

Development and validation of ultra high performance liquid chromatographic (UHPLC) method for the determination of roxithromycin in the broiler plasma

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ABSTRACT

Aims: The present study was designed to develop and validate the UHPLC method for quantitative determination of roxithromycin, a macrolide antimicrobial drug, in broiler plasma for the application of pharmacokinetic studies.

Methodology: UHPLC apparatus comprised of ultraviolet (UV) detector was used in the present study. Chromatographic separation was performed by using reverse phase C18 column. Mobile phase was combination of buffer and 55 acetonitrile in the ratio of 55: 45. Buffer part used was 0.1 % trifluoroacetic acid (v/v) having pH of 2.1. Erythromycin was used as an internal standard. Isocratic elution mode was employed with flow rate of 1 ml/min and effluents were monitored at wavelength of 220 nm. Liquid-liquid extraction using ice-cold acetonitrile was performed to extract roxithromycin from plasma samples. The data integration was performed using Chromeleon™ version 6.8 software.

Results: The linear calibration curve with a mean correlation coefficient (R^2) value of 0.9999 was observed for concentrations ranging from 0.20 to 12.80 µg/ml. At any concentration, accuracy was not found to be less than 90 %. The mean extraction recovery (n=5) for concentrations of 0.40 µg/ml was 81.36 %. The calculated intraday and interday C.V. % was not more than 7.70 % and 9.42 %, respectively, at any concentration studied. The specificity of the analysis was reflected by the narrow range of retention time ranging between 6.983 to 7.178 minutes. LOD and LOQ of the method under investigation were calculated as 0.131 and 0.398 µg/ml, respectively

Conclusion: A reliable, reproducible, accurate, precise, specific and sensitive method for analysis of roxithromycin in broiler plasma was developed and validated for application in the pharmacokinetic study of the roxithromycin.

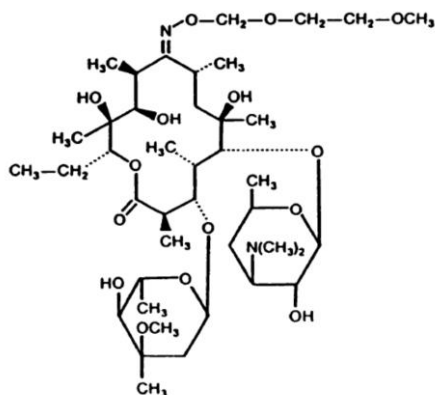
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Keywords: UHPLC, Roxithromycin, Method validation, Broiler plasma

1. INTRODUCTION

Roxithromycin is a semi-synthetic, long-acting, orally administered antibacterial drug of macrolide class [1]. One of the biggest advantages of roxithromycin is its power of intracellular concentration. The pharmacokinetic profile of roxithromycin is characterized by high plasma, tissue and body fluid concentrations and a long half-life permitting an extended dosage interval [2]. Macrolide antibiotics and their semi-synthetic derivatives like roxithromycin are most commonly used in poultry to treat avian mycoplasmosis and are considered to be bacteriostatic at low therapeutic concentrations, but can be slowly bactericidal at higher concentrations [3]. Roxithromycin is widely used as antimicrobial drug for treating many bacterial infections especially *Mycoplasma* spp. in broiler chickens. In Asian countries, its use is more in poultry industry for the treatment of Chronic Respiratory

26 Disease (CRD) caused by avian pathogen *Mycoplasma gallisepticum* [4]. The chemical
27 structure of roxithromycin is presented as figure 1.
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31 Fig. 1. Chemical structure of roxithromycin (9E-[O-((2-methoxyethoxy)-methyl)oxime]-
32 erythromycin).

