

**Antioxidant responses of three pepper
(*Capsicum annuum*) varieties against *Pepper
veinal mottle virus***

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ABSTRACT

Aims: This study aimed to investigate the changes in antioxidant activity and protein content between non-infected and infected leaves of three *Capsicum annuum* varieties against *Pepper veinal mottle virus*.

Material and methods: *Pepper veinal mottle virus* isolated from infected pepper plants was inoculated to three healthy varieties of pepper (Pepper Narval, Yolo Wonder and Chili pepper) by gently rubbing on the leaves of 14 days-old seedlings. Control peppers of each variety were treated in the same way with distilled water. The infection of inoculated plants was confirmed by the enzyme-linked immunosorbent assay method. Control and infected leaves were collected 21 days after inoculation (when symptoms manifested) and used for biochemical analyses. Change in different biochemical parameters (catalase, superoxide dismutase, malondialdehyde and protein) in infected pepper plant was observed compared to control non-infected ones.

Results: Catalase and superoxide dismutase activities were increased in Pepper Narval and Pepper Yolo Wonder infected leaves compared to non-infected, while a significant decrease was observed in infected Chili pepper compared to control. Higher malondialdehyde content was found in Pepper Yolo Wonder and Chili pepper infected leaves ($P < 0.05$) than control while a non-significant difference was shown between the infected and non-infected of Pepper Narval variety ($P > 0.05$). Infected Chili pepper showed high protein content compared to control ($P < 0.05$). An opposite trend was observed in pepper Narval and Yolo Wonder varieties ($P < 0.05$).

Conclusion: The results of this study showed that *Pepper veinal mottle virus* infection induces changes in enzymes, malondialdehyde and protein levels. These biochemical components were greatly expressed differentially between *Pepper veinal mottle virus* infected and non-infected in Pepper Yolo Wonder variety. Further studies with more biochemical parameters may contribute to improve the pepper tolerance mechanism to *Pepper veinal mottle virus* in a breeding program.

Keywords: *Pepper; PVMV; catalase; superoxide dismutase; malondialdehyde; protein.*

1. INTRODUCTION

Pepper (*Capsicum* spp.), including sweet pepper and hot pepper, is an important spice and vegetable crops worldwide [1]. Peppers belong to the Solanaceae family, genus *Capsicum* [2]. Among the five domesticated species of the genus *Capsicum* (*C. annuum*, *C. frutescens*,

21 *C. chinense*, *C. baccatum*, *C. pubescens*), *C. annuum* is the most widely grown in Africa [3].
22 In Burkina Faso, peppers are an important source of income for many small farmers. *C.*
23 *annuum* crop is grown in open fields on an estimated area of 1639 hectares with a production
24 of 8230 tons/year [4]. However, their average yields are severely affected by the presence of
25 pests and diseases. Pepper crop is infected by several fungal, bacterial, and viral diseases.
26 Among viral diseases, *Pepper veinal mottle virus* is endemic and the most devastating
27 pepper virus and other solanaceous crops in several West African countries [5]. The virus
28 was first discovered in Ghana [6] and then in other West African countries [7,5], Ethiopia [8],
29 and South Africa [9].

30 *Pepper veinal mottle virus* (PVMV) is a virus species in the genus Potyvirus of the family
31 Potyviridae [6]. PVMV is transmitted by aphids in a non-persistent manner and is
32 transmissible experimentally by mechanical inoculation. Symptoms expressed on the leaves
33 of plants infected with PVMV are characterized by chlorotic vein banding, mottling, mosaic,
34 and distortion with puckering of leaves. Infected plants may show stunting with reduced and
35 distorted fruit set [10]. PVMV causes significant losses for growers of solanaceous crops in
36 several African countries [11,12]. The incidence of the virus can reach 50 to 100%, leading
37 to significant losses in production causing whole field to be abandoned before harvest and in
38 some areas [13,14].

39 To control these pests, synthetic chemical pesticides are the most used. However,
40 chemicals could have secondary sides effect such as intoxication of farmers and consumers,
41 environmental pollution and the selection of strains resistant to pesticides [15,16].

