

**Subclinical mastitis survey on milk composition in dairy sheep in  
Kurdistan region of Iraq**

**ABSTRACT:**

Mastitis is an inflammatory condition of the mammary gland, characterized by the changes in the physical characteristics of the udder or milk. A cross sectional study was carried out June, 2017 to December, 2018 to estimate the effect of subclinical mastitis (SCM) on milk composition in dairy sheeps in Kafri city of Kurdistan region of Iraq. Milk samples were gathered from residences of 295 sheeps with subclinical mastitis (California mastitis test (CMT) positive and somatic cell counts (SCC) >600,000 cells/ml in individual quarter foremilk), as well as from 50 healthy controls. Contrasted to the levels watched in milk from healthy quarters, milk from quarters with subclinical mastitis exhibited raised high chloride (>0.12 in contrast with <0.12 g/dl), pH (5.65 in comparison to 5.57), sodium (86.87 vs 47.81 mg/dl), albumin (4.52 in contrast with 1.75 g/dl), immunoglobulins (24.66% in comparison to 5.73%) and lactate dehydrogenase (LDH) activity (1344.14 vs 449.84 IU/L). In compare, reduced values were discovered for potassium (147.47 in comparison to 161.34 mg/dl), inorganic phosphorous (19.42 in comparison to 26.48 mg/dl), calcium (86.35 vs 121.12 mg/dl),  $\beta$ -lactoglobulin (30.22% in comparison to 52.18%) and  $\alpha$ -lactalbumin (19.15% vs 24.52%). In this study, no changes were seen in blood serum LDH activity. Moreover, an increase in positive response to CMT was found to be accompanied by an almost proportionate increase in immunoglobulin values to 44.32% and reduce of  $\alpha$ -lactalbumin levels in milk serum ( $P<0.01$ ). These alterations in LDH activity, pH, mineral concentrations and protein fractions in milk of quarters display the presence of tissue injury provoked by SCM. Therefore, these parameters can be used in the diagnosis of mastitis. The current study revealed that changes of the foremilk chemical composition are connected to

28 the subclinical mastitis; and that mastitis progression of quarters (CMT scores) influenced protein  
29 fractions in milk.

30

31 **Keywords:** Composition , CMT, Kafri, Kurdistan, Iraq, Milk, Subclinical mastitis

32

### 33 **1. INTRODUCTION:**

34 Mastitis is the single most costly disease of dairy animals. Although large technological advances  
35 in the prevention and treatment of mastitis have been made in recent years, mastitis continues to  
36 cause major economic losses in dairy industry [1, 2]. This disease is usually connected with  
37 physical and chemical abnormalities of milk and udder through which it can be grouped into  
38 clinical or subclinical [3]. The gold standard diagnostic tool in both clinical and subclinical  
39 mastitis is the identification of the causative agent by culture [4,5]. Anyway, California mastitis  
40 test, somatic cells count (SCC), and changes in milk constituents are other important tools for  
41 detection subclinical mastitis in bovine [6] and these tests may be used for ovine mastitis  
42 detection (Please, insert here one reference for this affirmation). . In addition, Mastitis is an  
43 important problem causing very large economic losses in dairy industry throughout the world [7].  
44 Many of the intramammary infections (IMI) originate during the dry or non lactating period and  
45 result in clinical or subclinical mastitis during early lactation [8,9]. Subclinically infected udder  
46 quarters can improve clinical mastitis and the rate of new infections can be high [11]. Dairy sheep  
47 produce about 12.2 million metric tons (MT) of milk, accounting for about 1.5% of the world  
48 total amount of milk produced by livestock species, the largest amount of sheep milk is produced  
49 in India, followed by Iraq and Sudan [12]. The dairy sheep industry is quickly gaining in  
50 importance throughout the world in new years. Among the several problems hindering the  
51 livestock development in Iraq, sanitary problems constitute a serious threat to the successful  
52 production of livestock and its industry. Hence, any factor that adversely affects the quantity and  
53 quality of cattle and goat milk is of sheep financial interest. Milk quality is mainly influenced by  
54 bacterial contamination of the mammary gland, which causes clinical or subclinical mastitis [13].  
55 Mastitis is described as an inflammation of the mammary gland, affects lactating animals  
56 including sheep, goats, cattle, buffaloes and camels and is almost always caused by bacterial

