

## Review Article

# Salicylic Acid Alleviates Postharvest Fruit Decay of Strawberry (*Fragaria x ananassa* Duch.-A mini review

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

### ABSTRACT

Strawberry (*Fragaria x ananassa* Duch.) fruits are highly perishable; and fruit quality decrease rapidly after harvesting, thereby it has a limited scope of long duration storage. Among several synthetic chemicals suggested for minimizing postharvest losses of fruits, Salicylic acid (SA) is a natural phenolic compound widely distributed in plants s and considered as a hormone because of its regulatory role in plants and has received a particular attention because of its role in the modulation of the plant response to biotic and abiotic stresses. Current scientific knowledge on the salicylic acid application in postharvest management of strawberry fruits suggests that SA has a potential role in minimizing fruit decay and maintaining the fruit quality. These predictors, however, need further work to validate reliability in postharvest management of strawberry fruits in a larger perspective.

Comment [PK1]: It would be better to rewrite this sentence into two separate sentences. (too lengthy)

Keywords: *Fragaria x ananassa* Duch., Fruit decay, Postharvest, Salicylic acid, Strawberry

### 1. INTRODUCTION

Modern cultivated strawberry (*Fragaria x ananassa* Duch.) is an octaploid ( $2n=8x=56$ ) hybrid species in the Family *Rosaceae* [1], that arose during the mid-1700s in France from two New World species *F. chiloensis* and *F. virginiana* [2]. Worldwide, strawberries are grown in a broad range of climates including temperate, grassland, Mediterranean, and subtropical regions [3], and globally stands most important berry fruit [4]. In India, temperate Jammu Kashmir, Himanchal Pradesh, and Uttarakhand have been known for strawberry cultivation, but in the recent decades, there has been an expansion in area and production of strawberry in plains of subtropical regions and North-Eastern states of India [5]; however, the runner production in plains are very poor [6]. Strawberries are the most preferred berry fruit due to its ~~their~~ pleasant aroma, colour, taste, flavor, and bioactive compounds [7,8]. Strawberry fruits are rich in nutritional content, phenolic compounds, anthocyanins, tannins, phenolic acids, and have potential health benefits [9].

The strawberry fruits are highly perishable; it has limitations in long-long-distance transport in the conventional system of fruit growing. Besides fruit maturity, method of packaging and distance of transportation influence the postharvest losses in strawberry [10,11], also postharvest management practices and storage conditions affects the shelf life and quality of fruits [12,13]. Strawberry is a non-climacteric fruit, implying its respiration rate and ethylene production does not increase during ripening [14,15]. During the ripening phase, the fruit continues to grow in size ~~grow~~ [16,17] and exhibits tissue softening rate, water loss, and susceptibility to physical damage during postharvest handling [12,18]. Due to the high metabolic activity of the fruit, the susceptibility to mechanical damage, physiological deterioration, water loss, and microbial spoilage, the quality of the fruit drops rapidly after

~~harvest~~harvest, and it tends to post-harvest losses [19]. Fruit softening occurs due to ~~the~~ disassembly of ~~the~~ cell wall as a result of ~~the~~ dissolution of middle lamellae during ripening [20]. Strawberry postharvest methods attempt to reduce respiration and water loss, while also preserving fruit firmness and limiting disease development. Various synthetic compounds have been used to extend strawberry postharvest life, however public concerns about fungicide residues have arisen. The negative consequences of pesticides on human health and the environment have prompted scientists to look for better fungicide alternatives [21]. In ~~the~~ past few decades, ~~researches~~research on the use of natural compounds in postharvest quality management of fruits have been gained attention among ~~the~~ scientific community and the ~~end-end~~-users. Among others, salicylic acid is one of ~~the~~ natural compounds ~~that~~ has been found beneficial in maintaining the quality of several fruits including ~~strawberry-strawberries~~ during storage [22-26]. ~~Present-The present~~ review encompasses the use of salicylic acid on postharvest fruit decay control of ~~strawberry strawberries~~ by salicylic acid applications.

