

## **Eosinopenia Associated with Infection is an Independent Risk Factor for 28-day Mortality in *Staphylococcus aureus* Bloodstream Infection**

### **Abstract**

**Aims:** This retrospective study aimed to evaluate the impact of eosinopenia on 28-day mortality in *Staphylococcus aureus* bloodstream infection (SABSI) .

**Methods:** A retrospective study was designed to evaluate the impact of eosinopenia on 28-day mortality in SABSI.

**Results:** Patients who were  $\geq 16$  years old with SABSI at Sun Yat-Sen Memorial Hospital between January 1<sup>st</sup> 2014 and December 31<sup>st</sup> 2018 were included. The overall 28-day mortality of all patients was 14.3% (44 out of 307). Patients with eosinopenia in the onset of SABSI had a significantly higher 28-day mortality than those without eosinopenia (22.4% vs 6.5%;  $P < 0.01$ ). For patients who developed SABSI after the first 48 hours in the hospital, eosinophils decreased significantly from the baseline ( $P < 0.01$ ). Kaplan–Meier survival curve showed that patients with eosinopenia had a lower survival rate than those without eosinopenia ( $P < 0.01$ ). Multivariate Cox regression analysis revealed that eosinophils in the onset of SABSI were associated independently with 28-day mortality (hazard ratio [HR], 2.84; 95% confidence interval [CI], 1.36–5.91;  $P < 0.01$ ).

**Conclusion:** Eosinopenia associated with infection might be an independent risk factor for 28-day mortality in SABSI.

**Keywords:** *Staphylococcus aureus*, bloodstream infection, prognosis, eosinopenia

**Abbreviation:** SABSI, *Staphylococcus aureus* bloodstream infection

### **1. Introduction**

The *Staphylococcus aureus* bloodstream infection (SABSI) is a common condition with a high fatality rate, an important cause of morbidity and mortality all over the world [1]. In North America, the incidence of SABSI has ranged between 20 and 40 cases per 100,000 population, with an increase demonstrated over the last two decades in some regions [2]. In

an epidemiological study from 2011 to 2013 in Alberta of Canada, there were 299 cases of Methicillin-Resistant *Staphylococcus aureus* bloodstream infection, equating to 3.95 cases per 100,000 population[3].

So far, foreseeing the prognosis of SABSI in the early stage remains a huge challenge. Some biological parameters such as C-reactive protein and procalcitonin, have been used to determine the diagnosis of infection or bacteraemia, but these biomarkers are not necessarily associated with the prognosis[4-6]. Moreover, as the resources are limited, the use of some biomarkers remains unavailable in some developing countries. Therefore, an ideal biomarker which is highly specific and sensitive, easy to measure and inexpensive is in urgent need.

Unlike the biological parameters mentioned above, the routine analysis of blood is economical, thus put into use extensively among patients. Eosinophils are multifunctional cells of the innate immune system linked

to allergic and parasitic inflammation in the traditional perspective, and are generally interpreted as purely detrimental. Surprisingly, the intricate relationship between eosinophils and infectious diseases has been delineated detailed in recent researches, in which eosinophils have been shown to have a unique protective role in the setting of nonparasitic infectious diseases. In infectious-related asthma patients, the phagocytosis of bacteria by eosinophils might be a dominating pathophysiological process [7]. Caroline et al [8] confirmed that degranulating airway eosinophils promoted survival in virus infection, and activated eosinophils from both *Aspergillus* antigen and cytokine-driven asthma models were profoundly antiviral and promoted survival in an otherwise lethal pneumonia virus of mice infection. In mice infected with influenza A virus, eosinophils were susceptible to the virus and responded by activation, piecemeal degranulation and upregulation of antigen presentation markers. The transfer of eosinophils from lungs of allergen-sensitized and challenged mice to influenza virus-infected mice reduced morbidity and viral burden, improved lung compliance, and increased CD8(+) T cell numbers in the airways[9].

To the best of our knowledge, neither any work has investigated the incidence of eosinopenia in SABSIs, nor the potential association between theosbee patients.

