

## Original Research Article

# Antioxidant and Antimicrobial Activities of Various Extracts from Safou (*Dacryodes edulis*) Cultivated in Côte d'Ivoire

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### ABSTRACT

Safou (*Dacryodes edulis*) is a fruit with very interesting nutritional and pharmacological properties. However, this fruit remains rather unknown to the Ivorian population.

**Aims:** The objective of this study was to highlight the antioxidant and antimicrobial potential of the seed and pulp of Safou (*Dacryodes edulis*) cultivated in Côte d'Ivoire for its valorization.

**Study design:** Microbiological and biochemical studies

**Place and Duration of Study:** Laboratory of Biotechnology and Food Microbiology, Abidjan, Côte d'Ivoire, between September 2020 and February 2021.

**Methodology:** Aqueous, ethanolic and methanolic extracts of the pulp and seed of Safou were prepared. Moisture content, pH, yield, total polyphenols, diphényl-2-picrylhydrazyle (DPPH) radical scavenging test of the different extracts were determined. Aqueous, ethanolic and methanolic extracts of the seed and pulp at varying concentrations of 200 mg/mL, 100 mg/mL, 50 mg/mL, 25 mg/mL, 12.5 mg/mL and 6.25 mg/mL was tested against some selected human pathogens such as *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Aspergillus fumigatus* and *Penicillium chrysogenum*.

**Results:** pH, moisture content and yield ranged from 3.16 to 4.74; 9 to 18.30%; and 10.79 to 18.78%, respectively. Total polyphenol content ranged from 1.45 mgEAG/g to 4.56 mgEAG/g. The highest levels of total polyphenols were observed for the methanolic extracts of pulp. The different extracts strongly scavenged the DPPH radical with percentages of anti-free radical activity that varied from 50.76 to 64.43%. The antimicrobial susceptibility results of the methanolic extracts of *D. edulis* seed showed highest zones of inhibition to the microbial isolates tested. The organisms were resistant to the aqueous extracts of both the pulp and seed.

**Conclusion:** The presence of bioactive compounds and the antimicrobial activity of Safou extracts could justify its use in the food and pharmaceutical fields.

*Keywords:* Safou (*Dacryodes edulis*), total polyphenols, antioxidant activity, antimicrobial activity.

## 1. INTRODUCTION

Plants are used in developing and developed countries for both dietary and medicinal reasons. According to a World Health Organization (WHO) report, 80% of the world's populations rely primarily on traditional therapies that involve the use of plant extracts or their active substances [1]. Among these plants, there is the safoutier (*Dacryodes edulis* (G. Don) H. J. Lam, Burseraceae) native to the countries of Central Africa and the Gulf of Guinea (Cameroon, Congo, Democratic Republic of Congo, Gabon, Equatorial Guinea and Nigeria) [2]. The different parts of this plant are used in many parts of Africa in the treatment of several diseases. The leaves are reportedly used to cure ringworm, scabies, and skin rashes [3]. The bark is said to be used to treat dysentery and anemia [4]. *D. edulis* fruit (Safou) has shown excellent nutritional qualities and interesting agro-industrial properties [5]. Its pulp and seed are rich in fatty acids and amino acids [6]. Nwokonkwo [7] in her study reported that the seed extracts of *Dacryodes edulis* and its secondary metabolites possessed potential antibacterial activity against some human pathogens in varying degrees. Antibacterial effect of the essential oil as well as the organic extracts of *D. edulis* plant has been reported [8,3]. It was indicated that the leaf extract of *Dacryodes edulis* possess broad spectrum antibacterial activity in a recent study [9].

In Côte d'Ivoire, the first Safou (*D. edulis*) trees were introduced before 1960s but disappeared [10]. The second introduction of Safou trees comes from Cameroon Safou seeds in 1980 [11]. However, few scientific works have been published either on their nutritional and therapeutic potential. This study investigated the antioxidant and antimicrobial potential of the seed and pulp of Safou (*Dacryodes edulis*) cultivated in Côte d'Ivoire necessary for their wider use.

## 2. MATERIAL AND METHODS

### 2.1 Sample Collection

*Dacryodes edulis* fruits (Safou) were harvested in the south of Côte d'Ivoire (Azaguié-blida, Agboville) from September to December 2020. The fruits were transported in net bags to Biocatalysis and Bioprocessing laboratory of NANGUI ABROGOUA University.