42 Considerable efforts have focused on the development of pepper varieties resistant to the
43 virus. Early work resulted in materials that were tolerant or only partially resistant [17,18].

44 Another approach to select resistant plant by using physiological and biochemical
45 parameters was developed [19,20].

46 Indeed, the contact of the plant with the pathogen induces physiological and biochemical
47 reactions leading to the production of defense substances. The level of antioxidant activity
48 and total phenolic content of peppers infected with the virus reflects the condition of
49 resistance or susceptibility of pepper plants [21]. The identification of pepper biochemical
50 products expressed under virus infection will be helpful to improve *Capsicum annuum*
51 tolerance mechanism to PVMV in a breeding program. This study aimed to investigate the
52 changes in antioxidant activity and protein content of three peppers varieties against *Pepper*
53 *veinal mottle virus* infection in order to understand the biochemical tolerance mechanism of
54 *Capsicum annuum*.

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56 2. MATERIAL AND METHODS

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58 2.1 Plant material and experimental dispositive

59 Seeds of *C. annuum* were purchased from a commercial supplier in Ouagadougou, Burkina
60 Faso. Three varieties of peppers, pepper Narval (Na), pepper Yolo Wonder (Y) and Chili
61 pepper were sown in pots (25 cm diameter) containing sterilized sand and peat (1:1). For
62 each variety, two seeds were sown in pots in three replications. A control group and infected
63 group were defined for each variety. Plants were well watered and grown in a greenhouse
64 under insect-proof conditions all the experiment.

65 2.2 Inoculation of plants with *Pepper veinal mottle virus*

66 Virus isolates were obtained from the PVMV infected pepper plants grown in greenhouses
67 and propagated in pepper plants. The isolated PVMV was confirmed serologically by DAS-
68 ELISA. The inoculum was prepared according to the method described by Dikilitas *et al.*
69 [22]. Each pepper variety was then inoculated with the supernatant containing PVMV by
70 gently rubbing on the leaves of 14 days-old seedlings [23]. Control plants were treated in the

71 same manner using distilled water. All tests were performed in triplicate. After 21 days of
72 inoculation (when symptoms manifested), the leaves of each plant were collected to carried
73 out the ELISA and biochemical tests were carried out.

74 2.3 ELISA test

75 Pepper leaf samples were tested for the presence of PVMV in inoculated peppers by double
76 antibody sandwich enzyme-linked immunosorbent assay (DAS-ELISA). Leaves were ground
77 in phosphate buffered saline with Tween 20 using a pre-chilled mortar and pestle. The
78 extract was tested using polyclonal antisera produced by SEDIAG. Samples were
79 considered positive when absorbance values at 405 nm (A405) were at least three times
80 greater than the mean absorbance value of five healthy control samples [24,11,25].

81 2.4 Determination of antioxidant enzymes activities

82 2.4.1 Extraction of antioxidant enzymes

83 Fresh leaves (500 mg) were homogenized with 50 mM sodium phosphate (pH 7.8) and
84 centrifuged at 4000 rpm during 10 minutes. The supernatant was used to measure
85 superoxide dismutase and catalase enzymes activities.

86 2.4.2 Superoxide Dismutase (SOD) enzyme activity

87 Superoxide Dismutase activity was measured using the method described by Ranjitha and
88 Vijiyalakshmi [26] at 420 nm. The enzyme activity was expressed in terms of $\mu\text{mol/g}$ protein.

89 2.4.3 Catalase (CAT) enzyme activity

90 Catalase activity was measured using the method described by Ranjitha and Vijiyalakshmi
91 [26]. The absorbances were measured at 240 nm for each interval of 30 seconds during 3
92 minutes. The CAT activity was expressed in terms of μmol of H_2O_2 consumed/g protein.