Therefore, the study designed to evaluate the impact of eosinopenia on 28-day mortality in SABSIs. In this study, we showed that eosinophils were susceptible to SABSIs and eosinopenia associated with infection was an independent risk factor for 28-day mortality in SABSIs.

## 2. Materials and Methods

### 2.1. Study population

We performed a retrospective cohort study at Sun Yat-Sen Memorial Hospital between January 1st 2014 and December 31st 2018. Sun Yat-Sen Memorial Hospital is a 2800-bed primary care and tertiary referral centre in South China. Patients who were  $\geq 16$  years of age with SABSIs were chosen from the computerized database of the hospital's clinical microbiology laboratory. For patients who had more than one episode of SABSIs, only the first episode was selected for this study. Patients with a length of stay shorter than 48 hours after the episode of SABSIs were excluded.

### 2.2. Study design

The study reviewed retrospectively the medical records of all the eligible patients. The data collected included demographic, clinical, microbiological data and the outcome. The main outcome was 28-day mortality. All of the patients had given their informed consent for the medical and the study was approved by the Institutional Review Board.

### 2.3. Definitions

SABSIs were defined as the isolation of *Staphylococcus aureus* in a blood culture. Initial antibiotic therapy was considered adequate when at least one adequate antimicrobial was given within 24 hours of SABSIs onset, and the dose and pattern of administration must be in accordance with current medical standards. The adequacy of the antimicrobial was determined by *in vitro* susceptibility for the causative microorganism[10]. There has been no unified standard for eosinopenia so far. The standards adopted

in the previous researches were varied from  $0.01 \times 10^9/L$  to

$0.04 \times 10^9/L$  and the definition  $0.02 \times 10^9/L$  adopted in the present research actually was based on the distribution of eosinophils in the onset of SABS. I.

2.4. Microbiological and biochemical testing Blood cultures, consisting of aerobic and anaerobic samples, were processed at the clinical laboratory of the hospital. The Vietk 2 system (bioMérieux, Marcy l'Etoile, France) was used for isolate identification and antimicrobial susceptibility testing. Minimum inhibitory concentrations were classified according to the Clinical Laboratory and Standards Institute criteria used in the corresponding year. Blood counts were measured by a XE-5000 haematology analyzer (Sysmex, Kobe, Japan), and levels of serum creatinine and total bilirubin were measured by a TBA-2000FR hematology biochemical analyzer (Toshiba, Tokyo, Japan). Patients were divided into two cohorts depending on whether the SABS. I was onset in the first 48 hours in the hospital or not. The data of blood counts in the onset of SABS. I were collected in all patients, and data of blood counts in the first hospital day were also collected in those who had SABS. I onset after the first 48 hours in the hospital.

#### 2.5. Statistical analysis

outcome was all-cause 28-day mortality. For convenience, some continuous parameters were dichotomised at the median, including leukocytes, platelets, haemoglobin, neutrophils, lymphocytes, monocytes, eosinophils, basophils, serum creatinine and total bilirubin.

Firstly, baseline differences between survivors and non-survivors were compared. Parametric variables (except the dichotomized ones) were described as mean (standard deviation, SD), while non-parametric variables as median (interquartile range, IQR). Means were executed with Student's t test or paired Student's t test when appropriate. Medians and comparative analysis were executed with Mann–Whitney U test, Kruskal–Wallis test or chi-square test when appropriate.

Secondly, the Kaplan–Meier survival curve was used to assess the association between eosinopenia and all-cause 28-day mortality, and log rank test was done to compare the survival curves.

Thirdly, Cox proportional hazards regression models were performed, in which hazard ratio (HR) and 95% confidence interval (CI) were reported, to assess predictors of 28-day mortality. Variables with a  $P < 0.05$  in univariate analysis, along with age and sex, were entered in the multivariate Cox regression model. All P values were 2-tailed and statistical significance was set at  $P < 0.05$ . All statistic analyses were performed using SPSS version 20.0 (SPSS, Inc., Chicago, IL, USA).

