

**EFFECTS OF THE ETHANOL EXTRACTS OF THE ROOT, STEM AND LEAVES OF  
BALANITES AEGYPTIACA AGAINST SELECTED MICROBES**

**ABSTRACT**

*Balanites aegyptiaca* has been used in many traditional treatments of microbial infections. This work was carried out with the aim of determining the phytochemical compounds present in the ethanol extracts of the root, stem and leaves of *Balanites aegyptiaca* and their effects against selected microorganisms. Phytochemical screening indicated the presence of; Alkaloids, Tannins, flavonoids, Saponins, Steroid, and anthraquinones, although anthraquinones were absent in ethanol extracts of root and stem bark of *Balanites aegyptiaca* but present in the ethanol extract of the leaf. Antibacterial effects were tested against *Staphylococcus aureus* and *Klebsiella pneumoniae*. Results obtained from the ethanol extract of the root stem and leaves showed significant zones of inhibition against *Klebsiella pneumoniae* and *Staphylococcus aureus* at all concentrations of 100mg/ml, 50mg/ml, 25mg/ml, 12.5mg/ml and 6.125mg/ml. The positive control performed better than the extracts in terms of zone of inhibition. The results of the minimum inhibitory concentration (MIC) indicated that the leaf ethanol extract had the greatest activity against *K. pneumoniae* with MIC value of 12.5mg/ml, while the results from the minimum bactericidal concentration (MBC) showed that the leaf ethanol extract had the greatest activity against *K. pneumoniae* with an MBC value of 12.5mg/ml.

**Key words:** *Balanites aegyptiaca*, phytochemical compounds, microorganisms, plant extracts

**INTRODUCTION**

The antimicrobial activities of plant oils and extracts have formed the basis of many applications, including raw and processed food preservation, pharmaceuticals, alternative medicine and natural therapies. Moreover, the increasing use of plant extracts in the food, cosmetics and pharmaceutical industries suggests that in order to find active compounds, a systematic study of medicinal plants is very important [1]. Traditional medicine has remained the most affordable and easily accessible source of treatment in the primary health care system of resource poor communities and local therapy is the only means of medical treatment for such communities [2]. According to [3] about 80% of Nigerian homes, maintain some sort of private family traditional medicine practitioner. Existing data and contemporary researchers seem to authenticate the assumption for general health improvement of the masses by traditional healers. Plants have broader uses than just food and genetic reservoirs. Medicinal plants

34 have been used for centuries to treat a wide variety of ailments [4]. The presence of secondary  
35 metabolites in plants has been associated in most of their therapeutic activities [5]. Herbal medicines are  
36 now considered a part of Complementary and Alternative medicine (CAM) and are gaining popularity due  
37 to their potent antioxidant activity, minimal side effects and economic viability [6]. Many research efforts  
38 have been directed towards the provision of empirical proofs to back up the use of plants species in trade  
39 and medicinal practices in recent years [7].

## 40 **MATERIALS AND METHODS**

### 41 **Source and preparation of plant materials**

42 The plant materials were collected from Gombe State University botanical garden Biological science  
43 department; Gombe State University, it located in Gombe between the latitude 10°00N to 10°20N and  
44 longitude 11°0E to 11° E. These were brought and identified by a taxonomist with voucher number  
45 900191 at the Herbarium unit of the Department of Biological Sciences, Gombe State University. The  
46 plant parts were air-dried for two weeks at room temperature (25°C) in the laboratory and then ground to  
47 powder.

### 48 **Extraction procedures**

49 The ground plant parts were extracted at the Department of Pharmacognosy, Faculty of  
50 Pharmaceutical Sciences, Gombe State University, following the methods of [8].

### 51 **Preparation of Ethanol Extraction of *Balanites aegyptiaca***

52 Approximately 800 g of the dried leaves and roots of *B. aegyptiaca* were extracted with 10 litres of 80%  
53 (v/v) ethanol by maceration at (25°C) for 3days. The total mixture then is strained and filtered. The filtrate  
54 was concentrated to dryness on a water bath at 100° C so as to obtain the dry extract after which was  
55 stored at -20°C for further studies.