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34 Simple HPLC - UV (using ultra-violet detector) methods are reported for the roxithromycin
35 detection from tablets or pharmaceutical formulations and used for stability studies [5, 6] but
36 these methods are not suitable for analysis from biological matrices like plasma. A number
37 of advanced methods are reported for analysis of roxithromycin in biological matrices like
38 rat plasma [7], rat lung tissues [8], human serum [9], human urine [10] and human plasma [11,
39 12]. But all these methods involved advanced and costly detection techniques using
40 electrochemical, amperometric, photo-diode array and fluorescence (with derivitization)
41 detectors. One report of roxithromycin detection using HPLC - UV method was reported, that
42 too for human plasma [13]. For broiler plasma, no HPLC - UV method is reported, however
43 costly advanced LC-MS technique is reported for detection of roxithromycin from broiler
44 tissues implicated mainly for residue studies [14]. Thus, the present study was undertaken to
45 develop and validate a simple UHPLC-UV method for determination of roxithromycin in
46 broiler chicken's plasma for studying pharmacokinetic studies.

47 48 2. MATERIAL AND METHODS

49 50 2.1 Drugs and Chemicals

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52 Certified E.P. (European Pharmacopoeia) grade of roxithromycin and erythromycin (for
53 internal standard) pure drug powders were procured from Sigma-Aldrich Chemicals Pvt. Ltd.,
54 Bengaluru, India. Water, acetonitrile, trifluoroacetic acid, sodium hydroxide and other
55 reagents of HPLC grade were procured from S. D. Fine-Chem Limited, Mumbai, India.
56 Triethylamine of HPLC grade was procured from Merck, Mumbai, India.

57 58 2.2 Broiler Plasma

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60 The blank drug free plasma from broiler chickens was supplied by the Department of
61 Livestock Production and Management, College of Veterinary Science and Animal
62 Husbandry, SDAU, Sardarkrushinagar, Gujarat, India.

63 64 2.3 Instruments

65

66 Ultra high performance liquid chromatography (UHPLC) apparatus (Dionex ultimate 3000[®],
67 Thermo Fisher, Germany) comprised of ultraviolet (UV) detector and gradient solvent
68 delivery pump was used in the present study. Chromatographic separation was performed
69 by using reverse phase C₁₈ column (ODS, 25 cm x 4.6 mm ID, 4.5 μ; Purospher[®] Star RP-
70 18, Merck-Millipore, Mumbai, India) at ambient temperature (~28°C). The data integration
71 was performed by software 'Chromeleon™ version 6.8' Chromatography Data System.
72

73 2.4 Drug Extraction from Plasma Standards

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75 Drug-free broiler plasma was used for making different roxithromycin concentrations of 0.20,
76 0.40, 0.80, 1.60, 3.20, 6.40, and 12.80 mg/ml. After drug extraction, each of these standards
77 was run in quintuple (n=5) to get area response in UHPLC. Liquid-liquid extraction using ice-
78 cold acetonitrile was performed to extract roxithromycin from plasma samples. Exactly 400
79 μL of plasma sample was taken into 2 ml Eppendorf[®] micro-centrifuge tube, and then 40 μL
80 1 M NaOH was added to it and vortexed for about 10 seconds. After this alkalization, 1200
81 μL ice-cold acetonitrile was added in same tube and vortex mixed for 3 minutes at 2400
82 RPM. Then the mixture was centrifuged at 5000 RPM for 10 minutes at 4°C. Upper organic
83 phase was collected and dried under nitrogen (N₂) evaporator (AT-EV-50, Athena
84 Technology, Mumbai, India). Exactly, 20 μL of internal standard (200 mg/ml erythromycin
85 dissolved in diluent) was mixed with 80 μL diluent (made up of equal parts of ice-cold
86 acetonitrile and HPLC grade water) for reconstitution of each extracted and evaporated
87 sample. Thus, dried residues were reconstituted with total 100 μL diluent. The prepared
88 sample was finally centrifuged (5000 RPM, 5 minutes, 4°C) and 20 μL of the upper clear
89 portion was manually injected into UHPLC.
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91 2.5 Optimized Chromatographic Conditions