93 2.5 Lipid peroxidation assay

94 The Malondialdehyde (MDA) content as the marker of lipid peroxidation was determined as
95 described by Mahi *et al.* [27]. The MDA content of samples was expressed in micromole per
96 milligram ($\mu\text{mol.mg}^{-1}$) of leaves fresh weight.

97 2.6 Protein content

98 Leaves (500 mg) were homogenized in 5 ml of 0.1 M NaCl. The samples were centrifuged at
99 4400 rpm during 30 min, and the supernatant was used to determine the protein content.
100 Protein concentration was determined by Bradford method as described by Mimouni *et al.*
101 [28].

102 2.7 Statistical analysis

103 The results are presented as mean \pm SD for triplicate analysis and were subjected to one-
104 way analysis of ANOVA variation with Tukey's Significant Difference test and $P < 0.05$ was
105 considered significant. The statistical analysis was performed using XLSTAT Version Pro-
106 2017 and the graphs were drawn using Graph Pad Prism software version 5.0.

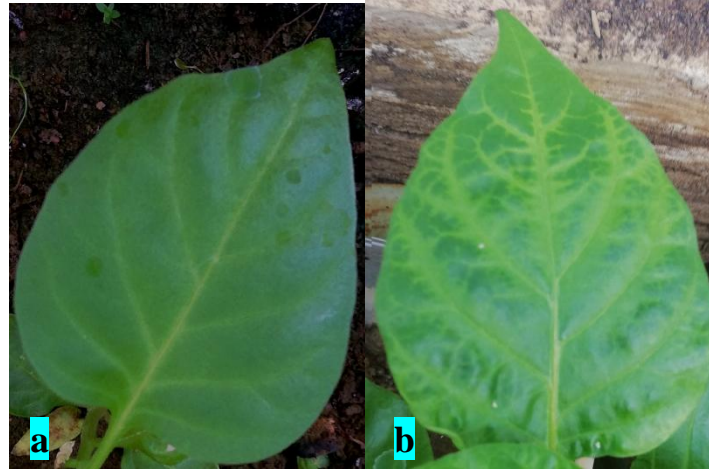
107 3. RESULTS

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109 3.1 Pathogenicity test

110 The different varieties of pepper inoculated with PVMV showed more or less severe
111 symptoms. Inoculated Chili pepper developed disease symptoms 2 weeks after inoculation
112 and developed severe symptoms such as chlorotic vein banding, mottling, mosaic, and
113 distortion. However, the inoculated Pepper Na and Pepper Y varieties developed slight
114 symptoms of chlorotic vein banding on some leaves three weeks after inoculation. These
115 observations were confirmed by ELISA-positive result for PVMV. On the contrary, no
116 symptoms were observed in control peppers plant and confirmed by ELISA-negative result
117 for PVMV. The results of the pathogenicity test are presented in Fig. 1.

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Fig. 1. Pepper leaf structures (a) healthy, (b) infected

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3.2 Enzymes antioxidant activities of non-infected and infected pepper varieties

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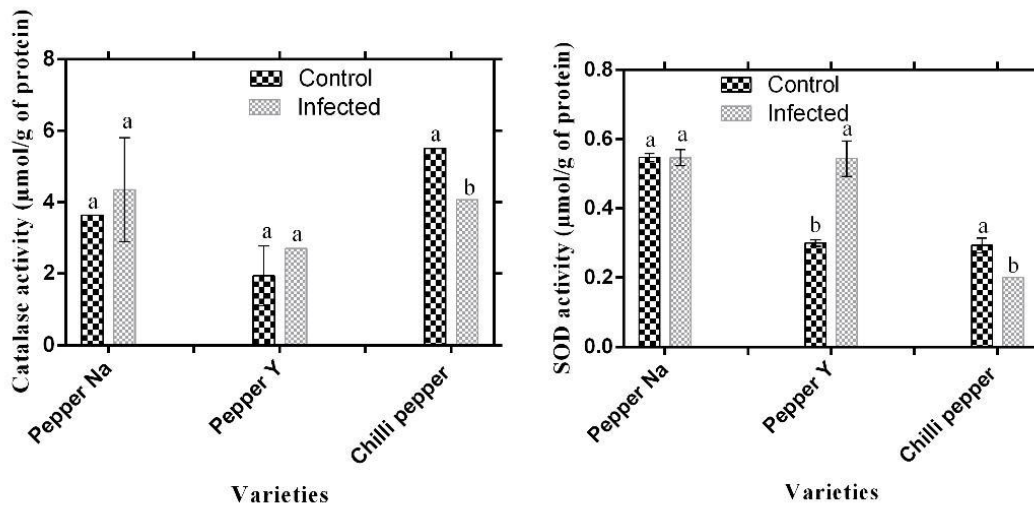
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Catalase (CAT) and superoxide dismutase (SOD) activities of control and PVMV-infected peppers are shown in Fig. 2. The activities of these enzymes were increased non-significantly in infected pepper Na variety compared to control ones. The activity of catalase enzyme increased insignificantly while the SOD enzyme activity increased significantly ($P < 0.05$) in infected pepper Y as compared to non-infected. A significant decrease ($P < 0.05$) in catalase and superoxide activities was observed in Chili pepper variety.