56

57

## 58 **Phytochemical analysis**

59 The method of [8] was employed for the test of the presence of the phytochemical properties.

## 60 **Source and preparation of test microorganisms**

61 The stock cultures of the test microorganisms were obtained from the Department of Microbiology,  
62 Gombe State University. Their validity was determined by sub culturing onto nutrient agar and confirmed  
63 by standard cultural, morphological and biochemical techniques as described by [9]. The inocula of the  
64 test organisms were standardized by the method of [10]. This was done by suspending each test  
65 organism in 5ml of nutrient broth and the turbidity was compared with that of 0.5 McFarland standard.  
66 McFarland standard was prepared by adding 0.6ml of 1% barium chloride (BaCl<sub>2</sub>) to 99.4ml of 1%  
67 sulphuric acid (H<sub>2</sub>SO<sub>4</sub>) solution. The turbidity of the 0.5 McFarland standards was used for estimation of  
68 the number of bacteria in broth culture (culture for 24 hours at 37<sup>0</sup>C) to pour into 5ml of distilled water in  
69 order to obtain a standard bacterial suspension of 1 x 10<sup>5</sup> cfu/ml [11].

## 70 **Preparation of concentration of extracts**

71 Approximately 1g of each extract was dissolved in 5mls of distilled water to yield 200mg/ml. 1ml of the  
72 200mg/ml was taken and added to 1ml of distilled water to give a concentration of 100mg/ml. 1ml of the  
73 100mg/ml extract concentration was also taken and added to 1ml of distilled water to get a concentration  
74 of 50mg/ml. The procedure was repeated twice to give concentrations of 25mg/ml and 12.5mg/ml.

## 75 **Antibacterial susceptibility testing**

76 The antibacterial activity of the fractions of *B. aegyptiaca* was determined using the well method (Kirby-  
77 Bauer Methods) as described by [12]. Standard aseptic Microbiological methods were followed  
78 throughout this antibacterial study.

79

## 80 **Well method for antibacterial activity**

81 The well method was employed to assay the plant fractions for antibacterial activity. Petri dishes were  
82 poured with nutrient agar and allowed for 30 minutes to solidify (This was done in duplicate for each

83 fraction and test organism). The test organisms were then inoculated by spreading on the inocula on the  
84 surface of the medium using a sterile swab stick. A sterile Cork borer (size 3) was used to bore 4 wells in  
85 the medium. The different concentration of the plant fractions were placed in the wells using a sterile  
86 syringe and needle (different for each sample and test organism). These were then allowed a diffusion  
87 time of 1 hour after which the plates were incubated at 37 °C for 24 hours. The positive control was  
88 Ceftriaxone (100mg/ml). The potency of the extracts was determined by the clear zones of inhibition  
89 around the wells and was respectively measured as the diameter zones of inhibition. MIC was determined  
90 using the method of [13], while MBC was determined using the method of [14].

### 91 **Data analysis**

92 One way Analysis of Variance (ANOVA) of was used to assess the efficacy of the plant parts in terms  
93 of the activity as was shown by the zones of inhibition. Since there was no significant difference,  
94 Duncan's multiple range test (DMRT) and Student t-test was not carried.

### 95 **Result and Discussion**

96 The result of preliminary phytochemical screening (table 1) revealed the presence of alkaloids,  
97 flavonoids, tannins, saponins and steroids in the roots and stem bark, while anthraquinones were present  
98 in the leaf only. This is similar to that of [15] who investigated the activity of root, stem-bark and leaves  
99 extracts of *Terminalia glaucescens*' against some pathogenic organisms. Phytochemical screening of the  
100 fractions of *Terminalia glaucescens*' also revealed the presence of alkaloids, tannins, saponins, steroids,  
101 flavonoids, anthraquinones and phlobatannins (mostly in root and stem-bark). Also, [16] in the screening  
102 of the methanolic and water extracts of the stem bark of *Jatropha curcas* revealed the presence of  
103 saponin, steroids, tannin, glycosides, alkaloids and flavonoids in the extracts. [17] Found tannins and  
104 anthraquinones (the largest group of quinones) to possess antibacterial effects by inhibiting nucleic acid  
105 synthesis. Anthraquinones were absent in the root and stem-bark extracts. This is similar to the work of  
106 [18] who investigated the ethanol extract of *Maranta arundinacea* rhizomes and found alkaloids to be  
107 present in the ethanol extract, but absent in the ethyl-acetate fraction. The presence of these  
108 phytochemicals has been reported to account for the exertion of antimicrobial activity by plants [19].