### 2.2 Preparation of Extract

The fruits were de-fleshed to separate the seed from the pulp. The fresh seed and pulp Safou plant were dried in an oven 45°C/ 72h. The seed and pulp of the plant were then grind to fine powder with the aid of a mechanical grinder. Ethanol, methanol and water were the extracting solvents used in extracting the phytochemicals from *D. edulis* [12] modified. Fifty gram each of the powdered seed and pulp were soaked in 500 ml of the solvents (ethanol, methanol and water) in separate beakers and the set up allowed to run for 24 hours. After 24 hours of extraction, the suspensions were vigorously shaken and filtered with Whatmann No 1 filter paper. Ethanolic and methanolic solvents were evaporated at 40°C using a rotary evaporator. The concentrated extracts were stored in airtight bottles and labelled.

### 2.3 Analysis of physico-chemical constants

Extraction yield [13], moisture content [14], pH [15] as well as the color and texture of the different Safou extracts were determined.

### 2.4 Antioxidant Assay

#### 2.4.1 2,2-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay

The free radical scavenging activities of the crude extracts of *D. edulis* were evaluated using the DPPH (2, 2-diphenyl- 1-picrylhydrazyl) assay method as described by [16]. Fifty (50) µL of each methanolic solution of the extracts at different concentrations (from 0.01 to 0.2 mg/mL) is added to 1.95 mL of the DPPH methanolic solution (0.025 g/L). At the same time, a negative control is prepared by mixing 50 µL of methanol with 1.95 mL of the DPPH methanolic solution. The absorbance reading is taken against a blank prepared for each

concentration at 517 nm after 30 min of incubation in the dark at room temperature. Ascorbic acid (Vitamin C) was used as a control. Each assay was done in triplicate and the results, recorded as the mean  $\pm$  SD of the three findings, and were illustrated in a tabular form. The radical scavenging activity (RSA, %) was calculated as follows:  $RSA (\%) = [(A_{DPPH} - A_{sample}) / A_{DPPH}] \times 100$  (where A = Absorbance). The radical scavenging percentages were plotted against the logarithmic values of the concentration of test samples and a linear regression curve was established in order to calculate IC<sub>50</sub>, which is the amount of sample necessary to inhibit 50% of free radical DPPH.

#### **2.4.1 Total phenols contents (TPC)**

TPC of different extracts of Safou was determined using the Folin-Ciocalteu reagent method described by [17]. A volume of two hundred (200)  $\mu$ L of each extract was mixed with 1 mL of Folin-Ciocalteu reagent. The mixture was thoroughly homogenized by manual shaking. After 3 min, a volume of 1 mL of 20% aqueous sodium carbonate solution was added and the volume was adjusted to 10 mL with distilled water. The mixture was placed in the dark for 30 min and the absorbance was measured at 765 nm with a spectrophotometer against the blank. A calibration curve was established with a concentration range of gallic acid solution from 0 to 0.1 mg/mL. The total phenol content was expressed as milligram equivalent of gallic acid per gram of extract (mgGAE/g).

### **2.5 Determination of the antimicrobial activity of different Safou extracts**

#### **2.5.1 Microbial strains**

*Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Proteus mirabilis* and *Aspergillus fumigatus*, *Penicillium chrysogenum* were collected from Pasteur Institute of Côte d'Ivoire and Laboratory of Biotechnology and Food Microbiology of NANGUI ABROGOUA University respectively.

#### **2.5.2 Preparation of the microbial suspensions**

The density of selected organisms was adjusted equal to that of the 0.5 McFarland standards ( $1.5 \times 10^8$  CFU/mL) by adding them to nutrient broth for bacteria and Sabouraud broth for molds. A 24 h old culture was used for the preparation of microbial suspension. McFarland standards were used as a reference to adjust the turbidity of microbial suspension so that the number of microorganisms would be within a given range.