57 infection. Mastitis in sheeps is mainly subclinical [14,15,16]. It is one of the serious problems of  
58 the dairy industry worldwide including Iraq. Subclinical mastitis is 10 to 35 times more common  
59 than the clinical form, is of long duration and difficult to discover [17, 18]. In Bangladesh, the  
60 prevalence of SCM is recorded from 20 to 44% at cow level based on California Mastitis Test  
61 (CMT) [19, 20]. The efficacy of antibiotic therapy for intramammary infections (IMIs) early in  
62 lactation is rare and Slight, with the ones carried out reporting mixed results. The response to  
63 therapy with intramammary (IMM) cephalosporin sodium on CMT positive quarters in lactating  
64 sheeps on cure rates and somatic cell count [21]. It was determined that by the 4-week post-  
65 calving evaluation, quarters treated with cephalosporin sodium had significantly increased cure rates,  
66 and SCC were significantly decreased.

67 Lessening the exposure of the udder to potential pathogens and/or increasing the immune  
68 response of dairy animals against infection remain some of the most effective mastitis control  
69 measures today [18]. There have been some research studies that proved the effectiveness of  
70 vaccination programs with a different combination of agents against mastitis in dairy sheep and  
71 cattle [22, 23]. Unluckily, most of the mastitis vaccines are only labeled for dairy sheep. Early  
72 identification of udder health problems is necessary for dairy farmers and veterinarians to ensure  
73 not only the animal well-being but also the milk quality and dairying productivity. Economic  
74 aspects interfere with the routine application of bacteriologic test of quarter milk samples. For  
75 this reason, alternative parameters are used to identify trends in the development of the udder  
76 health in a dairy herd, despite the fact that these parameters show inflammation. The aim of this  
77 study was to investigate the effects of relationship between a set of chemical parameters  
78 including pH, mineral concentrations, lactate dehydrogenase (LDH) activity and protein fractions  
79 and subclinical mastitis occurred naturally on dairy sheep.

80

## 81 **2. MATERIALS AND METHODS**

### 82 **2.1. Study area**

83 Field survey for this study, aiming to evaluate the effects of subclinical mastitis on milk  
84 composition, was done at three dairy herds located in some villages in Kafri city, Kurdistan  
85 region of Iraq. All the laboratory investigations were conducted at the Biology Laboratory of

86 College of Agriculture - Kifri, Garmian University, Kalar, As Sulaymaniyah, KRG of Iraq. The  
87 study was conducted for the three years period of from June, 2017 to December, 2018.

88 Sheeps were in the second to fifth lactation and were milked twice daily by hand milking. They  
89 were fed ad libitum by a total mixed diet that had been formulated to meet the nutritional  
90 requirements of a 350-kg sheep, yielding 10–18 kg of milk/d with about 1.2% protein and 1.1%  
91 milk fat. All sheeps were subjected to post dipping , those were dried off nearly two months  
92 before anticipated calving and all mammary glands of sheeps were infused with an antibiotic  
93 preparation for use in non-lactating sheeps following the last milking of lactation.

94

### 95 **2.2.Milk sampling and milk component analysis**

96 Milk samples were collected from quarters of 295 sheeps with subclinical mastitis (SCM), as  
97 well as from 50 healthy controls just before morning milking. Teats were scrubbed  
98 comprehensively and dried with a single use paper towel. The first three flows of milk from each  
99 teat were discarded. The teat end and aperture was disinfected with cotton swabs drenched in  
100 90% ethyl alcohol and nearly 8 ml foremilk sample were gathered from each quarter of sheep in a  
101 sterile tube held horizontally.

102

### 103 **2.3.California Mastitis Test (CMT)**

104 The experimental material was divided into four groups according to the California mastitis test  
105 (CMT) results—0 = negative or trace, 1 = weak positive, 2 = distinct positive and 3 = strong  
106 positive—obtained from the test performed directly in the herds, using the method described by  
107 Schrick *et al.* [11]. Blood samples were also gathered from jugular vein for the LDH assay.  
108 Samples were right away placed in crushed ice and submitted to the laboratory analysis within 3–  
109 5 hrs. To diagnosis of SCM, the total somatic cell count of milk was decided, using Breed's  
110 smudges with Newman's stain and leukocyte count more than 600,000 cells/ml of individual  
111 quarter milk was taken as a positive index of mastitis [6]. In all other cases, the samples were  
112 considered uninfected (healthy). All milk and blood samples were tested at midlactation and none  
113 of the ewes were sampled twice in the study.