109 The result of antibacterial activity (Table 2, 3 and 4) of the ethanol extracts of root, stem bark and leaf of  
 110 *Balanites aegyptiaca* revealed the potentiality of the plant in treating and curing diseases cause by the  
 111 tested bacteria and other microorganisms. The ethanol extract of the root, stem bark, and leaves has  
 112 inhibited the growth of *Klebsiella pneumoniae* and *Staphylococcus aureus*, with MIC values with low  
 113 concentration of 25mg/ml for *S. aureus* and 12mg/ml for *K. pneumoniae* respectively.

114  
 115 It was shown from previous studies that the root extract of the plant has more and lethal effect on  
 116 microorganisms, followed by the bark among the various part tested [20]. But based on these work and  
 117 the statistical analysis, the leaf extract showed MIC value with low concentration of 12.5mg/ml for *S.*  
 118 *aureus* sand 50mg/ml for *K. Pneumoniae*, than the root and stem bark extract on both the tested bacteria  
 119 (Table 5). The root showed MIC value with concentration of 50mg/ml for both the tested organisms. The  
 120 result of the MIC and MBC also showed that *Balanites aegyptiaca* is bactericidal (Table 6).

121

122 **Table 1:** Qualitative phytochemical screening of *Balanites aegyptiaca*

	Root	Stem Bark	Leaf
	Ethanol	Ethanol	Ethanol
Alkaloids	+	+	+
Tannins	+	+	+
Flavonoids	+	+	+
Saponins	+	+	+
Steroids	+	+	+
Anthraquinones	-	-	+

123 + = Present, - = Absent.

124

125

126 **Table2:** The mean of the sensitivity test of the microorganisms to the ethanol root extract of *Balanites*  
 127 *aegyptiaca*

Microorganisms	Zones of inhibition (mm)					
	100mg/ml	50mg/ml	25mg/ml	12.5mg/ml	6.125mg/ml	control
<i>K. pneumoniae</i> <sup>13</sup>	10	5	2	2	26	
<i>S. aureus</i>	16	13	9	7	4	24
Control: Ceftriaxone						

135 **Table 3:** The mean of the sensitivity test of the microorganisms to the ethanol stem bark extract of  
 136 *Balanites aegyptiaca*

Microorganisms	Zones of inhibition (mm)					
	100mg/ml	50mg/ml	25mg/ml	12.5mg/ml	6.125mg/ml	control
<i>K. pneumoniae</i>	14	11	9	5	3	24
<i>S. aureus</i>	8	12	9	7	5	25
Control: Ceftriaxone						

143 **Table 4:** The mean of the sensitivity test of the microorganisms to the ethanol leaf extract of *Balanites*  
 144 *aegyptiaca*

Microorganisms	Zones of inhibition (mm)					
	100mg/ml	50mg/ml	25mg/ml	12.5mg/ml	6.125mg/ml	control
<i>K.pneumoniae</i>	16	12	9	7	5	26
<i>S. aureus</i>	20	16	13	10	6	24
Control: Ceftriaxone						

151 **Table 5:** Minimum inhibitory concentration (MIC) for microorganism of different extracts of *Balanites*  
 152 *aegyptiaca* in mg/ml