### 3. Results

#### 3.1. Study population

During the study period, a total of 307 patients with SABSIs were identified. The mean age ( $\pm$ SD) of these patients was  $56.9\pm 17.7$  years, and 65.5% were male ( $n=201$ ). The most common underlying disease was hypertension ( $n=107$ , 34.9%), followed by solid organ malignancy ( $n=94$ , 30.6%). Comparisons between survivors and non-survivors in the whole study population were summarized in Table 1. The disease severity in the non-survivor group was much more critical than that of the other group. Drug-resistance bacteria and receiving an inappropriate initial empirical antimicrobial therapy were more common in the non-survivors. For the routine analysis of blood in the onset of SABSIs, no significant differences in the leukocytes, neutrophils, monocytes, basophils or lymphocytes were observed between the two groups, while the reduction of platelets, haemoglobin and eosinophils in non-survivors were much more frequent. The median of eosinophil counts in the onset of SABSIs

was  $0.02\times 10^9/L$ , which happened to be the boundary of eosinopenia according to the criterion used in the clinical laboratory. There were 101 patients who had SABSIs onset in the first 48 hours in the hospital, while 206 patients developed SABSIs after 48 hours. For patients who developed SABSIs after 48 hours in the hospital, paired Student's *t* test showed that eosinophils decreased significantly after the SABSIs (Difference value=0.04,  $P<0.01$ ).

### 3.2. 28-day mortality and predictors of mortality

The overall 28-day mortality of all patients was 14.3% (44 out of 307). Patients in the onset of SABSIs had a significantly higher 28-day mortality than those without eosinopenia (22.4% vs 6.5%;  $P<0.01$ ). The survival curve showed that patients with eosinopenia had a lower probability of survival than those without eosinopenia (Fig. 1, log rank test,  $P<0.01$ ). In the subgroup analysis, for patients who developed SABSIs after 48 hours in hospital, the survival curve showed the same trend among the

prognosis and eosinopenia in the onset of SABSİ (Fig. 2, log rank test,  $P<0.01$ ), while eosinopenia presenting in the first day in hospital in these patients was not associated with the prognosis ( $P=0.09$ ). By multivariate Cox proportional hazards regression analysis, eosinophils in the onset of SABSİ were associated independently with 28-day mortality (HR, 2.84; 95% CI, 1.36–5.91;  $P<0.01$ ), while the other blood hemocytes were not (Table 2). Factors associated with 28-day mortality

also included male, serum creatinine, total bilirubin and inappropriate empiric antimicrobial therapy. In the subgroup analysis, for patients who developed SABSİ after 48 hours in the hospital, eosinophils in the onset of SABSİ were associated independently with 28-day mortality (HR, 3.20; 95% CI, 1.23–8.14;  $P<0.01$ ), while eosinophils in the first day were not

(Table 3).

#### 4. Discussion

Our study demonstrated that eosinopenia in the onset of SABSİ might be a predictive factor of the 28-day mortality of SABSİ. It was the first time to explore the relationship of evolution of eosinophil and the prognosis of SABSİ. As the eosinophil count is given for each patient, it would be a great gain for no additional outlay if eosinopenia is proven to be a useful prognostic marker.

Eosinophils have been traditionally perceived largely as end-stage, cytotoxic effector cells associated with allergy and parasitic diseases[11-13]. Production of eosinophils is tightly regulated by interleukin-3, interleukin-5 and granulocyte–macrophage colony-stimulating factor[14,15]. Studies have confirmed that the decline of eosinophil count is associated with any of the three processes: peripheral sequestration of eosinophils, suppression of egress of mature eosinophils and suppression of eosinophil production[16]. By expressing specific cytokines associated with inner immunity, eosinophils might play an indispensable role in viral infection. In mouse airways in vivo and in isolated human

eosinophils in vitro, eosinophils mediated the antiviral effect via the production of nitric oxide and by serving as a dead-end host for virus infection. Eosinophils produced nitric oxide in response to virus and to a synthetic agonist of the virus-sensing innate immune receptor, Toll-like receptor (TLR) 7 [17]. In another research, eosinophils were able to limit lung dysfunction associated with the respiratory syncytial virus, via surface and intracellular TLR associated with antiviral immunity and responding functionally to TLR ligands[18]. Shigeharu et al[19] demonstrated that activated human eosinophils can undergo extracellular DNA trap cell death (ETosis) that cytolytically releases free eosinophil granules, and EETosis resulted in the generation of histone-bearing nuclear DNA extracellular nets and cell-free granules, both of which may exert biological activities for eosinophils postmortem.