### **Salicylic acid: a hormone with a diverse regulatory role in plants**

Salicylic acid is considered a potent plant hormone because it has a variety of regulatory effects in plant metabolism [27]. Salicylic acid (SA) and methyl salicylate (MeSA) play a central role in plant development, regulating stress response and disease resistance [28]. Under constantly changing environmental conditions, exogenous application of salicylic acid has been proven to be beneficial to crop growth and biological productivity, because of the key role of salicylic acid in photosynthesis, plant water relations, various enzyme activities, and its effect on the plants exposed to various biotic and abiotic stresses [29-31]. Plants are attacked by various pathogens, but plants protect themselves by activating certain types of defense mechanisms, such as locally acquired resistance (LAR) and systemic acquired resistance (SAR) against pathogens [32]. The effect of exogenous salicylic acid depends on many factors, such as plant type and developmental stage, application method and concentration of salicylic acid, and the endogenous content of salicylic acid in plants under given environmental conditions [30]. Salicylic acid has recently attracted special attention because it plays a key role in regulating the expression of multiple modes in plant responses to biological activities [33-35] and abiotic stresses [31,36,37]. The accumulation of large amounts of salicylic acid in the host is one of the mechanisms to protect it from pathogens. Therefore, salicylate plays an important role in the signal transduction pathway of plants and plays an important role in disease resistance [38] as the exogenous application of salicylic acid or acetylsalicylic acid induces the expression of pathogenic genes and confers resistance to pathogens [39-41].

### **Salicylic acid in mitigation of postharvest fruit decay**

Salicylic acid plays a key role in the signal transduction pathway leading to systemic acquired resistance (SAR), an enhancement of resistance against a broad spectrum of pathogens [34,42]. One reason for this is that salicylic acid induces a range of ~~defense~~ defense genes, most notably those encoding the pathogenesis-related (PR) proteins [43,44] and several of these PR proteins possess antimicrobial activities such as chitinase or  $\beta$ -1,3-glucanase activity [45-47]. Others, including members of the well-studied PR<sup>-1</sup> family, have no known biochemical activity but have been shown to inhibit ~~the~~ growth of oomycete pathogens and true fungi in vitro [48]. It has been established that ~~the~~ Salicylic acid also activates defense responses against pathogens and plays a role in the development of systemic acquired resistance [49,50]. According to Asghari and Aghdam [51], the application of salicylic acid, especially preharvest, for inducing the defense resistance systems against postharvest diseases may be a useful and promising measure for controlling postharvest decay on a commercial scale.

Strawberry fruit ~~are is~~ very delicate; they can get bruised during harvest and postharvest handling. Bruising is the most undesirable damage that seriously limits not only the appearance of fruits but also such fruits easily affected by fungal pathogens leading to decay and shorter shelf life. Fungi *Botrytis cinerea*, *Rhizopus stolonifer*, *Mucor* spp., *Colletotrichum* spp., *Penicillium* spp. are the main pathogens that cause post-harvest rot of strawberry fruits [52], and the gray mold caused by *Botrytis cinerea* is the most serious disease after harvest when conditions become favorable to disease development [53,54]. Although, a lot of effort has been put in by researchers for minimizing decay losses of fruits, the ~~low-low~~ temperature conditions around 1°C ~~are~~ considered ~~an~~ effective tool in postharvest quality prolongation. Different fungicides, soil fumigants, and bio-agents have been used to control the diseases [54-58] but most of these approaches have ~~an~~ unsavory impact due to the increment safe strains of pathogens against different chemical fungicides. Recently, few researchers worked on the improvement in plant defense mechanisms in fruits after harvest against the decay through ~~the~~ salicylic acid application.