Microorganisms	Root	Stem bark	Leaf
<i>K. pneumoniae</i>	25	25	12.5

155 *S. aureus* 50 25 25

156

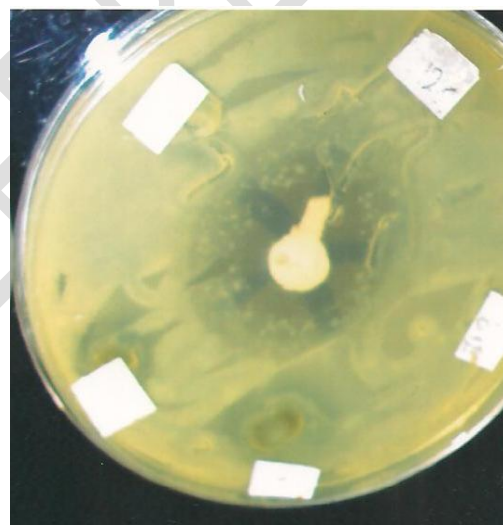
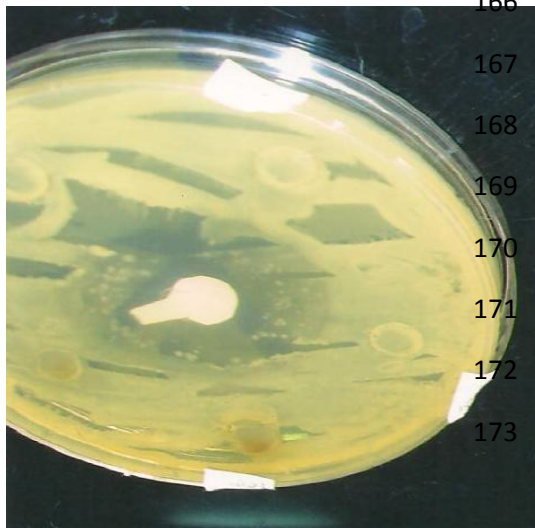
157 **Table 6:** Minimum Bactericidal Concentration (MBC) for microorganism of different extracts of *Balanites*  
158 *aegyptiaca* in mg/ml

159 Microorganisms	Root	Stem bark	Leaf
160 <i>K. pneumoniae</i>	50	25	12.5
161 <i>S. aureus</i>	50	25	50

162

163

164 **Plate 1:** Inhibition zone of the root extract at different concentrations against *Klebsiella pneumoniae* and  
165 *Staphylococcus aureus*



174 *Klebsiella pneumoniae*

*Staphylococcus aureus*

175

176

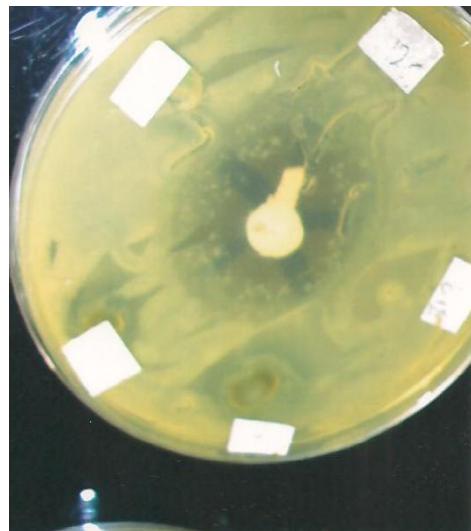
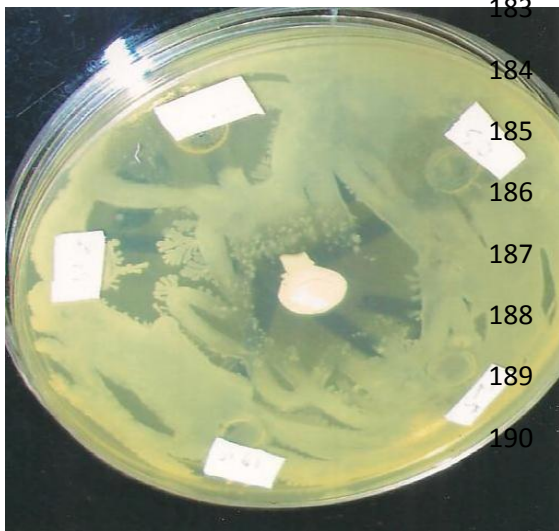
177

178

179

180

181 **Plate 2:** inhibition zone of the stem bark extract at different concentrations against *Klebsiella pneumoniae*  
182 and *Staphylococcus aureus*