On the contrary, in the territory of bacterial infection, the relationship between the eosinophil count and the inner immunity are poorly understood. The mechanism of eosinopenia in bacterial infectious diseases is much less reported. It is a conventional knowledge that cortisol increases leukocyte and neutrophil counts, whereas it reduces lymphocyte, monocyte and eosinophil counts. Some study suggested that eosinopenia can develop from acute severe stress of infectious or noninfectious, which is mediated by adrenal glucocorticoids and epinephrine[20-22]. Basic research showed that *Staphylococcus aureus* mediated rapid eosinophil cell death, and the cytolysin was a major contributory factor in eosinophil death[23]. Bass showed that both the infectious and noninfectious stimuli of acute inflammation markedly suppressed eosinophilia, which suggested that eosinopenia was a response to the acute inflammatory process rather than to a specific type of pathogen[16]. Erica et al[24] found an essential role for eosinophils in the immune response that reduces pathology associated with *Clostridium difficile* infection. In this process, the eosinophil number increased via microbiota-regulated interleukin-25. In the present study, eosinopenia in the onset of SABSIs was found to be an independent risk factor of 28-day mortality. For patients who developed SABSIs after 48 hours in the hospital, eosinophils in the onset of SABSIs decreased significantly from the baseline values in the first day, and the subgroup analysis shows that only eosinopenia associated with the infection was an independent risk factor of mortality. Therefore, given that patients were under severe stress of the bloodstream infection, we assume that there might be a powerful stimulation of adrenal glucocorticosteroid produced by the stress of the infection, though the level of patients' glucocorticosteroid had not been detected. As an acute physiological reaction to the SABSIs, eosinopenia did harm to the SABSIs in turn. Although eosinophils are not usually considered to play a crucial role in immune defences against bacteria, some studies have shown they possess anti-bacterial capabilities, mediated by granule contents[25,26] or release of mitochondrial DNA[27]. A retrospective study also found that eosinophil behaved as a protective cell in patients with ventilator-associated pneumonia caused by

Staphylococcus aureus[28].

## 5. Conclusion

In conclusion, the decline of eosinophils is an early message associated with the severity of SABSI, and eosinopenia associated with infection might be an independent risk factor for mortality in SABSI. Further studies are needed to demonstrate how the infection and eosinopenia interact with each other.

## 6.4. Patient Consent

All authors declare that written informed consent was obtained from the patients for publication of this report. A copy of the written consent is available for review by the Editorial office/Chief Editor/Editorial Board members of this journal.

## 6.5. Ethical approval

The study protocol has been approved by the research institute's

committee on human research.