#### **2.5.3 Antimicrobial susceptibility test**

The National Committee for Clinical Laboratory Standards was the reference in determining antimicrobial susceptibility through the Well Diffusion Method. The different extracts of Safou were tested using standard Mueller Hinton II plates to detect the antimicrobial activity of these testing solutions [18]. The plates with inoculated microorganisms were provided with 100  $\mu$ L aliquots in 6 mm diameter punched wells. The surface of the agar plate was streaked over the entire sterile agar surface rotating the plate to ensure an even distribution of inoculum with a final swab around the rim. All of the plates were placed in the incubator set at a temperature of 37°C/ 24h. The antibacterial and antifungal activities of methanolic, ethanolic and aqueous extract of Safou were measured using a ruler by determining the zone of inhibition. This was carried out in at least an average of three parallel independent trials. A substance is said to be ineffective if the diameter of inhibition is less than 8 mm while it is said to be effective if the diameter is between 9 and 14 mm. It is considered very effective when the diameter is between 15 and 19 mm and then extremely effective if the diameter is greater than 20 mm.

#### **2.5.4 Determination of the Minimum Inhibitory Concentration (MIC)**

MIC determination was performed with a series of concentrations of the different extracts: 200 mg/mL, 100 mg/mL, 50 mg/mL, 25 mg/mL, 12.5 mg/mL and 6.25 mg/mL. The plates with

inoculated microorganisms were provided with 100  $\mu$ L aliquots in 6 mm diameter punched wells. The MIC was obtained from the lowest concentration of the plant extract for which no bacterial and fungal colonies were obtained [19].

## 2.6 Statistical analysis

The one-way analysis of variances (ANOVA) was carried out with the XLSTAT software to compare the variables measured on the different hydroalcoholic products. This software was used to calculate the means and standard deviations of the analysis parameters

## 3. RESULTS

### 3.1 Physicochemical properties of Safou (*Dacryodes edulis*) extracts

The texture and color of the different extracts vary according to the type of extract. The aqueous extracts of pulp and seed have a black pasty texture. While the methanolic and ethanolic extracts of the pulp are pasty and green and those of the seed show a brown pasty texture. The yields of methanolic, ethanolic and aqueous extracts of safou pulp are 10.79%, 17.39% and 18.8% respectively. Those of the seed are 10.79%, 10.89% and 10.91% respectively. The moisture contents of the pulp extracts (methanolic, ethanolic and aqueous) are 9.22%, 9.40% and 19.00% respectively. Those of the seed are 18.30%, 13.09% and 15.34% respectively. pH of the methanolic, ethanolic and aqueous extracts of the pulp and seed of Safou is acidic. pH is 3.43, 3.16 and 4.24 for the methanolic, ethanolic and aqueous extracts of the pulp; and 3.68, 3.57 and 4.74 for those of the seed respectively (Table 1).

Table 1: Physico-chemical parameters of Safou (*Dacryodes edulis*) extracts

Extract	Texture	Color	Yields (%)	pH	Moisture content (%)
MEP	pasty	green	17.07 $\pm$ 0.61 <sup>a</sup>	3.43 $\pm$ 0.12 <sup>a</sup>	9.22 $\pm$ 1.53 <sup>a</sup>
MES	pasty	brown	10.79 $\pm$ 0.58 <sup>b</sup>	3.68 $\pm$ 0.01 <sup>a</sup>	18.30 $\pm$ 1.55 <sup>b</sup>
EEP	pasty	green	17.39 $\pm$ 1.06 <sup>a</sup>	3.16 $\pm$ 0.06 <sup>a</sup>	9.40 $\pm$ 1.27 <sup>a</sup>
EES	pasty	brown	10.89 $\pm$ 2.11 <sup>b</sup>	3.57 $\pm$ 0.04 <sup>a</sup>	13.09 $\pm$ 1.40 <sup>c</sup>
AEP	pasty	black	18.78 $\pm$ 2.23 <sup>c</sup>	4.24 $\pm$ 0.01 <sup>b</sup>	9.00 $\pm$ 1.98 <sup>a</sup>
AES	pasty	black	10.91 $\pm$ 0.78 <sup>b</sup>	4.74 $\pm$ 0.03 <sup>b</sup>	15.34 $\pm$ 1.27 <sup>d</sup>

In column, the averages affected by the same letter are not significantly different at the 5% threshold according to the Newmann-Keuls test. EEP= ethanolic extract pulp; MEP: methanolic extract pulp; AEP: aqueous extract pulp; EES: ethanolic extract seed; MES: methanolic extract seed; AES: aqueous extract seed

