51	20.45	0.16	7.49	6.90	0.24
Proso millet	33.84	1.59	20.50	1.39	21.55	7.01	0.28
Cassava	10.12	1.54	21.82	1.57	6.80	6.40	0.32
Sweet potato	12.50	1.50	23.62	0.25	6.34	6.51	0.42

309 *Average values of three measurements (For n=3 ± SD). All data reported on a dry basis.*

310

311 3.3. Amylose and amylopectin content of isolated starch

312 Starch consists of two major forms of polysaccharides such as amylose and amylopectin.
 313 Amylose is essentially a linear α -1,4 glucan with some molecules having α (1,6). Amylopectin
 314 is a branched molecule found in starch and the linear chain, the D-glucopyranose units are
 315 connected by α -1,4 linkages with 5-6% of its bonds being α -1,6 branch linkages. Amylose
 316 and amylopectin content in starches plays a significant role in influencing the functional
 317 properties of starches. Amylose and amylopectin content varied significantly ($P=0.05$) among
 318 the starches studied with values ranging from 17.08 to 27.83% and 72.16 to 82.91%,
 319 respectively (Table 4). According to Oduro [30], the differences in amylose content of these
 320 studied starches may be due to genotypic differences, environmental factors and starch
 321 processing methods. Interestingly, among the starches studied, proso millet showed the
 322 highest amylose content similar to commercial corn starches while the lowest was found in
 323 cassava starch. Cassava starch contains about 17% amylose and similar results were also
 324 found in Wang [31]. High amylose containing starches are characterized by their high

gelling strength which is indicating their usefulness in the production of pasta, sweets and bread-like products [32]. The amylose content affects gelatinization and retrogradation properties, swelling power and enzymatic susceptibility of starches [33]. Therefore, it is quite important to quantify the amylose content to determine the usability of the starches.

3.4. pH of isolated starch

The pH values of the starches are shown in Table 4. The pH values of studied starches were close to neutrality. However, the pH of commercial corn starch was within the acidic range (4.38). The higher pH values can increase the effect of the degree of ionization which has a significant effect in the hydration behavior of starches to permit the interaction between water molecules and amylopectin and amylose chains [34]. According to [16], the paste clarity of starches increased at very low pH and which decreased sharply towards high pH and in very acidic solutions, negatively charged phosphate groups are neutralized, and the ionization of hydroxyl groups is suppressed. Therefore, lysophospholipid may complex with amylose chains that contain only electropositive nitrogen and the presence of Coulombic repulsion between these positive nitrogen on adjacent amylose chains decrease the compactness of the amorphous region, thus increasing the transmission [35].

3.5. Gelatinization temperature

Gelatinization temperature is the temperature of the initial point that the starches loss their granule structure. The results in Table 4 showed that the corn starch has the highest gelatinization temperature than all other selected starches. Gelatinization temperatures of starches were ranged from 50°C to 76°C. Proso millet has the lowest gelatinization temperature and foxtail millet has the highest among the studied sources and though significantly lower compared with the gelatinization temperature of the commercial corn starch. The gelatinization property of starch is a determining factor in its functionality in food applications.