## References:

- 1 Widmer AF, Lakatos B, Frei R: Strict infection control leads to low incidence of methicillin-resistant *Staphylococcus aureus* bloodstream infection over 20 years. *Infect Control Hosp Epidemiol* 2015;36:702-709.
- 2 Kern WV: Management of *Staphylococcus aureus* bacteremia and endocarditis: Progresses and challenges. *CURR OPIN INFECT DIS* 2010;23:346-358.
- 3 Taylor G, Bush K, Leal J, Henderson E, Chui L, Louie M: Epidemiology of methicillin-resistant *Staphylococcus aureus* bloodstream infections in Alberta, Canada. *J HOSP INFECT* 2015;89:132-135.
- 4 Durnas B, Watek M, Wollny T, Niemirowicz K, Marzec M, Bucki R, Gozdz S: Utility of blood procalcitonin concentration in the management of cancer patients with infections. *Onco Targets Ther* 2016;9:469-475.
- 5 Ozsurekci Y, Oktay AK, Bayhan C, Karadag-Oncel E, Emre AA, Gurbuz V, Hascelik G, Ceyhan M: Can procalcitonin be a diagnostic marker for catheter-related blood stream infection in children? *J Pediatr (Rio J)* 2016;92:414-420.
- 6 Guo SY, Zhou Y, Hu QF, Yao J, Wang H: Procalcitonin is a marker of gram-negative bacteremia in patients with sepsis. *AM J MED SCI* 2015;349:499-504.
- 7 Cohen SG, Sapp TM: Phagocytosis of bacteria by eosinophils in infectious-related asthma. *The Journal of allergy* 1969;44:113.
- 8 Percopo CM, Dyer KD, Ochkur SI, Luo JL, Fischer ER, Lee JJ, Lee NA, Domachowske JB, Rosenberg HF: Activated mouse eosinophils protect against lethal respiratory virus infection. *BLOOD* 2014;123:743-752.
- 9 Samarasinghe AE, Melo RC, Duan S, LeMessurier KS, Liedmann S, Surman SL, Lee JJ, Hurwitz JL, Thomas PG, McCullers JA: Eosinophils promote antiviral immunity in mice infected with influenza a virus. *J IMMUNOL* 2017;198:3214-3226.
- 10 Chen HC, Lin WL, Lin CC, Hsieh WH, Hsieh CH, Wu MH, Wu JY, Lee CC: Outcome of inadequate empirical antibiotic therapy in emergency department patients with community-onset bloodstream infections. *J Antimicrob Chemother* 2013;68:947-953.
- 11 Carranza-Rodriguez C, Escamilla-Gonzalez M, Fuentes-Corripio I, Perteguer-Prieto MJ, Garate-Ormaechea T, Perez-Arellano JL: [Helminthosis and eosinophilia in Spain (1990-2015)]. *Enferm Infecc Microbiol Clin* 2016
- 12 Yenigun A, Sezen S, Calim OF, Ozturan O: Evaluation of the eosinophil-to-lymphocyte ratio in pediatric patients with allergic rhinitis. *AM J RHINOL ALLERGY* 2016;30:21-25.
- 13 Fabre V, Beiting DP, Bliss SK, Gebreselassie NG, Gagliardo LF, Lee NA, Lee JJ, Appleton JA:

- Eosinophil deficiency compromises parasite survival in chronic nematode infection. *J IMMUNOL* 2009;182:1577-1583.
- 14 Bass DA, Gonwa TA, Szejda P, Cousart MS, DeChatelet LR, McCall CE: Eosinopenia of acute infection: Production of eosinopenia by chemotactic factors of acute inflammation. *J CLIN INVEST* 1980;65:1265-1271.
- 15 Hogan SP, Rosenberg HF, Moqbel R, Phipps S, Foster PS, Lacy P, Kay AB, Rothenberg ME: Eosinophils: Biological properties and role in health and disease. *CLIN EXP ALLERGY* 2008;38:709-750.
- 16 Bass DA: Behavior of eosinophil leukocytes in acute inflammation. II. Eosinophil dynamics during acute inflammation. *J CLIN INVEST* 1975;56:870-879.
- 17 Drake MG, Bivins-Smith ER, Proskocil BJ, Nie Z, Scott GD, Lee JJ, Lee NA, Fryer AD, Jacoby DB: Human and Mouse Eosinophils Have Antiviral Activity against Parainfluenza Virus. *AM J RESP CELL MOL* 2016;55:387-394.
- 18 Phipps S, Lam CE, Mahalingam S, Newhouse M, Ramirez R, Rosenberg HF, Foster PS, Matthaei KI: Eosinophils contribute to innate antiviral immunity and promote clearance of respiratory syncytial virus. *BLOOD* 2007;110:1578-1586.
- 19 Ueki S, Melo RC, Ghiran I, Spencer LA, Dvorak AM, Weller PF: Eosinophil extracellular DNA trap cell death mediates lytic release of free secretion-competent eosinophil granules in humans. *BLOOD* 2013;121:2074-2083.
- 20 Spreng M: Possible health effects of noise induced cortisol increase. *NOISE HEALTH* 2000;2:59-64.
- 21 Levinson AT, Casserly BP, Levy MM: Reducing mortality in severe sepsis and septic shock. *Semin Respir Crit Care Med* 2011;32:195-205.
- 22 Brohee D, Vanhaeverbeek M, Kennes B, Neve P: Leukocyte and lymphocyte subsets after a short pharmacological stress by intravenous epinephrine and hydrocortisone in healthy humans. *INT J NEUROSCI* 1990;53:53-62.
- 23 Prince LR, Graham KJ, Connolly J, Anwar S, Ridley R, Sabroe I, Foster SJ, Whyte MK: Staphylococcus aureus induces eosinophil cell death mediated by alpha-hemolysin. *PLOS ONE* 2012;7:e31506.
- 24 Buonomo EL, Cowardin CA, Wilson MG, Saleh MM, Pramoongago P, Petri WA: Microbiota-regulated IL-25 increases eosinophil number to provide 1 protection during Clostridium difficile infection. *CELL REP* 2016;16:432-443.
- 25 Torrent M, de la Torre BG, Nogues VM, Andreu D, Boix E: Bactericidal and membrane disruption activities of the eosinophil cationic protein are largely retained in an N-terminal fragment.