### 3.2 Antioxidant activity of Safou (*Dacryodes edulis*) extracts

The table 2 showed antioxidant activity of methanolic, ethanolic and aqueous extracts of Safou. The highest levels of total polyphenol were observed for the methanolic extracts of the pulp with 4.56 mgGAE/g and of the seed with 4.03 mgGAE/g followed by the ethanolic extracts of the seed with 3.93 mg GAE/g and of the pulp with 2.93 mgGAE/g). The lowest

contents of total polyphenols were obtained with the aqueous extracts of pulp with 1.98 mg GAE/g and of seed with 1.45 mg GAE/g.

All the methanolic, ethanolic and aqueous extracts of Safou have high antioxidant power. Methanolic extracts of pulp (64.43), ethanolic extracts of seed (62.68), methanolic extracts of seed (60.03) and ethanolic extracts of pulp (58.96) of Safou had the highest activities.

The median inhibitory concentration (IC<sub>50</sub>) values of Safou extract range from 12.5 to 21 µg/mL. The highest values were obtained with extracts from Safou seeds.

Table 2: Antioxidant activity of Safou (*Dacryodes edulis*) extracts

Extract	Total polyphenol (mgGAE/g)	Percentage of DPPH radical inhibition (%)	IC <sub>50</sub>
MEP	4.56 ± 0.08 <sup>a</sup>	64.43 ± 0.40 <sup>a</sup>	13.5 <sup>a</sup>
MES	4.03 ± 0.07 <sup>a</sup>	60.03 ± 0.51 <sup>b</sup>	21 <sup>b</sup>
EEP	2.93 ± 0.23 <sup>b</sup>	58.96 ± 0.91 <sup>c</sup>	14.6 <sup>a</sup>
EES	3.93 ± 0.04 <sup>c</sup>	62.68 ± 0.22 <sup>a</sup>	19.5 <sup>b</sup>
AEP	1.98 ± 0.09 <sup>d</sup>	53.95 ± 0.30 <sup>d</sup>	12.5 <sup>a</sup>
AES	1.45 ± 0.18 <sup>d</sup>	50.76 ± 0.12 <sup>e</sup>	18.7 <sup>b</sup>
Vit C	-	90.02	6

In column, the averages affected by the same letter are not significantly different at the 5% threshold according to the Newmann-Keuls test. EEP= ethanolic extract pulp; MEP: methanolic extract pulp; AEP: aqueous extract pulp; EES: ethanolic extract seed; MES: methanolic extract seed; AES: aqueous extract seed

### 3.3 Antimicrobial activity of Safou (*Dacryodes edulis*) extract

The results showed that methanolic and ethanolic extract of *D. edulis* showed good, noticeable, and remarkable activity against the test organisms as compared with aqueous extract while no zone of inhibition was observed on *Pseudomonas aeruginosa* (Table 3). Methanolic extracts showed highest zone of inhibition with *Escherichia coli*, *Staphylococcus aureus*, *Klebsiella pneumoniae* and *Proteus mirabilis*. The aqueous extracts of Safou showed no zone of inhibition when tested against all the collected microorganisms isolates.

Table 3: Antimicrobial activity of Safou (*Dacryodes edulis*) extract: diameter of zones of inhibition (mm)

Extract	<i>Escherichia coli</i>	<i>Staphylococcus aureus</i>	<i>Pseudomonas aeruginosa</i>	<i>Klebsiella pneumoniae</i>	<i>Proteus mirabilis</i>	<i>Aspergillus fumigatus</i>	<i>Penicillium</i>
MEP	17 ± 0.4	16 ± 0.2	-	12 ± 0.5	9 ± 0.4	4 ± 0.2	5 ± 0.2
MES	20 ± 0.6	18 ± 0.2	-	16 ± 0.3	10 ± 0.3	5 ± 0.1	6 ± 0.1

<b>EEP</b>	10 ± 0.5	12 ± 0.4	-	8 ± 0.3	4 ± 0.1	-	-
<b>EES</b>	14 ± 0.4	15 ± 0.4	-	10 ± 0.3	5 ± 0.1	-	-
<b>AEP</b>	-	-	-	-	-	-	-
<b>AES</b>	-	-	-	-	-	-	-

EEP= ethanolic extract pulp; MEP: methanolic extract pulp; AEP: aqueous extract pulp; EES: ethanolic extract seed; MES: methanolic extract seed; AES: aqueous extract seed

### 3.4 Minimum Inhibitory Concentrations (MICs) of Safou (*Dacryodes edulis*) extracts

The methanolic extract gave the lowest MIC value compared to the ethanolic extract. The MIC values ranged from 12.5 - 100 mg/ml for methanolic extract and 50–200 mg/ml for ethanolic extract as depicted in table 4.