Table 2.1: Amylose, Amylopectine content, pH and gelatinization of starches

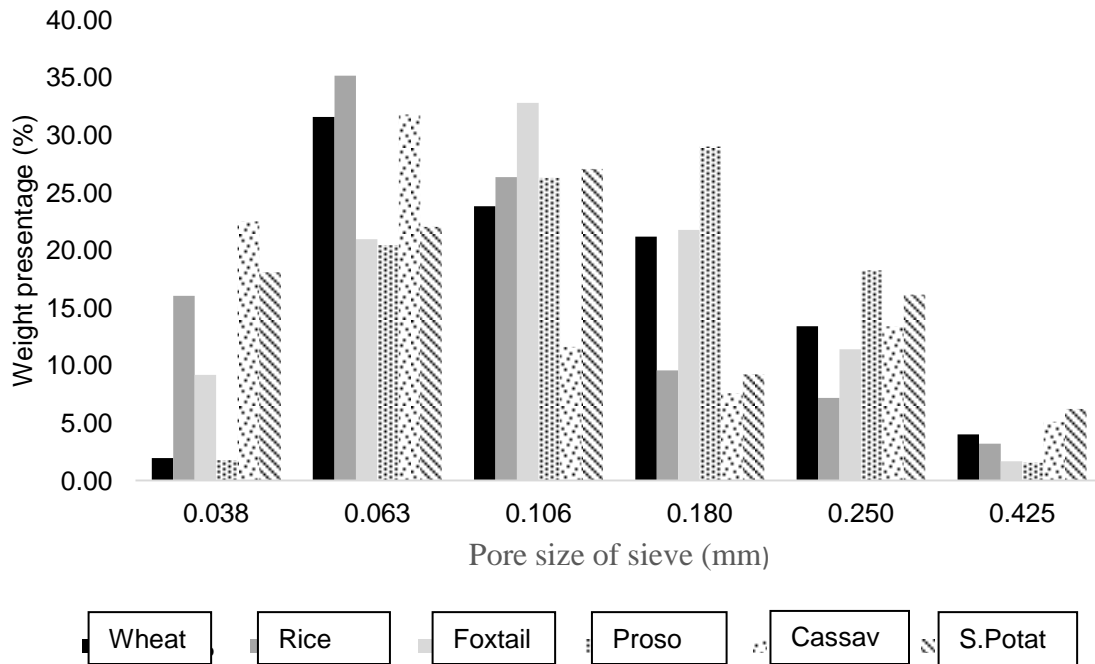
Starch	Amylose Content	Amylopectin content	pH	Geletinization Temp. (°C)	
				Start	Finish
Wheat	25.67±0.32 ^b	74.32±0.32 ^c	6.33±0.56	52	60
Rice	22.44±0.35 ^c	77.56±0.35 ^c	7.58±0.63	54	74
Proso millet	27.56±0.56^a	72.44±0.56 ^d	7.86±0.82	50	62
Foxtail	23.85±0.84 ^c	76.14±0.84 ^c	7.21±0.69	62	72
cassava	17.08±0.13 ^d	82.91±0.13 ^a	6.81±0.92	59	70
sweet potato	18.58±0.25 ^d	81.41±0.25 ^a	7.33±0.52	60	72
Corn	27.83±0.40^a	72.16±0.40 ^d	4.38±0.71	64	76

Average values of three measurements (n=3 ± SD). All data reported on a dry basis. Values followed by the same letter in each column are not significantly different (p < 0.05) by LSD test. pH values measured at 29 °C

3.6. Particle Size Distribution Analysis

The particle size distribution of isolated starches is presented in Figure 2. According to the analysis most of the particle size of wheat, rice, foxtail millet, proso millet, cassava and

365 sweet potato were at 0.180-0.063 mm, 0.106-0.063mm, 0.180-0.063 mm, 0.180-0.106 mm,
 366 0.063-0.038mm and 0.106-0.038mm range respectively. The geometric mean diameters of
 367 isolated starches are shown in Figure 3. It presented the geometric mean diameter of wheat,
 368 rice, foxtail millet, proso millet, cassava and sweet potato are 107 μm , 77 μm , 99 μm , 121
 369 μm , 78 μm , 93 μm and 73 μm respectively. The particle size of starch is one of the most
 370 important characteristics, which may influence other physicochemical properties such as
 371 swelling power, paste clarity, and water-binding capacity, among others [36].
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Figure 2: Particle size distribution of isolated starches

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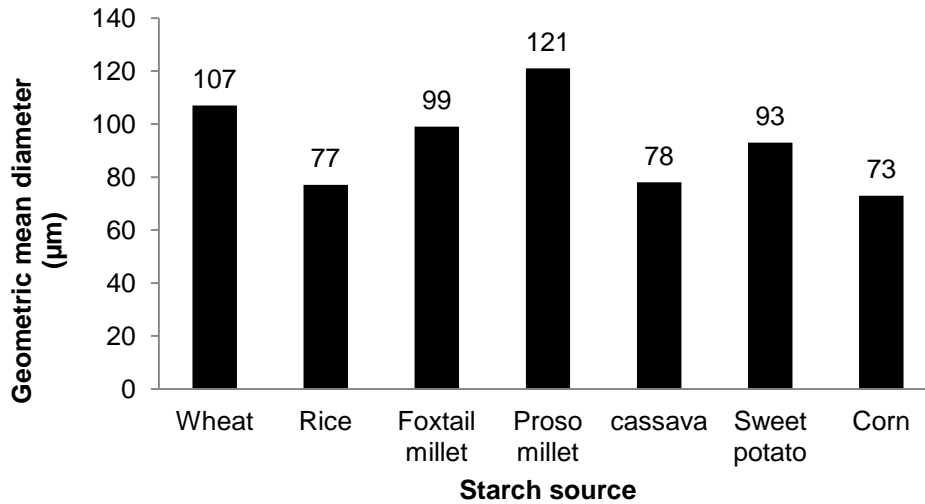