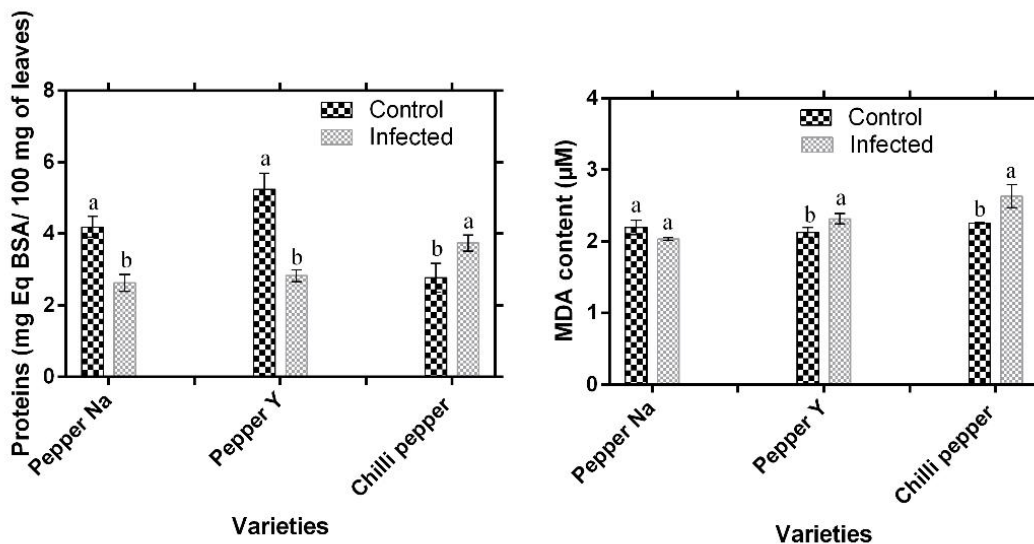


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130 **Fig. 2. Enzymes antioxidant activities of non-infected and PVMV-infected varieties of**
 131 **pepper**

132 **3.3 Lipid peroxidation of non-infected and infected pepper varieties**

133 The MalonDiAldehyde (MDA) content of control and PVMV-infected peppers is shown in Fig.
 134 3. The results showed **insignificant** decrease of MDA content only in infected Pepper Na
 135 variety compared with control ones. On the contrary, the MDA content of Pepper Y and Chilli
 136 pepper varieties infected with PVMV significantly increased ($P < 0.05$) when compared to
 137 control.