Table 4: Minimum Inhibitory Concentrations (MICs) of methanolic and ethanolic extract of Safou(*Dacryodes edulis*) against the test isolates

microorganisms	MIC (mg/ml)			
	MEP	MES	EEP	EES
<i>Escherichia coli</i>	12.5	12.5	25	25
<i>Staphylococcus aureus</i>	50	50	50	50
<i>Klebsiella pneumoniae</i>	100	100	200	200
<i>Proteus mirabilis</i>	-	100	-	-

EEP= ethanolic extract pulp; MEP: methanolic extract pulp; EES: ethanolic extract seed; MES: methanolic extract seed;

### 3. DISCUSSION

The results of this study showed that the yields vary depending on the part of the fruit and the type of extract. The difference in yield between extracts could be explained by the chemical composition which differs from one extract to another as well as by the type of solvent used. Indeed, several authors such as [20, 21] showed that the extraction efficiency was influenced by several parameters such as the nature of the phytochemicals, the method used for extraction, the temperature, the extraction time, the particle size as well as the solvent used. pH of the different extracts of Safou is acidic. According to [22] an acidic pH is recommended for food preservation as it would be harmful to bacteria. The high content of total polyphenols in the different extracts shows that Safou has chemical constituents that could be responsible for many pharmacological actions. Indeed, polyphenols play an important role in human health by preventing degenerative diseases such as cancers, cardiovascular diseases, osteoporosis, dyslipidemia, anxiety or constipation. The results of this study corroborate with those of [12, 23] who showed high levels of total polyphenols in safflower in Malaysia and Nigeria respectively. The methanolic extracts of pulp and seed showed the highest values. The extraction solvents have an effect on the extraction yield and the content of bioactive compounds, thus affecting the biological activity of the extract. The results of this study could be due to higher solubility of the compounds in methanol than the other solvents tested (ethanol and distilled water). Methanol is an efficient extractor for polyphenols. It is a solvent widely used in laboratories and industries. The results of this study are similar to those of [21] who obtained the higher levels of total polyphenols in methanolic extracts.

The antioxidant activity in Safou extracts is influenced by the extraction method and the part of the fruit studied. Antioxidants are defined as substances present in foods that act on human physiological functions by significantly reducing the harmful effects of reactive

oxygen species, nitrogenous oxygen species, or both [25]. These results suggest that the extracts of Safou could be potential antioxidant agents to be incorporated in some drugs. Antimicrobial properties of substances are useful and necessary weapons for microbiologists in the fight or eradication of (pathogenic) microorganisms most importantly in food industry (food spoilage) and in the treatment of infectious diseases, as their active components can inhibit the growth and interfere with the metabolism of microorganisms in a negative manner [24-25]. The result of this study show antimicrobial methanolic and ethanolic extracts of Safou (*Dacryodes edulis*) against microorganisms tested. These antimicrobial activities demonstrated could be dependent on the presence of secondary metabolites (steroid/triterpenes, saponins, tannins, alkaloids, cardiac glycosides, reducing sugars, flavonoids and Phenols). Methanolic extract of *D. edulis* had the highest activity against microorganisms tested. This result corroborates with investigations made by [26, 27] using methanol as solvents for the extraction. A previous study has indicated great potential for the plant in the battle against malaria and other diseases [12, 28]

#### 4. CONCLUSION

This study highlighted the antioxidant and antimicrobial potential of the pulp and seed of Safou (*Dacryodes edulis*) grown in Côte d'Ivoire. The aqueous, ethanolic and methanolic extracts of the pulp and seed of Safou are rich in total polyphenols and showed high antioxidant activity. Only the methanolic and ethanolic extracts have an antibacterial activity on the different microorganisms tested. Safou (*Dacryodes edulis*) could of immense benefit to pharmaceutical industries for the development of new antimicrobial or chemotherapeutic drugs to address unmet therapeutic and also can be used as natural preservatives in food against the well-known causal agents of foodborne diseases and food spoilage.