114 Milk serum (whey) was readied at a two-step centrifugation procedure. At first, milk samples  
115 were centrifuged at 5000 rpm for 15 min to remove their creams and cells. Samples were then  
116 treated with 0.2 M hydrochloric acid at the controlled pH of 3.5 for casein precipitation. Treated

117 samples were recentrifuged and the supernatants (whey) were gathered. The pH of milk samples  
118 was determined electrometrically. Total calcium and phosphorous concentrations were  
119 determined using by colorimetric method, a hand-held spectrophotometer by commercial kits  
120 based on cresolphthalein complexion and phosphomolybdic acid complex formation, at  
121 wavelengths of 500 and 310 nm, respectively. Albumin was determined by bromocresol green  
122 method, using commercial kit at wavelength of 546 nm; chloride based on rapid spot test using K  
123 chromate and sodium and potassium by flame photometer; and silver nitrate (observation of  
124 yellow colour, >0.15 g/dl and brownish colour less than that amount) [9]. LDH activity was  
125 calculated by spectrophotometer, using commercial kit by the method of Siddiquee et al. [10] at  
126 wavelength of 320 nm. Protein fractionation of milk was Segregated according to molecular mass  
127 by cellulose acetate membrane electrophoresis (Sebia preference, France) at 90 V for 20 min and  
128 barbital buffer; pH = 6.8. After fractionation, membranes were stained with fixative dye solution  
129 (4.5% trichloroacetic acid, 0.4% Ponceau red, 97.5% double distilled water) at 10 min and then  
130 decolorized and purified. After drying, the relative levels of proteins were determined using  
131 densitometry at wavelength of 430 nm.

132

#### 133 **2.4. SCC determination**

134 Milk samples for SCC determination were gathered before vaccine administration (T0) and on  
135 days 30 and 32 of the experiment. SCC was determined using spreading 0.03 ml of gently blend  
136 milk from each sample over 2 cm<sup>2</sup> area of a glass slide and staining by Newman-Lampert stain.  
137 The stained slides were then tested by the same technician every time by light microscope  
138 according to previously published procedure [24]. SCC was expressed in log<sub>3</sub>.

139

#### 140 **2.5. Statistical analysis of the experimental data**

141 The software of SPSS [25] was used of data analysis. Student's t-test was carried out to find the  
142 differences between the results of mastitic, non-mastitic milk and serum. The changes in the  
143 content of protein fractions in milk with different positive CMT scores were appraised by one-  
144 way analysis of variance (ANOVA) followed by Duncan's multiple range test. The results were  
145 given as mean ± SEM. A repeated measures ANOVA test was used to estimation milk  
146 composition variables over different sampling points in vaccinated and non-vaccinated normal  
147 ewes. p<0.05 was measured statistically significant.

148

149 **3. RESULTS**

150 Present study was done in order to investigate the effects of relationship between a set of  
151 chemical parameters including pH, mineral concentrations, lactate dehydrogenase (LDH)  
152 activity, protein fractions and subclinical mastitis occurred naturally on dairy sheep. The results  
153 of Table 1 showed that the concentrations of potassium, phosphorous and calcium were  
154 significantly lower in the milk of inflamed (SCM) mammary glands than those of normal glands  
155 ( $P<0.01$ ).

156 The concentrations of albumin, chloride and sodium were significantly higher in the milk of  
157 inflamed mammary glands than those in normal ones ( $P<0.01$ ). The pH was considerably higher  
158 in the subclinical mastitic milk than in the normal ones ( $P<0.01$ )

159

160 **Emplacement of Table 1**

161

162 The LDH activities of milk and blood serum samples of normal animals and animals affected by  
163 subclinical mastitic were presented in Table 2. The mean LDH activity was considerably higher  
164 in milk from inflamed (SCM) quarters than in normal milk ( $P<0.01$ ). No significant difference  
165 was observed in LDH serum values.