138

139 **Fig. 3. Protein and MDA contents of non-infected and PVMV-infected varieties of**
 140 **pepper**

141 **3.4 Proteins content of non-infected and infected pepper varieties**

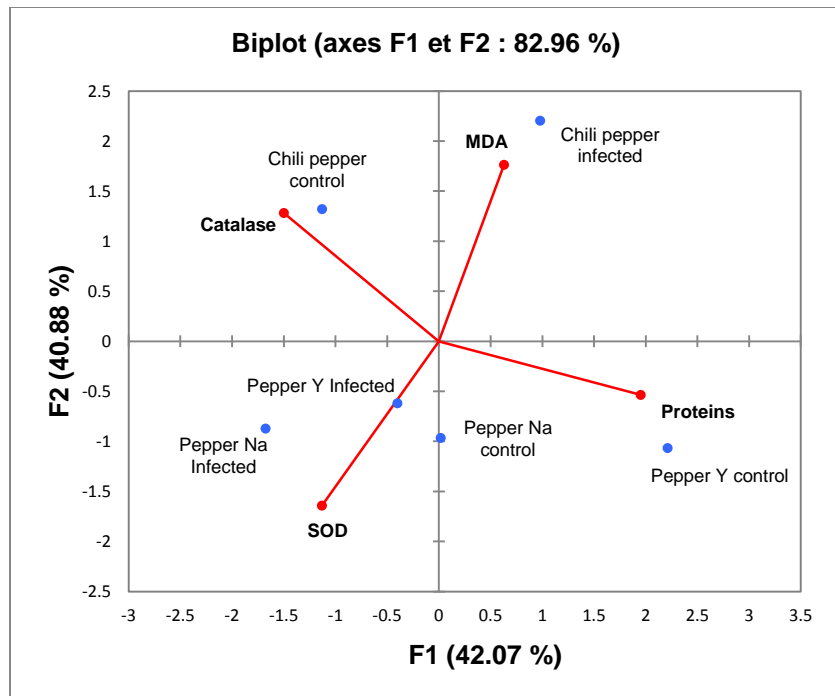
142 The protein content of control and PVMV-infected peppers is shown in Fig. 3. The protein
143 content differed significantly ($P < 0.05$) among control and infected of the three varieties of
144 pepper. The results revealed that the protein content decreased in infected pepper Na and
145 Y. However, Chili pepper showed significantly increased in protein content due to PVMV
146 infection compared with healthy ones.
147

148 **3.5 Comparative analysis of the protein content and antioxidant response of** 149 **pepper varieties to PVMV infection**

150 The principal component analysis was performed on the basis of variations in enzyme
151 antioxidant, MDA and proteins contents of different pepper varieties three (3) weeks after
152 inoculation. Fig. 4 presents the repartition of different parameters evaluated in the biplot axis.