#### COMPETING INTERESTS DISCLAIMER:

**AUTHORS HAVE DECLARED THAT NO COMPETING INTERESTS EXIST. THE PRODUCTS USED FOR THIS RESEARCH ARE COMMONLY AND PREDOMINANTLY USE PRODUCTS IN OUR AREA OF RESEARCH AND COUNTRY. THERE IS ABSOLUTELY NO CONFLICT OF INTEREST BETWEEN THE AUTHORS AND PRODUCERS OF THE PRODUCTS BECAUSE WE DO NOT INTEND TO USE THESE PRODUCTS AS AN AVENUE FOR ANY LITIGATION BUT FOR THE ADVANCEMENT OF KNOWLEDGE. ALSO, THE RESEARCH WAS NOT FUNDED BY THE PRODUCING COMPANY RATHER IT WAS FUNDED BY PERSONAL EFFORTS OF THE AUTHORS.**

#### REFERENCES

1. Riwom SH, NdoyéFoe FMCN, Nyegue MA, Wanki RA, Etoa FX. Chemical Composition and in Vitro Antibacterial Activity of the Essential Oils of the Leaves, Resin and Stem-Barks of *Dacryodes Edulis* (G. Don) H. J Lamgrowing in Cameroon on Diarrhea Associated Strains. *Journal of Applied Pharmaceutical Science*, 2015; 5 (10): 006-011.
2. Aubréville A. Flore du Gabon. Irvingiaceae, Simaroubaceae, Burseraceae. Vol. 3. Paris: Museum National d'Histoire Naturelle, 1962: 53-101.
3. Ajibesin KK, Essien EE, Adesanya SA. Antibacterial constituents of the leaves of *Dacryodes edulis*. *African Journal of Pharmacy and Pharmacology*, 2011;5 (15): 1782-1786