Salicylic acid application as evaluated by researchers has advocated that the effective concentration is variable with doses, method of application, and the time of application. Postharvest dip treatment of fruit with salicylic acid has been found most effective than preharvest foliar sprays. Bablar *et al.* [21] studied the effect of foliar applied salicylic acid (1, 2, and 4 mmol L<sup>-1</sup>) on postharvest fungal decay and overall quality index in strawberry fruits and they observed that the application of salicylic acid at vegetative and fruit development stages followed by postharvest treatment of fruits effectively reduced fungal decay and retained postharvest storage quality of fruits when applied at with 1 or 2 mmol L<sup>-1</sup> while the postharvest treatment at 4 mmol L<sup>-1</sup> slightly damaged the fruits and less effective in retaining fruit quality. Shafiee *et al.* [26] studied the strawberry cv. Camarosa fruit quality after 7 days storage at 2 °C as influenced by salicylic acid addition to nutrient solution (0.03 mM) and postharvest treatments of salicylic acid. They observed that SA in their nutrient solution had less fruit decay and higher firmness of fruits. In ~~an~~ another trial conducted by Abolfazl *et al.* [59], pre- or postharvest application of salicylic acid (0, 3, 5, and 7 mM) resulted ~~in~~ less weight loss of fruits and delayed onset of the climacteric peak of respiration and also inhibited ethylene production, which intern enhanced the quality of fruit. Lolaei *et al.* [60] observed delayed ripening of strawberry cv. Camarosa by pre and postharvest salicylic acid application. Salari *et al.* [61] examined five levels of salicylic acid (0, 1, 2, 3, and 4 mM) on three cultivars (Paros, Kamarosa, and Selva) on the postharvest durability of fruits stored in ~~the~~ refrigerator at 3°C±1°C for 12 days. ~~Highest~~ ~~The highest~~ rotten fruits percentage was obtained from control and the effect of SA was independent with cultivars for healthy fruits and rotten fruits percentage. Mahsa *et al.* [62] tested methods of application (spray SA on fruits and paper disk method) and variable concentration of salicylic acid (0, 25, 50, and 100 µ L<sup>-1</sup>) on the postharvest durability and quality characteristics of strawberry fruit wherein fruits treated with salicylic acid resulted in lower decay as compared to control and the paper disk method showed ~~a~~ higher effect on fruit decay and quality resulted in the longer storability.

Salicylic acid as ~~a~~ foliar spray at 3-4 leaf stage in the spring growth of strawberry and again 15 days after 1<sup>st</sup> spray found that salicylic acid treatments were effective in improving storability of fruits compared with control as it resulted ~~in a~~ significantly minimum physiological loss in weight and rotting percent, and retained higher values for biochemical parameters of fruits at 2, 4 and 6 ~~daydays~~ of storage at ambient conditions [64]. Yousef [65] emphasized that ~~the~~ salicylic acid can be considered ~~ed~~ one of the ~~most safesafestty~~ ways with ~~less-fewer~~ environmental hazards to improve the postharvest life and maintaining ~~the~~ quality of ~~strawberry-strawberries~~ and reducing the rate of rachis browning during postharvest storage. Preharvest foliar sprays of salicylic acid ~~are~~ also reported to be effective in postharvest quality management of fruit. SA 2 mM as pre- and postharvest application ~~was~~

found to be effective in prolonging the storage life of strawberry fruits by reducing the fruit spoilage and fungal decay up to 16 days of storage [65]. Postharvest dipping of dipping strawberry fruits after harvest in salicylic acid solution (2 and 4 mM) was found effective in suppression of the fungal growth on fruit [66]. Salicylic acid treatment with calcium was also found beneficial in postharvest quality retention of fruits. Niazi *et al.* [67] examined postharvest dip of fruits in 1 mM salicylic acid along with 2% calcium chloride, and a combination of salicylic acid and calcium chloride at two water temperatures of 20°C (cold water treatment) and 45°C (hot water treatment) for 5 min and then stored at 4°C for 14 days. They observed that the Salicylic acid and CaCl<sub>2</sub> along with hot water treatment maintained the fruit quality more efficiently during storage.

## CONCLUSION

Salicylic acid is a potential plant hormone having a variety of regulatory effects in plant development, regulating stress response and disease resistance. Exogenous application of salicylic acid has been proven to be beneficial in minimizing fruit decay and maintaining the fruit quality suggests that it has a potential role in postharvest management of strawberry fruits. Further research works in this direction are needed to validate the reliability of the applicability of applicability on a commercial scale.