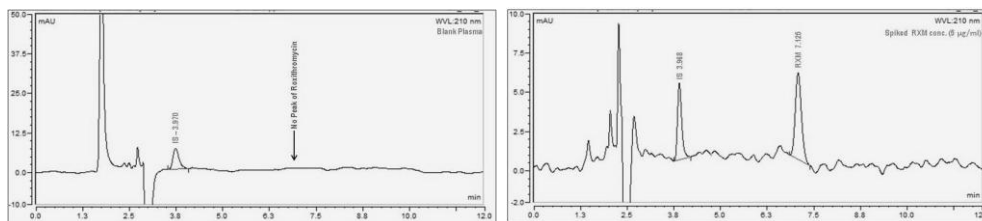
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93 The optimized chromatographic parameters for UHPLC analysis of roxithromycin
94 concentrations in broiler plasma sample are given in Table 1. Mobile phase comprises of 55
95 parts of buffer with 45 parts of acetonitrile. Buffer part was prepared by mixing trifluoroacetic
96 acid (TFA) with HPLC grade water to yield strength of 0.1 % TFA buffer (v/v) having pH of
97 2.1. It was filtered by filter paper (0.45 μm pore size) by using a vacuum pump and
98 degassed using ultrasonic sonicator before use. During sample run, intermittent washings of
99 microsyringe were done with washing solution (80 Acetonitrile: 20 HPLC grade water) to
100 avoid carry over effect.
101

102 **Table 1. Optimized chromatographic parameters for UHPLC analysis of roxithromycin**

S.N.	UHPLC parameters	Optimized values
1	Mobile phase components and ratio	55 (TFA Buffer) : 45 (Acetonitrile)
2	Elution mode	Isocratic
3	Flow rate	1.0 ml/min
4	Detection wavelength (λ_{max})	210 nm
5	Injection volume	20 μL
6	Run time	12 min
7	Retention time (RT)	7.1 min

103 The representative chromatograms are shown in Figure 2.
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106

107 Fig. 2. Chromatograms of (A) blank plasma, (B) plasma spiked with roxithromycin (5 µg/ml)

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109 UV detector responses were generated as chromatograms using 'Chromeleon™ version 6.8'

110 software at the data collection rate of 2.5 Hz and a time constant of 0.6 second.

111

112 2.6 Method Validation

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114 For validation of the present developed UHPLC method for analysis of roxithromycin, major

115 parameters like linearity, recovery, accuracy and precision (intraday and inter-day) were

116 calculated as per standard guidelines [15, 16]. Limit of detection (LOD) and Limit of

117 quantification (LOQ) were calculated using the standard deviation of responses (Std. Dev.)

118 and the slope value of the calibration curve [17]. Formulae used were $LOD = 3.3 \times (\text{Std. Dev.} / \text{Slope})$

119 and, $LOQ = 10 \times (\text{Std. Dev.} / \text{Slope})$.

120

121 2.7 Statistical Analysis

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123 All values are expressed as the mean \pm standard deviation. Precision C.V. (Co-efficient of

124 variance) was calculated as $\% = (\text{Standard Deviation} / \text{Mean of observed concentration}) \times$

125 100.

126 3. RESULTS AND DISCUSSION

127

128 3.1 Optimization of chromatographic parameters

129 The ultimate goal of optimization of any HPLC method is to achieve good resolution peaks

130 with acceptable retention times but without peak tailing, the absence of interfering

131 background peaks, and high and stable sensitivity [19]. Many buffers have been used in past

132 for reverse phased chromatographic estimation of roxithromycin mainly phosphate and

133 acetate buffers [7, 11, 13, 18], but in present study, trifluoroacetic acid (TFA) buffer gives

134 good chromatographic results in terms of better separation of peaks in plasma samples. It

135 has added advantage for having longer working life of silica columns. The precaution was

136 taken to prepare fresh 0.1 % TFA (v/v) each day prior to analysis due to its volatile nature.