26 Driss V, Legrand F, Hermann E, Loiseau S, Guerardel Y, Kremer L, Adam E, Woerly G, Dombrowicz D, Capron M: TLR2-dependent eosinophil interactions with mycobacteria: Role of alpha-defensins. BLOOD 2009;113:3235-3244.

27 Yousefi S, Gold JA, Andina N, Lee JJ, Kelly AM, Kozlowski E, Schmid I, Straumann A, Reichenbach J, Gleich GJ, Simon HU: Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. NAT MED 2008;14:949-953.

28 Rodriguez-Fernandez A, Andaluz-Ojeda D, Almansa R, Justel M, Eiros JM, Ortiz DLR: Eosinophil as a protective cell in *S. Aureus* ventilator-associated pneumonia. Mediators Inflamm 2013;2013:152943.

**Table 1**

**Comparisons between survivors and non-survivors in the whole study population**

Variable	Survivors (N=263)	Non-survivors (N=44)	All patients (N=307)	<i>P</i> value
Male gender	178(67.7%)	23(52.3%)	201(65.5%)	0.05
Age(Mean, $\pm$ SD)	56.3( $\pm$ 7.9)	60.4( $\pm$ 6.0)	56.9( $\pm$ 7.7)	0.13
LOS before BSI(Mean, $\pm$ SD)	15.2( $\pm$ 21.2)	16.9( $\pm$ 22.3)	15.2( $\pm$ 21.4)	0.58