Figure 3: Geometric mean diameter (µm) of isolated starches

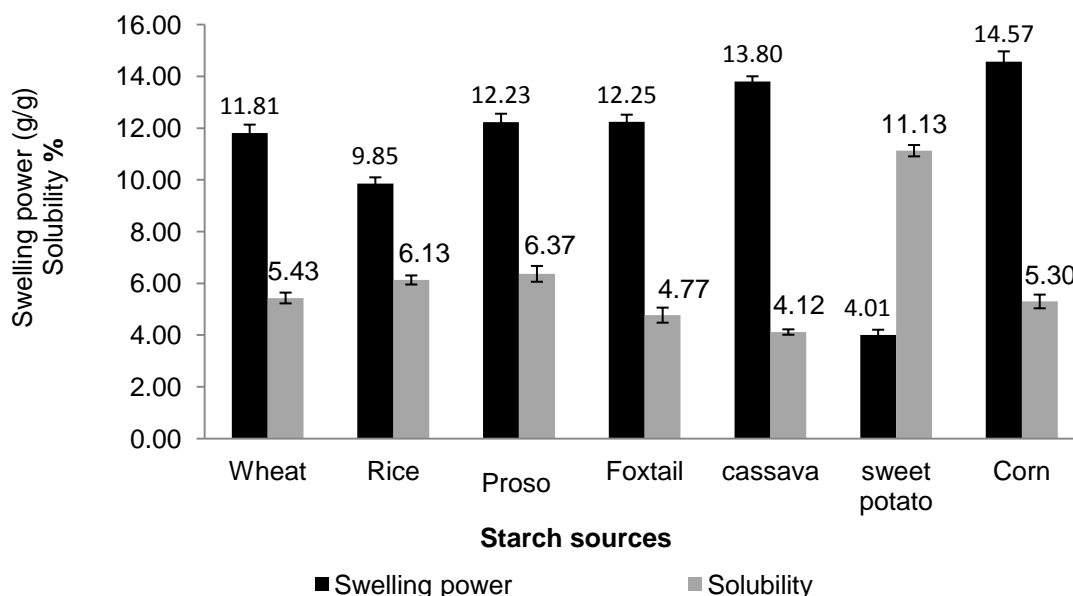
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3.7. Swelling power and solubility

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Swelling power and solubility values for the isolated starches are presented in Figure 4. The swelling power (at 90°C) followed the order: corn > cassava > foxtail millet > wheat > proso millet > rice > sweet potato. The swelling power of foxtail and proso millet and solubility of rice and proso millet are not significantly different ($P=0.05$). Sweet potato had the lowest swelling power and highest in cassava. All selected starches were lower than the swelling power of commercial corn. The swelling power of starches is often related to their protein and starch contents. Higher protein content in flour may cause the starch granules to be embedded within a stiff protein matrix, which subsequently limits the access of the starch to water and restricts the swelling power. In addition to protein content, a higher concentration of phosphorous may increase hydration and swelling power by weakening the extent of bonding within the crystalline domain [36]. Furthermore, the amylopectin is primarily responsible for granule swelling, thus higher amylose content would reduce the swelling factor of starch [37].



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Figure 2.1: Swelling power and solubility of starches at 90 °C.

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4. CONCLUSION

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The study showed that the starches extracted from selected sources exhibited differences in properties such as starch content, proximate composition, amylose, swelling power, solubility and gelatinization properties. Cereal starches contain a higher level of lipid and protein while tuber starches contain higher levels of moisture and fiber comparatively. The amylose content was high in proso and foxtail millets comparatively. Proso millet showed the highest gelatinization temperature whereas foxtail millet showed lowest comparatively. The study would be helpful to better understand the chemical, physical and microscopic characteristics of these starches and the application of novel starches obtain from non-conventional sources which are foxtail millet, proso millet and sweet potato as a thickening agent and a substitute to other common starches in food. Also, provide useful information for the further utilization of starches from food sources including two underutilized cereals for food and nonfood industries.

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COMPETING INTERESTS

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Authors have declared that no competing interests exist

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AUTHORS' CONTRIBUTIONS

425

426 This work was carried out in collaboration among all authors. Author HDKCW designed the study
427 and performed the statistical analysis. Author HDKCW wrote the first draft of the manuscript and
428 managed the analyses of the study. Author KDPPG managed the literature searches and wrote
429 the protocol. All authors read and approved the final manuscript
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