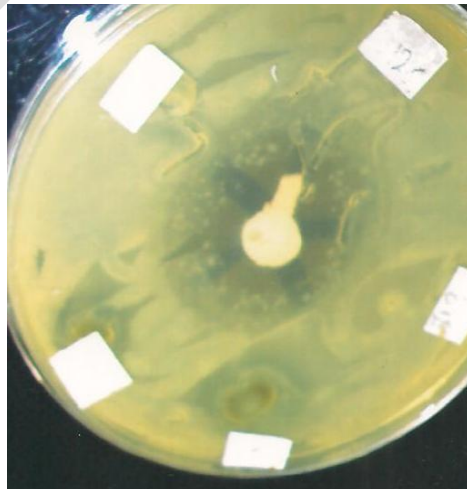
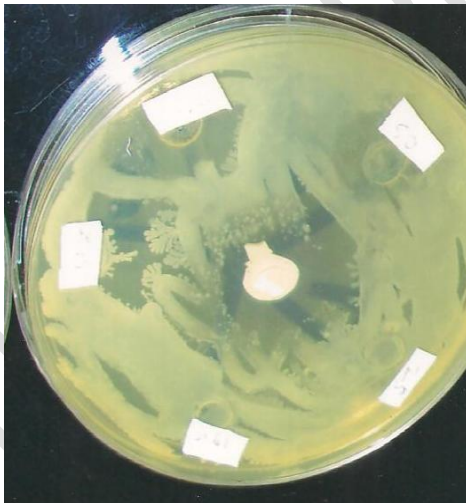


191 *Klebsiella pneumoniae*

*Staphylococcus aureus*

192

193 **Plate 3:** Inhibition zone of the leaf extract at different concentrations against *Klebsiella pneumoniae* and  
194 *Staphylococcus aureus*



204 *Klebsiella pneumoniae*

*Staphylococcus aureus*

205

206

207



## 208 **Conclusion**

209 *Balanites aegyptiaca* is traditionally used in treatment of various ailments. The phytochemical screening  
210 of the ethanol extract of root, stem bark, and leaves revealed the presence of; alkaloids, flavonoids,  
211 saponins, tannins, and steroids, in both and anthraquinones in the leaf only.

212  
213 The result of the antibacterial activity of the ethanol extract of the root, bark and leaf of *Balanites*  
214 *aegyptiaca* inhibited the growth of *Klebsiella pneumoniae* and *Staphylococcus aureus* with MIC value of  
215 12.5mg/ml for *K. pneumoniae* and 25mg/ml for *S. aureus*.

216  
217 The demonstration of antibacterial activity of the *Balanites aegyptiaca* is indeed a development that will  
218 help to discover new antibiotics that can serve for treatment of infections that are caused by bacteria that  
219 are becoming resistant to most of the antibiotics used for treatment of diseases caused by the  
220 microorganisms. The fact that plants are very common makes it a cheaper alternative for drugs  
221 development for human consumption and used.

## 223 **References**

- 224 1. Misrak, K., Amare, A. and Mohammed, Y. (2013). Efficacy of plant extracts, traditional materials and  
225 antibacterial chemicals against *Xanthomonas campestris* pv. *vesicatoria* on tomato seed. *African*  
226 *Journal of Microbiology Research*, 7(20): 2395-2400  
227
- 228 2. Yinger, H. and Yewhalaw, D. (2007). Traditional medicinal plant knowledge and use by local healers in  
229 Sekoru district, Jimmazon, Southwestern Ethiopia. *Journal of Ethnobiology and Ethnomedicine*. 3: 24-  
230 30
- 231 3. Alaribe, S.I. (2008). A Survey of the Importance and Problems of Traditional Health care medicine, A  
232 case study of Ezinihitte Mbaise L.G.A. Imo State. Unpublished B.Sc. project, A.I.F.C.E. Owerri, Imo  
233 State.  
234
- 235 4. Vaidya, A.D. and Devasagayam, T.P. (2007). Current status of herbal drugs in India: an overview.  
236 *Journal of Clinical Biochemical Nutrition*, 41:1-11.  
237
- 238 5. Ogunleye, D.S. and Ibitoye, S.F. (2003). Studies of antimicrobial activity and chemical constituents of  
239 *Ximema Americana*. *Tropical Journal of Pharmacology* 2:239-241.  
240  
241
- 242 6. Auddy, B., Ferreira, M., Blasina, F., Lafon, L., Arredondo, F., Dajas, F., Tripathi, P.C., Seal, T. and  
243 Mukherjee, B. (2003). Screening of antioxidant activity of three Indian medicinal plants, traditionally