137 HPLC grade Acetonitrile (ACN) was used as organic modifier part in mobile phase as it was

138 reported to having good solute retention capacity for roxithromycin [19]. Moreover, ACN is

139 good choice when detection is to be done at lower wavelengths of 200-220 nm as it has

140 lower UV cut-off value of 190 nm. Generally, macrolides are known to show higher

141 absorbance at lower wavelengths and produce weak chromatographic signals. UV-detection

142 of roxithromycin has been performed at λ_{max} value between 200 to 220 nm [5,

143 6, 10, 13]. The three wavelengths i.e. 205, 210 and 215 were tried in the present study and

144 larger peak areas were obtained at λ_{max} of 210 nm; hence this wavelength was selected for

145 chromatographic detection of roxithromycin.

146

147 3.2 Method validation

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149 **3.2.1 Linearity**

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151 The linearity was tested from the calibration curve prepared from spiked plasma samples for
152 concentrations in the range from 0.20 to 12.80 µg/ml. Thus, the method was validated over a
153 dynamic linear range of 0.20 – 12.80 µg/ml for roxithromycin with a mean correlation
154 coefficient (R^2) value of 0.9999.

155

156 **3.2.2 Recovery**

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158 Various liquid – liquid extraction (LLE) protocols with alkalization had been used to extract
159 roxithromycin from biological matrices using solvents like tert-butyl methyl ether [7], Diethyl
160 ether with isopentane [11], hexane plus isoamyl alcohol [13], dichloromethane [10] and
161 Acetonitrile [9, 12]. In present study, extraction was tried with dichloromethane and
162 acetonitrile and more recovery was observed with acetonitrile. The extraction recovery of
163 roxithromycin was measured by comparison of the areas of roxithromycin after injection of
164 the extracted spiked samples (n=5) with those obtained after injection of the standard
165 solution containing equivalent concentrations of the drug. The extraction recovery was
166 studied at three different concentrations of 0.40 µg/ml (low near to LOQ), 3.20 µg/ml
167 (medium) and 12.80 µg/ml (high concentration). The respective mean recoveries were
168 obtained as 81.36, 79.83 and 81.12 % (Table 2).

169

170 **Table 2: Mean extraction recoveries for roxithromycin from spiked plasma (n=5)**

171

S.N.	Spiked Plasma Concentration (µg/ml)	Recovery (%)	Range (%)	Mean Recovery (%)
1	0.400	75.07		
2	0.400	86.73		
3	0.400	72.71	72.71 – 94.91	81.36
4	0.400	90.93		
5	0.400	94.91		
6	3.200	70.69		
7	3.200	85.96		
8	3.200	71.46	70.69 – 92.63	79.83
9	3.200	91.23		
10	3.200	92.63		
11	12.800	72.98		
12	12.800	86.18		
13	12.800	70.80	70.80 – 94.50	81.12
14	12.800	94.50		
15	12.800	91.58		

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173 **3.2.3 Accuracy**

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175 Accuracy (in per cent) was estimated by deviation of observed concentration in plasma
176 sample with spiked known drug concentration in the plasma sample. Accuracy was studied
177 at three different concentrations in triplicates. At any concentration, accuracy was not found
178 to be less than 90% (Table 3).

179

180 **Table 3. Accuracy of the developed method for analysis of roxithromycin from broiler
181 plasma (n=3)**

S.N.	Spiked Plasma Concentration ($\mu\text{g/ml}$)	Observed Concentration ($\mu\text{g/ml}$)	Accuracy (%)
1	0.400	0.421	97.90
2	0.400	0.446	95.40
3	0.400	0.471	92.90
4	3.200	3.127	92.70
5	3.200	3.298	90.20
6	3.200	3.116	91.60
7	12.800	12.764	96.40
8	12.800	12.889	91.10
9	12.800	12.825	97.50

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3.2.4 Precision

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Intraday and inter-day precisions (n=5) were expressed in the terms of C.V. % (Co-efficient of variance). The highest intraday and interday C.V. % calculated was 7.70 % (at 0.20 $\mu\text{g/ml}$) and 9.42 % (at 0.40 $\mu\text{g/ml}$), respectively (Table 4). Thus, it was acceptable since CV% was within the acceptable 20% limit at the lower limit of quantification (0.400 $\mu\text{g/ml}$) and within 15% for other concentrations studied.