4. Ayuk E, Duguma B, Franzel S, Kangué J, Mollet M, Tiki-Manga T, Zekeng P. Uses, management and economic potentiel of *Dacryodes edulis* (Burseraceae) in the humid lowlands of Cameroon. *Economic Botany*, 1999 ; 53(3): 292-301.
5. Poligui R, Mouaragadja I, Haubruge E, Francis F. La culture du safoutier (*Dacryodes edulis* (G. Don) H.J. Lam (Burseraceae)) : enjeux et perspectives de valorisation au gabon (synthèse bibliographique). *Biotechnologie Agronomie société et Environnement*, 2013 ; 17(1) : 131-147.
6. Kinkela T, Kama-Niamayoua R., Mampouya D, Silou T. Variations in morphological characteristics, lipid content and chemical composition of safou (*Dacryodes edulis* (G. Don) H.J.LAM.) according to fruit distribution. A case study. *African journal of Biotechnology*, 2006 ; 5 (12): 1233-1238.
7. Nwokonkwo DC. The Phytochemical Study and Antibacterial Activities of the Seed Extract of *Dacryodes Edulis* (African Native Pear). *American Journal of Scientific and Industrial Research*, 2014; 5(1): 7-12.
8. Obame LC, Edou P, Bassolé IHN, Koudou J, Agnani H, Eba F, Traore AS. Chemical composition and antimicrobial properties of the essential oil of *Dacryodes edulis* (G. Don) H. J. Lam from Gabon. *African Journal of Microbiology Research*, 2008; 2: 148-152.
9. Oyetunji OO, Opeyemi AP. Antibacterial and Antioxidant Activities of *Dacryodes edulis* Methanolic Leaf Extract. *Journal of Advances in Medical and Pharmaceutical Sciences*, 2017; 14(1): 1-11
10. Bourdeaut. Le safoutier (*Pachylobus edulis*). *Fruits*, 1971; 26 (10) : 663-666
11. Sonwa D, Okafor J-C, MpungiBuyungu P, Weise S, Tchatat M, Adesina A, Nkongmeneck A, Ndoye O, Endamana D. *Dacryodes edulis*, a neglected non-timber forest species for the agroforestry systems of West and Central Africa, *Forests, Trees and Livelihoods*, 2002; 12 (1): 41-55.
12. Amise AF, Lennox JA, Agbo BE. Antimicrobial Potential and Phytochemical Analysis of *Dacryodes edulis* Against Selected Clinical Bacterial Isolates. *International Journal of Pharmacognosy and Phytochemical Research*, 2016; 8(11); 1795-1800.
13. Stanojević L, Mihajlo S, Vesna N, Ljubiša N, Dušica R, Jasna Č-B, Vesna T. Antioxidant Activity and Total Phenolic and Flavonoid Contents of *Hieracium pilosella* L. Extracts. *Sensors*, 2009; 9 (7) : 5702-5714
14. AOAC (Association of Official Analytical Chemists) (1990). *Official Methods of Analysis of the Association of Analytical Chemists*. 15th Edition. Washington, DC, USA, 684p.
15. AOAC (Association of Official Analytical Chemists) (2000). *Official Methods of Analysis of the Association of Analytical Chemists*, 17th Edition Washington, DC, USA.
16. Sanchez-Moreno C, Larrauri JA, Saura-calixto F. A procedure to measure the antiradical efficiency of polyphenols. *Journal Science Technology International*, 1998; 8: 121-137.
17. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols and other oxydant substrates and antioxidants by means of Folin-ciocalteu reagent. *Methods in Enzymology*, 1999; 299: 152-178.
18. Kirby W, Bauer AW. *Microbiology: disks diffusion sensitivity method*. University of Washington Medical School, Washington. 1960; 830-837.
19. Mordi RC, Olasehinde GI, Okedere AP, Elegwule AN, Ayo-Ajayi JI, Johnathan HO, Onibokun AE, Ajayi AA, Uchenna DO. Antibacterial activity of moderately volatile components of the oil extracted from the seeds of *Dacryodes edulis* G. LAM. *Asian Journal of Pharmaceutical and Clinical Research*, 2019; 12 (3): 246-249.
20. Quy DD, Artik EA, Phuong LT-N., Lien HH, Felycia ES, Suryadi I, Yi-Hsu J. Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatic*. *Journal of food and drug analysis*, 2014; 22: 296-302.
21. Truong Dieu-H, Dinh HN, Nhat TAT, Anh VB, Tuong HD, Hoang CN. Evaluation of the Use of Different Solvents for Phytochemical Constituents, Antioxidants, and In Vitro Anti-Inflammatory Activities of *Severiniabuxifolia*. *Journal of Food Quality*, 2019; 19: 9 p.



22. Soro S, Konan G, Elleingand E, N'guessan D, Koffi E. Formulation d'aliments infantiles à base de farines d'igname enrichies au soja. *African Journal of Food agriculture nutrition and development*, 2013; 3(5) :8313-8339.
23. Tee LH, Yang B, Prasad N, Ramanan N, Sun J, Seng CE, Ti TB, Azlan A, Ismail A, Lau CY, Jiang Y. Nutritional compositions and bioactivities of *Dacryodes species*: a review, *Food Chemistry*, 2014: 26p.
24. FNB (Food and Nutrition Board) Dietary Reference Intake: Elements. Institute of Medicine. National Academy of Sciences: Washington, D.C. 2001.
25. Kamba AS, Hassan LG. Phytochemical Screening and Antimicrobial Activities of African Star Apple (*Crysopyllum albidum*) Leaves, Stems Against Some Pathogenic Microorganisms, *International Journal of Pharmaceutical Frontier Research*, 2011; 1(2):119-129.
26. Nna PJ, Tor-Anyiin TA, IgoliJO, Khan ME, Anyam JV. Phytochemical and Antimicrobial extracts of *Dacryodes edulis*. *Screening of Root Biotechnology Journal International*, 2017; 19(3): 1-9.
27. Ikpa CCB, Maduka TOD. Antimicrobial Properties of Methanol Extract of *Dacryodes edulis* Seed and Determination of Phytochemical Composition Using FTIR and GCMS. *Chemistry Africa*, 2020: 9p.
28. Lennox JA, Agbo BE. Medicinal Properties of *Dacryodes edulis* against Selected Clinical Bacterial Isolates. *Microbiology Research Journal International*, 2017; 19(5): 1-7.