- 244 used for the management of neurodegenerative diseases. *Journal of Ethno-Pharmacology*, 84:131-  
245 138.  
246
- 247 7. Parker, J. D. and Burkepile, D. E. (2017). Recent advances in plant-herbivore interactions. *Public*  
248 *Medicine*, 1(6): 1-3
- 249 8. Sofowora, A. (2006). *Medical Plants and Traditional Medicine in Africa*. (2<sup>nd</sup> edn). Spectrum books Ltd,  
250 Ibadan. Nigeria. PP. 150 –153.  
251
- 252 9. Cowan, S. T., and Steel, K. J. (2004). *Manual for the identification of medical bacteria*. Cambridge  
253 University Press, New York. PP. 45 – 63.  
254
- 255 10. Barry, A. L. and Thornsberry, C. (1991). Susceptibility tests: diffusion test procedures. In Balows, A.  
256 (ed.): *Manual of Clinical Microbiology*, 40: 213-219.  
257
- 258 11. Bauer, A.W., Kirby, M.D.K., Sherras, J.C. and Trick, M. (2003). Antibiotic susceptibility testing by  
259 standard single disc diffusion method. *American Journal of clinical Pathology* 45:490-496.  
260
- 261 12. Abalaka, M.E., Mann, A. and Adeyemo, S.O. (2011). Studies on in-vitro antioxidant and free radical  
262 scavenging potential and phytochemical screening of leaves of *Ziziphus mauritiana* L and *Ziziphus*  
263 *spinachristi* L compared with Ascorbic acid. *Journal of Medical Genetics and Genomics*, 3(2):28-34.  
264  
265
- 266 13. Doughari, J. H., Pukuma, M. S. and De, N. (2007) Antibacterial effects of *Balanites aegyptiaca* L. Drel.  
267 and *Moringa oleifera* Lam. on *Salmonella* Typhi. *African Journal of Biotechnology*. 6 (19), pp. 2212-  
268 2215.
- 269 14. Rotimi, V. O., Laughon, B. E., Bartlett, J. G. and Mosadami, H. A. (1988). Activities of Nigerian chewing  
270 stick extracts against *Bacterioides melaninogenius*. *Antimicrobial Agents of Chemotherapy*, 32: 598-  
271 600.  
272
- 273 15. Adebayo, E. A. and Ishola, O. R. (2009). Phytochemical and antimicrobial screening of crude extracts  
274 from the root, stem bark and leaves of *Terminalia glaucescens*. *African Journal of Pharmacy and*  
275 *Pharmacology* 3 (5): 217-222.  
276  
277
- 278 16. Igbinosa, O. O., Igbinosa, E. O. and. Aiyegoro, O. A (2009). Antimicrobial activity and phytochemical  
279 screening of stem bark extracts from *Jatropha curcas* (Linn). *African Journal of Pharmacy and*  
280 *Pharmacology*. 3 (2): 058-062
- 281 17. Harbone, J. B. (1999). *Phytochemical methods*. Chapman and Hall Limited, London. PP. 149 – 188  
282
- 283 18. Nishaa, S., Vishnupriya, M., Sasikumar, J.M. And Gopalakrishnan, V.K. (2013). Phytochemical  
284 Screening and GC-MS Analysis of Ethanolic Extract of Rhizomes of *Maranta arundinacea*. *Research*  
285 *Journal of Pharmaceutical, Biological and Chemical Sciences*. 4 (2): 52  
286  
287
- 288 19. Lutterodt, G.D., Ismail, A., Basheer, R.H. and Baharudin, H.M. (1999). Antimicrobial effects of *Psidium*  
289 *guajava* extracts as one mechanism of its antidiarrhoeal action. *Malaysia Journal of Medical Science*.  
290 6(2): 17-20  
291

292 20. Chapaga, B. and Wiesma, Z. (2005). Larvacidal effects of aqueous extracts of *Balanites aegyptiaca*  
293 (desert date) against the larva of culexpiplens mosquitoes. *African Journal of Biotechnology*, 4:1351-  
294 1354

295

296

297 .

298

299

300 .

301

302

UNDER PEER REVIEW