166

167 **Emplacement of Table 2**

168

169 The contents of protein fractions were contingent upon the CMT progression. Statistically  
170 significant ( $P<0.01$ ) influence of high mastitis progression on the increase in milk  
171 immunoglobulin values to 35.20% was detected. Milks obtained from highly inflamed glands  
172 (milk samples with high score in CMT) had significantly ( $P<0.01$ ) lower albumin and pre-  
173 albumin and  $\alpha$ -lactalbumin, but the content of  $\beta$ -lactoglobulin in milk was comparable between  
174 quarters with different CMT scores (Table 3).

175

176

177 **Emplacement of Table 3**

178 The Table 4 shows that the concentrations of protein fractions were significantly different  
179 between normal and SCM milk ( $P < 0.01$ ). SCM caused increment in the immunoglobulin and  
180 albumin content in milk. While,  $\beta$ - lactoglobulin,  $\alpha$ -lactalbumin and pre-albumin content in SCM  
181 milk was reduced relationship normal milk.

182

#### 183 **Emplacement of Table 4**

184

#### 185 **4. DISCUSSION**

186 Mastitis is an important problem causing very large economic losses in dairy industry throughout  
187 the world [1, 2]. Many of the intramammary infections (IMI) are created during the dry or non  
188 lactating period and result in clinical or subclinical mastitis during early lactation. Swelling of the  
189 mammary gland leads to a diversity of compositional changes in milk either because of local  
190 results or because of serum components entering the milk and the movement of some normal  
191 milk components out of the alveolar lumen into the perivascular space [4]. Hypothetically, all  
192 changes in mammary discharge during swelling might be used to measure the effects of mastitis,  
193 but problems of instrumentation and standardisation have hampered farm application of most  
194 examinations. Albumin content of milk in subclinical mastitis was meaningfully increased  
195 compared to the healthy ones. The increase of albumin content in milk during mastitis has been  
196 reported in goats [26, 27], sheep [28] and goats [2]. Although, it be usual think that the main site  
197 of albumin synthesis is in the liver, and that the albumin enters the milk by leaking through the  
198 epithelial tight junction from the blood stream [18], the extrahepatic synthesis of albumin has  
199 been exhibited in mammary gland epithelial cells, albeit lesser than the liver [8]. The noticeable  
200 increases of albumin in mastitic animals propose that a great source of the increase in the content  
201 of albumin in milk under inflammatory situations is the inflamed gland itself. Our findings shows  
202 that tissue disturbances of the mammary gland in subclinical mastitis were accompanied by  
203 significant increase of LDH activity in the milk, but without obvious influence on enzyme levels  
204 in blood serum. Higher LDH activity in milk serum of inflamed udders has been previously  
205 reported in goats [10, 7] and cows [11]. The higher level of LDH in mastitic milks than in the  
206 blood serum reveals that blood serum was not the sole source of this enzyme during mastitis  
207 cases and that it is probably also liberated from disintegrated leukocytes and the parenchymal  
208 cells of the udder [27]. The pH of SCM milk was higher than that of normal milk, which is