Table 4: Intraday and inter-day precision of roxithromycin in broiler plasma (n=5) by UHPLC - UV detection

S.N.	Spiked Concentration ($\mu\text{g/ml}$)	Mean Observed Area (mAU*min)	Standard Deviation	Precision C.V. (%)
Intraday values (n=5)				
1	0.200	0.0437	0.0034	7.77
2	0.400	0.1076	0.0046	4.27
3	0.800	0.1996	0.0078	3.91
4	1.600	0.3732	0.0155	4.15
5	3.200	0.7479	0.0242	3.24
6	6.400	1.4912	0.0404	2.71
7	12.800	3.0070	0.1441	4.79
Interday values (n=5)				
1	0.200	0.0497	0.0033	6.65
2	0.400	0.1147	0.0108	9.42
3	0.800	0.2152	0.0167	7.76
4	1.600	0.3918	0.0329	8.40
5	3.200	0.8457	0.0645	7.63
6	6.400	1.6702	0.1065	6.38
7	12.800	3.3259	0.2463	7.41

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3.2.5 Specificity

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This parameter is concerned with the extent to which other substances interfere with the identification and, where appropriate, quantification, of the analyte(s) of interest. The retention times of roxithromycin were specifically ranged, without any interference peaks, from 6.983 to 7.178 (~ 7.1 \pm 0.1) minutes with a mean of 7.081 minutes.

201 **3.2.6 Sensitivity**

202

203 Limit of detection (LOD) and Limit of quantification (LOQ) were calculated using the standard
204 deviation of responses and the slope value of the calibration curve. LOD and LOQ of the
205 developed analytical method for roxithromycin were calculated as 0.131 and 0.398 µg/ml,
206 respectively. The target quantification limit of roxithromycin in plasma for pharmacokinetic
207 study should be above its MIC (minimum inhibitory concentration), that is 0.5 µg/ml against
208 most of common susceptible pathogens. The MIC cut-off point for roxithromycin separating
209 sensitive from resistant human pathogens was suggested to be 0.5 µg/ml [20] whereas, MIC
210 breakpoints for roxithromycin against avian pathogen *Mycoplasma gallisepticum* were
211 suggested as ≤ 1–4 (Sensitive), ≤ 4 (Intermediate) and > 4 (Resistant) µg/ml [21].

212

213 **3.2.7 Internal Standard**

214

215 Internal standard calibration involves the comparison of the instrument responses from the
216 target compound in the sample to the responses of reference standard added to the sample
217 or sample extract before injection [11, 22]. The difference between retention times of the
218 target compound and the internal standard may be used to know the relative retention time
219 which can be further used to compensate for small retention time shifts. Roxithromycin and
220 erythromycin were used as internal standard for quantification of each other [18]. In present
221 analytical method for roxithromycin, erythromycin (200 µg/ml concentration in diluent) was
222 found appropriate as an internal standard. Retention time of erythromycin was 4.0 ± 0.1
223 minutes in the present method.

224

225 **4. CONCLUSION**

226

227 The developed UHPLC method for determination of roxithromycin in broiler plasma was
228 found to be reliable, reproducible, accurate, precise, specific and sensitive method, and
229 serves the purpose for the application in the pharmacokinetic study with limit of quantification
230 equal to 0.398 µg/ml which is lower than the target MIC value of roxithromycin against
231 susceptible bacterial pathogens in broiler chickens which ranges from 0.500 to 1.000 µg/ml.

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234 **COMPETING INTERESTS**

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236 There is no competing interests exist.

237

238 **CONSENT**

239

240 It is not applicable.

241

242 **ETHICAL APPROVAL**

243

244 It is not applicable.

245

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