## CONSENT

Not applicable

## ETHICAL APPROVAL

## REFERENCES

1. Hummer KE, Bassil N, Njuguna W. *Fragaria*. In: Cole C, Editor. Wild crop relatives: genomic and breeding resources - temperate fruits. New York: Springer; 2011. [https://doi.org/10.1007/978-3-642-16057-8\\_2](https://doi.org/10.1007/978-3-642-16057-8_2).
2. Darrow, GM. The Strawberry: History, Breeding and Physiology, Holt, Rinehart and Winston, New York; 1966.
3. Hancock HF. Strawberries. In: Erez A, Editor. Temperate fruit crops in warm climates. Dordrecht: Kluwer Academic Publishers; 2000.
4. Stewart PJ. *Fragaria*: history and breeding. In: Folta KM, Kole C, Editors. Genetics, Genomics, and Breeding of Berries. Science Publishers, New Hampshire, USA; 2011 pp. 114-137. <https://doi.org/10.1201/b10922>.
5. Sharma VP, Sharma RR. The Strawberry. ICAR Publications, New Delhi; 2004.
6. Ali A, Gaur GS. Effect of organic mulches on runner production of strawberry. Asian Journal of BioScience. 2013;8(2):175-179.
7. Seeram NP. Berry fruits: Compositional elements, biochemical activities, and the impact of their intake on human health, performance, and disease. Journal of Agricultural and Food Chemistry. 2008;56:627-629. DOI: <https://doi.org/10.1021/jf071988k>.
8. Agehara S, Nunes MCDN. Season and nitrogen fertilization effects on yield and physicochemical attributes of strawberry under subtropical climate conditions. Agronomy. 2021;11:1391. <https://doi.org/10.3390/agronomy11071391>.
9. Giampieri F, Tulipani S, Alvarez-Suarez, JM, Quiles JL, Mezzetti B, Battino M. The strawberry: composition, nutritional quality, and impact on human health. *Nutrition*, 2012;28:9-19. <https://doi.org/10.1016/j.nut.2011.08.009>.

10. Duran M, Aday MS, Zorba NND, Temizkan R, Büyükcan MB, Caner C. Potential of antimicrobial active packaging containing natamycin, nisin, pomegranate and grape seed extract in chitosan coating to extend shelf life of fresh strawberry. *Food and Bioproducts Processing*, 2016;98:354-363. <https://doi.org/10.1016/j.fbp.2016.01.007>.
11. Matar C, Guillard V, Gauche G, Costa S, Gontard N, Guilbert S, Gaucel S. Consumer behaviour in the prediction of postharvest losses reduction for fresh strawberries packed in modified atmosphere packaging. *Postharvest Biology and Technology*. 2020;163:111-119. <https://doi.org/10.1016/j.postharvbio.2020.111119>.
12. Kader AA. Quality and its maintenance in relation to the postharvest physiology of strawberry. In: Luby JJ, Dale A, Editors. *The Strawberry into the 21<sup>st</sup> Century*. Portland, Oregon: Timber Press; 1991. pp.145-152.
13. Kader AA. Postharvest technology of horticultural crops. *Ethiopian Journal of Applied Science Technology*, 2013;1:1-8.
14. Biale JB, Young RE. Respiration and ripening in fruit retrospect– and prospect. In: Friend J, Rhodes MJC, Editors. *Recent advances in the biochemistry of fruits and vegetables*. London: Academic Press; 1981.
15. Perkins-Veazie PM, Huber DJ, Brecht JK. *In vitro* growth and ripening of strawberry fruit in the presence of ACC, STS, or propylene. *Annals of Applied Biology*. 1996;128:105-116. <https://doi.org/10.1111/j.1744-7348.1996.tb07094.x>.
16. Perkins-Veazie P, Huber DJ. Growth and ripening of strawberry fruit under field conditions. *Proceedings of the Florida State Horticultural Society*. 1987;100:253-256.
17. Abeles FB, Takeda F. Cellulase activity and ethylene in ripening strawberry and apple fruits. *Scientia Horticulturae*. 1990;42:269-275. [https://doi.org/10.1016/0304-4238\(90\)90050-O](https://doi.org/10.1016/0304-4238(90)90050-O).
18. Kader AA. Fruit maturity ripening and quality relationships. *Acta Horticulturae*. 1999;485:203-208. <https://doi.org/10.17660/ActaHortic.1999.485.27->.
19. Cordenunsi BR, Genovese MI, Nascimento JRO, Hassimotto NMA, Santos RJ, Lajolo FM. Effects of temperature on the chemical composition and antioxidant activity of three strawberry cultivars. *Food Chemistry*. 2005;91:113-121. <https://doi.org/10.1016/j.foodchem.2004.05.054>.
20. Paniagua C, Pose S, Morris V, Kirby A, Quesada M, Mercado J. Fruit softening and pectin disassembly: An overview of nan-structural pectin modifications assessed by atomic microscopy. *Annals of Botany*. 2014;114:1375-1383. <https://doi.org/10.1093/aob/mcu149>.
21. Babalar M, Asghari M, Talaei A, Khosroshahi A. Effect of pre and postharvest salicylic acid treatment on ethylene production, fungal decay, and overall quality of Selva strawberry fruit. *Food Chemistry*. 2007;105:449-453. <https://doi.org/10.1016/j.foodchem.2007.03.021>.
22. Romani RJ, Hess BM, Leslie CA. Salicylic acid inhibition of ethylene production by apple discs and other plant tissues. *Journal of Plant Growth Regulator*. 1989;8:63-69.
23. Zeng KF, Cao JK, Jiang WB. Enhancing disease resistance in harvested mango (*Mangifera indica* L. cv. Matisu) fruit by salicylic acid. *Journal of the Science of Food and Agriculture*. 2006;86:694-698. <https://doi.org/10.1002/jsfa.2397>.
24. Wang L, Chen S, Kong W, Li S, Archbold DD. Salicylic acid pre-treatment alleviates chilling injury and affects the antioxidant system and heat shock proteins of peaches during cold storage. *Postharvest Biology and Technology*. 2006;41:244-251. <https://doi.org/10.1016/j.postharvbio.2006.04.010>.
25. Mo Y, Gong D, Liang G, Han R, Xie J, Li W. Enhanced preservation effects of sugar apple fruits by salicylic acid treatment during postharvest storage. *Journal of the Science of Food and Agriculture*. 2008;88:2693-2699. <https://doi.org/10.1002/jsfa.3395>.
26. Shafiee M, Taghavi TS, Babalar M. Addition of salicylic acid to nutrient solution combined with postharvest treatments (hot water, salicylic acid, and calcium dipping)