209 agreement with the results of earlier reports [2]. The circuitous pH testing can be measured as a  
210 guide to detect the subclinical mastitis as this is economical, comfortably and rapid. It can be  
211 done in the field at the time of milk collection. Later determining pH, the positive samples can be  
212 checked to isolate the causative organism for further confirmation of SCM. Mastitis also  
213 noticeably changed the ionic environment. Chloride and sodium are increased while potassium,  
214 normally the predominant mineral in milk, is decreased. These increases in chloride and sodium  
215 and reduce in potassium levels have been verified by other authors as methods of monitoring  
216 udder health [8, 27]. Intramammary infection results in injury to the ductal and secretory  
217 epithelium, an opening of the “tight junctions” between secretory cells, and the increased  
218 permeability of the blood capillaries. Thus, chloride and sodium pour into the lumen of the  
219 alveolus and, in order to keep osmolarity, potassium levels reduce relatively. The levels of  
220 phosphorous and calcium is also influenced by mastitis. The reduction in phosphorous and  
221 calcium levels in the case of intramammary infections have been previously reported [16, 18].  
222 The current study showed that the types of proteins present in all of the milking fractions from  
223 quarters with subclinical mastitis undergo dramatic changes. Quarters with SCM revealed higher  
224 immunoglobulins and lower lactalbumin than did the corresponding milking fractions taken from  
225 healthy ones. The increased proportion of immunoglobulins connect to inflammatory responses  
226 of the udder compensated for the significantly lower proportion of lactalbumin. Actually, there is  
227 a near balance between this reduce and increase. Changes in protein fractions of milk acquired  
228 from mastitic sheep have been documented in previously studies [1, 22]. Immunoglobulins in  
229 mammary discharges are serum-derived or produced in the udder and pass into the milk through  
230 the mammary epithelium. The concentrations of immunoglobulins in normal milk are low and  
231 depend on the degree of vascular permeability of the udder tissues. When this penetrability  
232 barrier is broken during inflammation, immunoglobulin concentrations increase in discharges  
233 from infected glands. The immunoglobulin has several important functions. They are believed to  
234 prevent bacterial adherence to inhibit multiplication in epithelial membranes, agglutinate bacteria  
235 and neutralize toxins. Also, an important function of immunoglobulins is opsonization of  
236 microorganisms for phagocytosis. The increase in milk immunoglobulins may be effective in  
237 decreasing severity of mastitis [4]. Specific proteins are greatly synthesized in the mammary  
238 gland. This reduce in  $\alpha$ - lactalbumin connect to SCM could be due to the decreased synthetic  
239 activity of mammary gland. Some studies propose that  $\alpha$ -lactalbumin may leak out of the



240 alveolus between epithelial cells; this component has been calculated in urine or blood of sheeps  
241 with mastitis [28].  $\beta$ -lactoglobulin and  $\alpha$ -lactalbumin have physiological properties of whey  
242 proteins involving immunoenhancing effects. The possible role of  $\alpha$ -lactalbumin as an antitumour  
243 agent is being investigated [10].

244

## 245 **Conclusion**

246 The results of study showed that these alterations in LDH activity, pH, mineral concentrations  
247 and protein fractions in milk of quarters display the presence of tissue injury provoked by SCM.  
248 Therefore, these parameters can be used in the diagnosis of mastitis. The current study revealed  
249 that changes of the foremilk chemical composition are connected to the subclinical mastitis; and  
250 that mastitis progression of quarters (CMT scores) influenced protein fractions in milk.

251

## 252 **COMPETING INTERESTS**

253 Author has declared that no competing interests exist

254

## 255 **References**

- 256 1. Anderson, K.L.; Smith, A.R.; Shanks, R.D.; Whitmore, H.L.; Davis, L.E., Gustafsson, B.K.  
257 (1986). Endotoxin-induced bovine mastitis: immunoglobulins, phagocytosis, and effect of  
258 flunixin meglumine. *Am. J. Vet. Res.*, 47: 2405-2410.
- 259 2. Leitner, G., Chaffer, M., Shamay, A., Shapiro, F., Merin, U., Ezra, E., Saran, A., Silanikove,  
260 N. (2004a). Changes in milk composition as affected by subclinical mastitis in sheep. *J. Dairy*  
261 *Sci.*, 87: 46-52.
- 262 3. Mishra, S.K., Panda, B., Mohapatra, S.C., Ahuja, S.D. (1993). Genotype-protein interaction  
263 for egg production traits in Japanese quail. *India J. Poult. Sci.*, 29(1): 18-22.
- 264 4. Nazifi, S., Haghkhah, M., Asadi, Z., Ansari-Lari, M., Tabandeh, M.R., Esmailnezhad, Z.,  
265 Aghamiri, M. (2011). Evaluation of sialic acid and acute phase proteins (haptoglobin and  
266 serum amyloid A) in clinical and subclinical bovine mastitis. *Pakistan Veterinary Journal* 31:  
267 55-59.
- 268 5. Moroni, P., Vellere, F., Antonini, M., Pisoni, G., Ruffo, G., Carli, S. (2004). Antibiotic  
269 susceptibility of coagulase-negative staphylococci isolated from goat's milk. *Int. J.*  
270 *Antimicrob. Agents.*, 23:637-640.