Neutropenia <sup>a</sup>	9(3.4%)	4(9.1%)	13(4.2%)	0.19
Prior surgery or trauma <sup>a</sup>	84(31.9%)	19(43.2%)	103(33.6%)	0.14
With previous hospitalization in the preceding 90 days	122(46.4%)	27(61.4%)	149(48.5%)	0.07
Antibiotics therapy	121(46.0%)	30(68.2%)	151(49.2%)	<0.01
Prior chemotherapy or radiotherapy <sup>a</sup>	45(17.1%)	6(13.6%)	51(16.6%)	0.57
Dialysis or filtration <sup>a</sup>	9(4.0%)	3(7.7%)	12(4.6%)	0.55
Mechanical ventilation <sup>b</sup>	49(18.6%)	14(31.8%)	63(20.5%)	0.05
Indwelling central venous catheter <sup>b</sup>	101(38.4%)	27(61.4%)	128(41.7%)	<0.01
Indwelling nasogastric tube <sup>b</sup>	65(24.7%)	20(45.5%)	85(27.7%)	<0.01
Indwelling urinary catheter <sup>b</sup>	84(31.9%)	24(54.5%)	108(35.2%)	<0.01
<b>Underlying disease</b>				
Solid organ malignancy	78(29.7%)	16(36.4%)	94(30.6%)	0.37
Hematological malignancy	13(4.9%)	3(6.8%)	16(5.2%)	0.60
Chronic lung disease	14(5.3%)	1(2.3%)	15(4.9%)	0.62
Cerebrovascular disease	36(13.7%)	5(11.4%)	41(13.4%)	0.68
Chronic cardiac failure	38(14.8%)	11(25.0%)	50(13.6%)	0.09
Hypertension	90(34.2%)	17(38.6%)	107(34.9%)	0.57
Atrial fibrillation	11(4.2%)	4(9.1%)	15(4.9%)	0.31
Liver cirrhosis	18(6.8%)	6(13.6%)	24(7.8%)	0.21
Chronic renal failure	31(11.8%)	3(6.8%)	34(11.1%)	0.48
Diabetes mellitus	58(22.1%)	10(22.7%)	68(22.1%)	0.92
Autoimmune disease	36(13.7%)	8(18.2%)	44(14.3%)	0.43

Abbreviation: LOS=length of stay. Values

are n (%) unless otherwise noted. <sup>a</sup>Within

30 days preceding infection onset.

<sup>b</sup>Within 48 hours preceding infection onset.

**Table 1**

**Comparisons between survivors and non-survivors in the whole study population (continued)**

Variable	Survivors(N=263)	Non-survivors (N=44)	All patients (N=307)	P value
Charlson score(Mean, $\pm$ SD)	1.9( $\pm$ 1.9)	2.7( $\pm$ 2.2)	2.0( $\pm$ 1.9)	<0.01
Pitt score(Mean, $\pm$ SD)	1.7( $\pm$ 2.0)	3.1( $\pm$ 2.8)	1.9( $\pm$ 2.2)	<0.01
APACHE II score(Mean, $\pm$ SD)	11.3( $\pm$ 5.5)	16.3( $\pm$ 7.3)	12.0( $\pm$ 6.1)	<0.01
Drug-resistance bacteria	173(65.8%)	39(88.6%)	212(69.1%)	<0.01
MRSA	114(43.3%)	25(56.8%)	139(45.3%)	0.10
Inappropriate empiric antimicrobial therapy	109(41.4%)	31(70.5%)	140(45.6%)	<0.01
* Serum creatinine <sup>c</sup>	129(49.0%)	28(63.6%)	157(51.1%)	0.07
*Total bilirubin <sup>c</sup>	127(48.3%)	31(70.5%)	158(51.5%)	<0.01
*Leukocytes <sup>c</sup>	130(49.4%)	24(54.5%)	154(50.2%)	0.53
*Hemoglobin <sup>c</sup>	122(46.4%)	32(72.7%)	154(50.2%)	<0.01
*Platelets <sup>c</sup>	118(44.9%)	29(65.9%)	147(47.9%)	0.01
*Lymphocytes <sup>c</sup>	130(49.4%)	27(61.4%)	157(51.1%)	0.14
*Neutrophils <sup>c</sup>	127(49.0%)	22(52.4%)	149(49.5%)	0.69
*Monocytes <sup>c</sup>	129(49.0%)	28(63.6%)	157(51.1%)	0.07
*Basophils <sup>c</sup>	187(71.7%)	36(81.8%)	223(72.6%)	0.14
*Eosinophils <sup>c</sup>	118(44.9%)	34(77.3%)	152(49.5%)	<0.01

Abbreviation: MRSA= Methicillin Resistant Staphylococcus Aureus.

Values are n (%) unless otherwise noted.

<sup>c</sup>Data in the onset of Staphylococcus aureus Bloodstream Infection.

\*Parameters dichotomised at the median. Values are n (%) unless otherwise noted.

The numbers were patients' numbers with leukocytes, haemoglobin, platelets, neutrophils, lymphocytes, monocytes, basophils or eosinophils under the median, and patients' numbers with serum creatinine total bilirubin above the median.