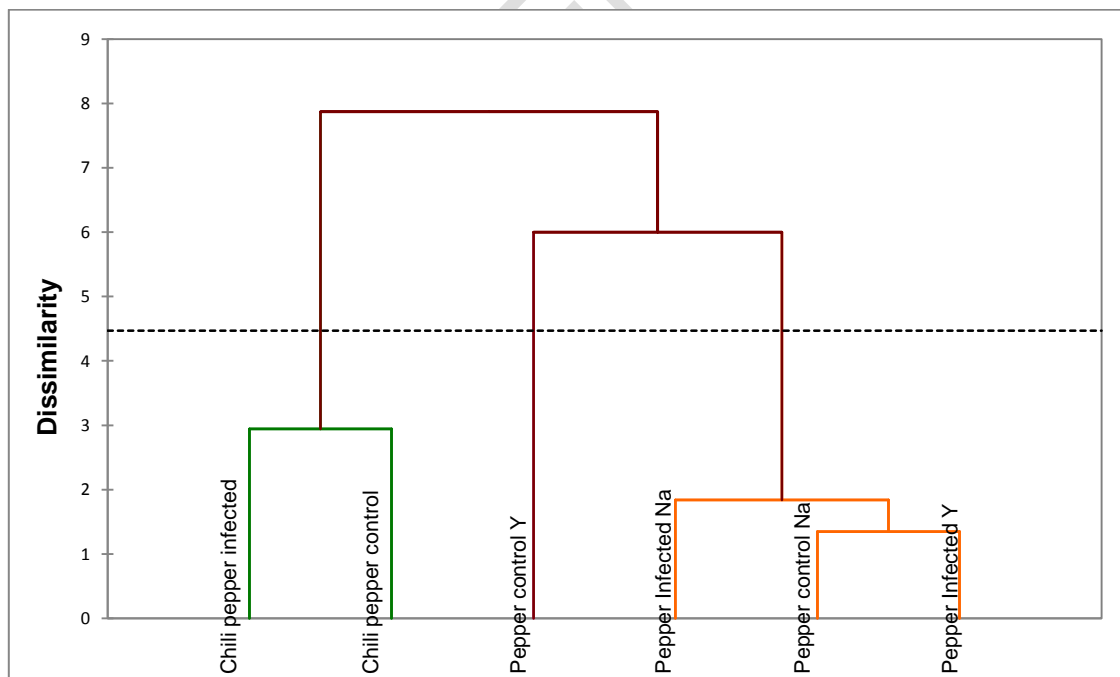
153 The first two principal components explained 82.96 % of the total variance. The first principal
154 component (F1) and the second principal component (F2) account, respectively for 42.07 %
155 and 40.88 % of the total variation. The first principal component (F1) separated Chili pepper
156 control from Chili pepper infected in one hand and Pepper Y infected, Pepper Na infected
157 from Pepper Y control and Pepper Na control in other hand. The F1 axis divided the control
158 non-infected group from infected group of the different varieties of pepper. The second
159 principal component (F2) separated Chili pepper (control and infected) from Pepper Y and
160 Pepper Na (control and infected). The F2 axis divided the pepper in different part according
161 to the type of pepper variety. Analysis of the correlations between the evaluated variables
162 and factors showed a strong contribution of Pepper Y infected, Pepper Na infected and
163 Pepper Na control to SOD while Chili pepper infected contributes strongly to MDA. Pepper Y
164 control contributes strongly to proteins, while Chili pepper control contributes strongly to
165 catalase. After the construction of dendrogram of the different treatments, the treatments
166 were grouped into three main classes, I, II and III (Fig. 5). Class I comprised Pepper Na
167 control, Pepper Na infected and Pepper Y **infected**. Class II comprised Chili pepper control
168 and Chili pepper infected. Class III is constituted only of Pepper Y control.

169



170

171 **Fig. 4. Principal component analysis of the different pepper varieties responses on**
 172 **the protein content and antioxidant activities to PVMV infection**



173

174 **Fig. 5. Dendrogram of the different pepper varieties response based on the protein**
 175 **content and antioxidant activities to PVMV infection.**

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177 4. DISCUSSION

178 This study was conducted to investigate the changes in antioxidant activity and protein
179 content of three peppers varieties against PVMV infection in order to understand the
180 biochemical tolerance mechanism of *Capsicum annum*.

181 The results of the pathogenicity test showed an early development of diseases symptoms
182 and severity in the Chili pepper variety due to PVMV infection compared to the Pepper Na
183 and Pepper Y varieties. Besides, the ELISA tests were positive for all **infected** peppers. The
184 Chili pepper **was** more **sensitive** to PVMV infection compared to the **other** varieties of
185 pepper. The susceptibility of Chili pepper is confirmed by the decrease of catalase and
186 superoxide dismutase activities after PVMV infection when compared to non-infected group.
187 Only pepper Y variety showed a significant increase of SOD activity in infected group
188 compared to non-infected group. The pepper Yolo Wonder variety is tolerant to PVMV by
189 increasing the activity of superoxide dismutase, enzyme involved against the biotic or abiotic
190 stress of plant. Appiah **et al.** [29] showed that pepper plants respond differently to a viral
191 infection based on their susceptibility or resistance to viruses. Antioxidant enzymes (CAT,
192 SOD) are produced by host plant to promote cells protection of oxidative damage from
193 pathogens [30]. They induce resistance against pathogen [31]. Similar results have been
194 reported by Siddique **et al.** [32]. They showed an increase of the CAT and SOD activities in
195 the leaves of resistant varieties of cotton and a decrease in the susceptible varieties after
196 infection with *Cotton Leaf Curl Burewala Virus*.