- 271 6. Leslie, K.E., Dingwell, R.T. (2000). Mastitis control: where are we and where are we  
272 going? In: Andrews, AH (Ed.), *The health of dairy cattle*. (1st. Edn.), Malden, Blackwell  
273 Series. PP: 370-381.
- 274 7. Neelesh, S., Gyu, J.R., Yeong, H.H., Tae, Y.K, Hak Kyo Lee, H.K. Tai-Young, H., Dong,  
275 K.J., (2012). Bovine Mastitis: An Asian Perspective. *Asian Journal of Animal and Veterinary*  
276 *Advances* 7: 454-476.
- 277 8. Shamay, A., Mabjeesh, S.J., Silanikove, N. (2002). Casein-derived phosphopeptides disrupt  
278 tight junction integrity, and precipitously dry up milk secretion in goats. *Life Sci.* 70:2707–  
279 2719.
- 280 9. Schaar, J., Funke, H. (1986). Effect of subclinical mastitis on milk plasminogen and plasmin  
281 compared with that on sodium, antitrypsin and *N*-acetyl-D-glucosaminidase. *J. Dairy Res.*  
282 53:515–528.
- 283 10. Siddiquee, N.U., Tripura, T.K., Islam, M.T., Bhuiyan, S.A., Rahman, A.K.M.A., Bhuiyan,  
284 A.K.F.H. (2013). Prevalence of sub-clinical mastitis in high yielding crossbred cows using  
285 draminski mastitis detector. *Bangladesh Journal of Veterinary Medicine* 11 (1): 37-41.
- 286 11. Merkhan, K., Alkassa, J. (2012). study on milk composition of black and Meriz goats raised  
287 under farm condition. *Journal of University of Duhok.* 15(1): 58-63.
- 288 12. FAOSTAT. 2008. <http://faostat.fao.org/default.aspx>.
- 289 13. Fthenakis, G.C., El-Masannat, E. T., Booth, J. M., Jones, J. E. T. (1991). Somatic cell count  
290 of ewes' milk. *Br. Vet. J.* 147:575–581.
- 291 14. Auldish, M.J., Coats, S., Sutherland, J. B., Mayes, J. J., McDowell, H. G. (1996). Effects of  
292 somatic cell count and stage of lactation on raw milk composition and the yield and quality of  
293 cheddar cheese. *J. Dairy Res.* 63:269–280.
- 294 15. Batavani, RA.; Mortaz, E.; Falahian, K., Dawoodi, M.A. (2003). Study on frequency,  
295 etiology and some enzymatic activities of subclinical ovine mastitis in Urmia, Iran. *Small*  
296 *Ruminant Res.*, 50: 45-50.
- 297 16. Fthenakis, G.C. (1994). Prevalence and aetiology of subclinical mastitis in ewes of Southern  
298 Iraq. *Small Rumin. Res.* 13:293–300.
- 299 17. Kotula A.W., Thomson, J.E. Kinner, J.A. (1960). Weight increase during chilling of broilers  
300 as influenced by methods of opening the abdominal cavity during evisceration. *Poult. Sci.*,  
301 39: 26-27.

- 302 18. Fantuz, F., Plidori, F., Cheli, F., Baldi, A. (2001). Plasminogen activation system in goat  
303 milk and its relation with composition and coagulation properties. *J. Dairy Sci.* 84:1786–  
304 1790.
- 305 19. White, E.C., Hickley, L.S. (1999). Prevalence of mastitis pathogens in goat milk. *Small*  
306 *Rum.Res.*, 33:117-121.
- 307 20. Singh, P.K., Khatta, V.K., Thakur, R.S., Dey, S., Sangwan, M.L. (2003). Effects of phytase  
308 supplementation on the performance of broiler chickens fed maize and wheat based diets with  
309 different levels of non-phytate phosphorus. *Asian Australas. J. Anim. Sci.*, 16(1):1642-1649.
- 310 21. Pandey, N.K. Shyamsunder, G. (1990). Carcass characteristics, meat yield and  
311 physicochemical properties of meat from white leghorn cockerels. *Indian J. Poult. Sci.*,  
312 25(4):249-252.
- 313 22. Ernstrom, C.A., Wong, N. P. (1974). Milk clotting enzymes and cheese chemistry. Pages  
314 662—771 in *Fundamentals of Dairy Chemistry*. H. B. Webb, A. H. Johnson, and J. A. Alford,  
315 ed. Avi Publ. Co., Inc., Westport, CT.
- 316 23. El-Dengawy, R.A. and Nassar, A.M. 2001. Investigation on the nutritive value and  
317 microbiological quality of wild quail carcasses. *Nahrung/Food* 45. No. 1, pp. 50-54.
- 318 24. Leitner, G., Merin, U., Silanikove, N. (2004b). Changes in milk composition as affected by  
319 subclinical mastitis in goats. *J. Dairy Sci.*, 87: 1719-1726.
- 320 25. SPSS. (Statistical Procedure for Social Sciences), 1998. SPSS 11.5 for windows.
- 321 26. Ndegwa, E.N., Mulei, C. M., Munyua, S.J. (2000). The prevalence of subclinical mastitis in  
322 dairy goats in Kenya. *J. S. Afr. Vet. Assoc.*, 71(1):25-27.
- 323 27. Ndegwa, E.N., Mulei, C.M., Munyua, S.J.M. (2001). Risk factors associated with  
324 subclinical subacute mastitis in Kenyan dairy goats. *Israel J. Vet. Med.* 56:4.
- 325 28. Leitner, G., M. Chaffer, Y. Caraso, E. Ezra, D. Kababea, M. Winkler, and A. Saran. 2003a.  
326 Udder infection and milk somatic cell count, *NAGase* activity and milk composition-fat,  
327 protein and lactose-in Israeli Assaf and Awassi sheep. *Small Rumin. Res.* 49:157–164.