- improved postharvest fruit quality of strawberry. *Scientia Horticulturae*. 2009;124:40-45. <https://doi.org/10.1016/j.scienta.2009.12.004>.
27. Raskin I. Role of salicylic acid in plants. *Annual Review of Plant Physiology and Plant Molecular Biology*. 1992;43:439-463. <https://doi.org/10.1146/annurev.pp.43.060192.002255>.
  28. Klessig DF, Malamy J. The salicylic acid signal in plants. *Plant Molecular Biology*. 1994;26:1439-1458. <https://doi.org/10.1007/BF00016484>.
  29. Senaratna T, Touchell D, Bunn E, Dixon K. Acetyl salicylic acid (aspirin) and salicylic acid induce multiple stress tolerance in bean and tomato plants. *Plant Growth Regulators*. 2000;30:157-161. <https://doi.org/10.1023/A:1006386800974>.
  30. Horvath E, Szalai S, Janda T. Induction of abiotic stress tolerance by salicylic acid signaling. *Journal of Plant Growth Regulators*. 2007;26:290-300. <https://doi.org/10.1007/s00344-007-9017-4>.
  31. Hayat Q, Hayat H, Irfan M, Ahmad A. Effect of exogenous salicylic acid under changing environment: A review. *Environmental and Experimental Botany*. 2010;68:14-25. <https://doi.org/10.1016/j.envexpbot.2009.08.005>.
  32. Vlot AC, Dempsey DA, Klessig DF. Salicylic acid, a multifaceted hormone to combat disease. *Annual Review of Phytopathology*. 2009;47:177-206. <https://doi.org/10.1146/annurev.phyto.050908.135202>.
  33. Malamy J, Klessig DF. Salicylic acid and plant disease resistance. *Plant Journal*. 1992;2:643-654. <https://doi.org/10.1111/j.1365-3113X.1992.tb00133.x>.
  34. Ryals JA, Neuenschwander UH, Willits MG, Molina A, Steiner HY, Hunt MD. Systemic acquired resistance. *Plant Cell*. 1996;8:1809-1819. <https://doi.org/10.2307/3870231>.
  35. Shah J. Lipids, lipases and lipid-lipid-modifying enzymes in plant disease resistance. *Annual Reviews of Phytopathology*. 2005;43:229-260. <https://doi.org/10.1146/annurev.phyto.43.040204.135951>.
  36. Ding CK, Wang CY, Gross KC, Smith DL. Reduction of chilling injury and transcript accumulation of heat shock protein genes in tomatoes by methyl jasmonate and methyl salicylate. *Plant Science*. 2001;161:1153-1159. [https://doi.org/10.1016/S0168-9452\(01\)00521-0](https://doi.org/10.1016/S0168-9452(01)00521-0).
  37. Ding C, K, Wang C Y. The dual effects of methyl salicylate on ripening and expression of ethylene biosynthetic genes in tomato fruit. *Plant Science*. 2003;164:589-596. [https://doi.org/10.1016/S0168-9452\(03\)00010-4](https://doi.org/10.1016/S0168-9452(03)00010-4).
  38. Park SW, Kaimoyo E, Kumar D, Mosher S, Klessig DF. Methyl salicylate is a critical mobile signal for plant systemic acquired resistance. *Science*. 2007;318:113-116. <https://doi.org/10.1126/science.1147113>.
  39. Morris SW, Vernooij B, Titaram S, et al. Induced resistance responses in maize. *Molecular Plant-Microbe Interactions*. 1998;11:643-658. <https://doi.org/10.1094/MPMI.1998.11.7.643>.
  40. Pasquer F, Isidore E, Zarn J, Keller B. Specific patterns of changes in wheat gene expression after treatment with three antifungal compounds. *Plant Molecular Biology*. 2005;57:693-707.
  41. Makandar R, Essig JS, Schapaugh MA, Trick HN, Shah J. Genetically engineered resistance to Fusarium head blight in wheat by expression of *Arabidopsis NPR1*. *Molecular Plant-Microbe Interactions*. 2006;19:123-129. <https://doi.org/10.1094/MPMI-19-0123>.
  42. Murphy AM, Chivasa S, Singh DP, Carr JP. Salicylic acid-induced resistance to viruses and other pathogens: a parting of the ways? *Trends in Plant Science*. 1999;4:155-160. [https://doi.org/10.1016/s1360-1385\(99\)01390-4](https://doi.org/10.1016/s1360-1385(99)01390-4).
  43. Bowles DJ. Defense-related proteins in higher plants. *Annual Review of Biochemistry*. 1990;59:873-907. <https://doi.org/10.1146/annurev.bi.59.070190.004301>.
  44. Carr JP, Klessig DF. The pathogenesis-related proteins of plants. In: Setlow J, Editors. *Genetic engineering, principles, and methods*. New York: Plenum Press; 1989.