197 Generally, infected plants show a high content of protein, which could be due to both of the
198 activation of the host defense mechanism and the pathogen attack mechanism [33]. In this
199 study, protein content significantly decreased in infected Pepper Na and Pepper Y compared
200 to control non-infected. An opposite trend was observed in Chili pepper plants. The increase
201 in protein content in Chili pepper after infection may be due to viral replication which could
202 explain it high susceptibility to PVMV. Indeed, Zinga **et al.** [34] showed that protein content is
203 higher in cassava leaves infected by *African Mosaic Virus* than in healthy ones. However,
204 other investigators have shown an increase in protein content in resistant infected varieties
205 [32,35].

206 **MalonDiAldehyde** is a general indicator of lipid peroxidation [36]. MDA produced during lipid
207 peroxidation is an indicator of cellular membrane damage to the cell membrane caused by
208 pathogenic infection [27]. Infection of Pepper Y and Chili pepper with PVMV resulted of an
209 increase of the MDA content compared to control non-infected. Previous studies have shown
210 that MDA content tends to increase in susceptible varieties due to infection. Lanubile **et al.**
211 [37] obtained the same result with maize leaves corn **infected** by *Aspergillus niger*. Analysis
212 of the principal components revealed a negative correlation between MDA and SOD. Chili
213 pepper infected contributes strongly to MDA while infected Pepper Na and infected pepper Y
214 contribute strongly to SOD. MDA increasing translates cellular degradation while SOD
215 enhancement induces cellular defense mechanism [27,31]. The Pepper Na and Y varieties
216 produce chemicals inducing resistance to PVMV than Chili pepper. Sama **et al.** [35] showed
217 that the leaves of susceptible varieties of *Jatropha* strongly contribute to the MDA content
218 after infection with *Lasiodiplodia theobromae*.

219 Combination in classes of three varieties of uninfected and infected peppers revealed a
220 relationship between control and infected Chili pepper in class II. This closeness might be
221 due to a weak response of the measured parameters (protein and MDA content, antioxidant
222 enzymes activities) of this variety to the viral infection. Chili pepper presented a susceptible
223 reaction against the PVMV. Pepper Y control and Pepper Y infected are in different classes.
224 Likewise, Pepper Na control and infected are in the same class but in different subclasses.

225 This may explain by the important biochemical response of Pepper Na and Y varieties due to
226 viral infection.

227 In view of parameters of the oxidative **stress** enzymes (CAT, SOD) and MDA of the infected
228 pepper varieties then the grouping into classes of the different uninfected and infected
229 varieties, we can conclude that Pepper Y induce resistance against PVMV infection than
230 Pepper Na and Chili pepper.

231 Biochemical responses of pepper varieties during disease reaction indicated that there was
232 a variation in the enzyme activity linked to infection. In our study, under viral infection
233 conditions, stimulation and increased SOD enzyme activity play an important role in defense
234 mechanisms of peppers. This enzyme could induce pepper tolerance to *Pepper veinal mottle*
235 *virus*.

236 4. CONCLUSION

237

238 This study found variations in the activity of oxidative **stress** enzymes, MDA and protein
239 contents between the three **infected** and non-**infected** pepper varieties. SOD is involved in
240 the biochemical defense mechanisms controlling the development of PVMV in Pepper Yolo
241 Wonder variety. Under stressful conditions such as viral infection, stimulation of biochemical
242 parameters plays a vital role in the defense mechanism. The results of this study suggest
243 that the Pepper Yolo Wonder variety is more tolerant to *Pepper veinal mottle virus* than the
244 Pepper Narval and Chili pepper varieties. Further studies with more biochemical parameters
245 related to pathogenicity may contribute to improve the pepper tolerance mechanism to
246 *Pepper veinal mottle virus* in a breeding program.

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249 COMPETING INTERESTS

250

251 Authors have declared that no competing interests exist.

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UNDER PEER REVIEW