328

329

330

331

332

333 Table 1: Comparisons of pH, albumin and minerals in milk of normal and milk of mammary  
334 glands with subclinical mastitis

Parameters	SCM milk	Normal milk
pH	5.65 ± 0.08	5.57 ± 0.01*
Albumin (mg/dl)	4.52 ± 0.1	1.75 ± 0.02*
Chloride(mg/dl)	<0.12*	>0.12
Potassium(mg/dl)	147.47 ± 201	449.84 ± 1.1*
Sodium(mg/dl)	86.87 ± 4.1	47.81 ± 1.1*
Calcium(mg/dl)	86.35 ± 1.1	121.12 ± 0.6*
Phosphorous(mg/dl)	19.42 ± 0.2	26.48 ± 0.2*

335

336

337 Table 2: : Comparisons of LDH in milk and blood sera of normal and infect animals (SCM)

	SCM milk	Normal milk	SCM serum	Normal serum
LDH (IU/L)	1340± 110.1	280.1± 11.3*	601.1± 18.14	640.2 ± 25.1

338

339

340 Table 3: Comparisons of milk albumin, pre-albumin, immunoglobulin, β-lactoglobulin and α-  
341 lactalbumin according different scores in positive CMT

	+++	++	+
Albumin	5.1±1.2 <sup>c</sup>	15.2 ± 1.3 <sup>b</sup>	25.1.3 <sup>a</sup>
Pre-albumin	0.1 ± 0.12 <sup>a</sup>	0.05 ± 0.2 <sup>b</sup>	0.1 ± 0.04 <sup>a</sup>
Immunoglobulin	45.1 ± 1.1 <sup>c</sup>	17.12 ± 0.32 <sup>b</sup>	9.1 ± 0.21 <sup>a</sup>
β-Lactoglobulin	32.1 ± 1.2 <sup>a</sup>	33 ± 1.01 <sup>a</sup>	28.1 ± 1.01 <sup>a</sup>
α-Lactalbumin	10.3 ± 1.03 <sup>c</sup>	22.1 ± 1.1 <sup>b</sup>	26.1 ± 0.6 <sup>a</sup>

342

343

344 Table 4: Comparisons of albumin, pre-albumin, immunoglobulin,  $\beta$ -lactoglobulin and  $\alpha$ -

345

lactalbumin in milk of normal or mastitic mammary glands (SCM milk)

346

	SCM milk	Normal milk
Albumin	15.2 $\pm$ 1.1	5.3 $\pm$ 43 *
$\alpha$ -Lactalbumin	19.15 $\pm$ 0.54	24.52 $\pm$ 0.4*
$\beta$ -Lactoglobulin	30.22 $\pm$ 1.1	52.18 $\pm$ 0.5*
Pre-albumin	0.06 $\pm$ 0.3	0.15 $\pm$ 0.02*
Immunoglobulin	24.66 $\pm$ 0.52	5.73 $\pm$ 0.22*

347

348

349