45. Schlumbaum A, Mauch F, Vogeli U, Boller T. Plant chitinases are potent inhibitors of fungal growth. *Nature*. 1986;324:365-367. <https://doi.org/10.1038/324365a0>.
46. Mauch F, Hadwiger LA, Boller T. Antifungal hydrolases in pea tissue 1. Purification and characterization of 2 chitinases and 2 beta-1,3-glucanases differentially regulated during development and in response to fungal infection. *Plant Physiology*. 1988;87:325-333. <https://doi.org/10.1104/pp.87.2.325>.
47. Salzman RA, Tikhonova I, Bordelon BP, Hasegawa PM, Bressan RA. Coordinate accumulation of antifungal proteins and hexoses constitutes a developmentally controlled defense response during fruit ripening in grapes. *Plant Physiology*. 1998;117:465-472. <https://doi.org/10.1104/pp.117.2.465>.
48. Niderman T, Genetet I, Bruyere T, Gees R, Stintzi A, Legrand M, et al. Pathogenesis related PR1 proteins are antifungal isolation and ~~characterisation~~ **characterization** of 3 14-kilodalton proteins of tomato and of a basic PR-1 of tobacco with inhibitory activity against *Phytophthora infestans*. *Plant Physiology*. 1995;108:17-27. <https://doi.org/10.1104/pp.108.1.17>.
49. Vernooij B, Friedrich L, Morse A, Reist R, Kolditz-Jawhar R, Ward E, et al. Salicylic acid is not the translocated signal responsible for inducing systemic acquired resistance but is required in signal transduction. *Plant Cell*, 1994;6(7):959-965. <https://doi.org/10.1105/tpc.6.7.959>.
50. Mauch MB, Metraux JP. Salicylic acid and systemic acquired resistance to pathogen attack. *Annals of Botany*, 1998;82:535-540. <https://doi.org/10.1006/anbo.1998.0726>
51. Asghari M, Aghdam MS. Impact of salicylic acid on post-harvest physiology of horticultural crops. *Trends in Food Science and Technology*. 2010;21:502-509. <https://doi.org/10.1016/j.tifs.2010.07.009>.
52. Feliziani E, Romanazzi G. Postharvest decay of strawberry fruit: Etiology, epidemiology, and disease management. *Journal of Berry Research*. 2016;6:47-63. DOI:10.3233/JBR-150113. <https://doi.org/10.3233/JBR-150113>.
53. Petrasch S, Knapp SJ, van-Kan JAL, Blanco-Ulate B. Grey mould of strawberry, a devastating disease caused by the ubiquitous necrotrophic fungal pathogen *Botrytis cinerea*. *Molecular Plant Pathology*. 2019;20(6):877-892. <https://doi.org/10.1111/mpp.12794>.
54. Zavala JFA, Wang SY, Wang CY, Aguilar GAG. Effect of storage temperatures on antioxidant capacity and aroma compounds in strawberry fruit. *Food Science and Technology*. 2004;37:687-695. <https://doi.org/10.1016/j.lwt.2004.03.002>.
55. LaMondia JA, Douglas SM. Sensitivity of *Botrytis cinerea* from Connecticut greenhouses to benzimidazole and dicarboximide fungicides. *Plant Disease*. 1997;81:729-732. <https://doi.org/10.1094/PDIS.1997.81.7.729>.
56. Kim JH, Lee SH, Kim CS, Lim EK, Choi KH, Kong HG, et al. Biological control of strawberry gray mold caused by *Botrytis cinerea* using *Bacillus licheniformis* N1 formulation. *Journal of Microbiology and Biotechnology*. 2007;17(3):438-44.
57. Deising HB, Reimann S, Pascholati SF. Mechanisms and significance of fungicide resistance. *Brazilian Journal of Microbiology*. 2008;39(2):286-295. <https://doi.org/10.1590/S1517-83822008000200017>.
58. Boutin C, Strandberg B, Carpenter D, Mathiassen SK, Thomas PJ. Herbicide impact on non-target plant reproduction: What are the toxicological and ecological implications? *Environmental Pollution*. 2014;185:295-306. <https://doi.org/10.1016/j.envpol.2013.10.009>.
59. Abolfazl L, Behzad K, Mohammad AR, Mojtaba KR, Rana M. Effect of Pre and postharvest treatment of salicylic acid on ripening of fruit and overall quality of strawberry (*Fragaria ananassa* Duch. cv. Camarosa) fruit. *Annals of Biological Research*. 2012;3(10):4680-4684.
60. Lolaei, A., B. Kaviani, M.A. Rezaei, M.K. Raad, and R. Mohammadipour. Effect of pre and postharvest treatment of salicylic acid on ripening of fruit and overall quality of

- strawberry (*Fragaria ananassa* Duch cv. Camarosa) fruit. *Annals of Biological Research*. 3(10): 4680-4684.
61. Salari N, Bahraminejad A, Afsharmanesh G, Khajehpour G. Effect of salicylic acid on post-harvest quantitative and qualitative traits of strawberry Cultivars. *Advances in Environmental Biology*. 2013;7(1):94-99.
  62. Mahsa G, Sadegh S, Vahid A. Extending postharvest longevity and improving quality of strawberry (*Fragaria ananassa* Duch cv. 'Gaviota') fruit by postharvest salicylic acid treatment. *Journal of Agricultural Studies*. 2015;3(2):17-36. <https://doi.org/10.5296/jas.v3i2.7274>.
  63. Baba TR. Effect of Exogenous application of salicylic acid and triacontanol on growth, yield, and quality of strawberry (*Fragaria x ananassa* Duch.) cv. Camarosa. M.Sc. Thesis, submitted to SKUAST-Kashmir, Srinagar, J&K, India, 2016.
  64. Yousif DYM. Evaluation of salicylic acid solution on fungus *Botrytis cinerea* that caused strawberry gray mold. *Plant Archives*. 2019;19(1): 229-238.
  65. Kumar S, Kaur G. Effect of pre and post-harvest applications of salicylic acid on quality attributes and storage behavior of strawberry cv. Chandler. *Journal of Pharmacognosy and Phytochemistry*. 2019;8(4):516-522.
  66. El-Mogy MM, Ali MR, Darwish OS, Rogers HJ. Impact of salicylic acid, abscisic acid, and methyl jasmonate on postharvest quality and bioactive compounds of cultivated strawberry fruit. *Journal of Berry Research*. 2019;9(2):333-348. <https://doi.org/10.3233/JBR-180349>.
  67. Niazi AR, Ghanbari F, Moghadam JE. Simultaneous effects of hot water treatment with calcium and salicylic acid on shelf life and qualitative characteristics of strawberry during refrigerated storage, *Journal of Food Processing and Preservation*. 2020;45(10):e15005 1-10. <https://doi.org/10.1111/jfpp.15005>.