**Table 2**

**Cox proportional hazards regression analysis for mortality in  
Staphylococcus aureus bloodstream infection**

Variable	Univariate HR(95%CI)	P value	Multivariate HR(95%CI)	P value
Male gender	0.54(0.30-0.97)	0.04	0.43(0.22-0.81)	0.01
Age, +10 years	1.11(0.94-1.31)	0.24	1.00(0.83-1.18)	0.91
Neutropenia <sup>a</sup>	2.43(0.87-6.80)	0.09		
With previous hospitalization in the preceding 90 days	1.80(1.00-3.30)	0.06		
Antibiotics therapy	2.32(1.23-4.38)	0.01		
Mechanical ventilation <sup>b</sup>	1.92(1.02-3.62)	0.04		
Indwelling central venous catheter <sup>b</sup>	2.33(1.27-4.28)	<0.01		
Indwelling nasogastric tube <sup>b</sup>	2.28(1.26-4.12)	<0.01		
Indwelling urinary catheter <sup>b</sup>	2.35(1.30-4.25)	<0.01		
APACHE II score, +1 score	1.11(1.07-1.15)	<0.01		
Pitt score, +1 score	1.22(1.11-1.34)	<0.01		
Charlson score, +1 score	1.18(1.04-1.35)	0.01		
Drug-resistance bacteria	3.61(1.42-9.17)	<0.01		
Inappropriate empiric antimicrobial therapy	2.96(1.55-5.66)	<0.01	4.01(2.04-7.92)	<0.01
Total bilirubin	2.32(1.22-4.44)	0.01	2.20(1.13-4.29)	0.02
Serum creatinine	1.75(0.95-3.24)	0.07	2.08(1.07-4.02)	0.03
Monocytes <sup>c</sup>	1.74(0.94-3.21)	0.08		
Eosinophils <sup>c</sup>	3.98(1.97-8.06)	<0.01	2.84(1.36-5.91)	<0.01

<sup>a</sup>Within 30 days preceding infection onset.

<sup>b</sup>Within 48 hours preceding infection onset.

<sup>c</sup>Data in the onset of infection.

**Table 3**

**Cox proportional hazards regression analysis for mortality in  
Staphylococcus aureus bloodstream infection developed after 48  
hours in hospital**

<b>Variable</b>	<b>Univariate HR(95%CI)</b>	<b>P value</b>	<b>Multivariate HR(95%CI)</b>	<b>P value</b>
Male gender	0.58(0.29-1.15)	0.12	0.55(0.27-1.13)	0.10
Age, +10 years	1.08(0.88-1.32)	0.46	1.06(0.85-1.33)	0.61
Antibiotics therapy	1.92(0.92-4.02)	0.09		
Mechanical ventilation <sup>a</sup>	1.82(0.91-3.67)	0.09		
Indwelling central venous catheter <sup>a</sup>	2.14(1.02-4.49)	0.05		
Indwelling nasogastric tube <sup>a</sup>	2.20(1.11-4.36)	0.02		
Indwelling urinary catheter <sup>a</sup>	2.26(1.11-4.59)	0.02		
Chronic cardiac failure	2.46(1.11-5.46)	0.03		
Liver cirrhosis	2.39(1.00-5.80)	0.05	3.17(1.23-8.14)	0.02
Charlson score, +1 score	1.21(1.04-1.39)	0.01		
APACHE II score, +1 score	1.12(1.07-1.17)	<0.01		
Pitt score, +1 score	1.22(1.10-1.36)	<0.01		
Serum creatinine	2.29(1.09-4.81)	0.03	2.56(1.18-5.56)	0.02
Total bilirubin	2.24(1.04-4.82)	0.04		
Eosinophils <sup>b</sup>	3.02(1.43-6.34)	<0.01	3.20(1.23-8.14)	<0.01

<sup>a</sup>Within 48 hours preceding infection onset.

<sup>b</sup>Data in the onset of infection.

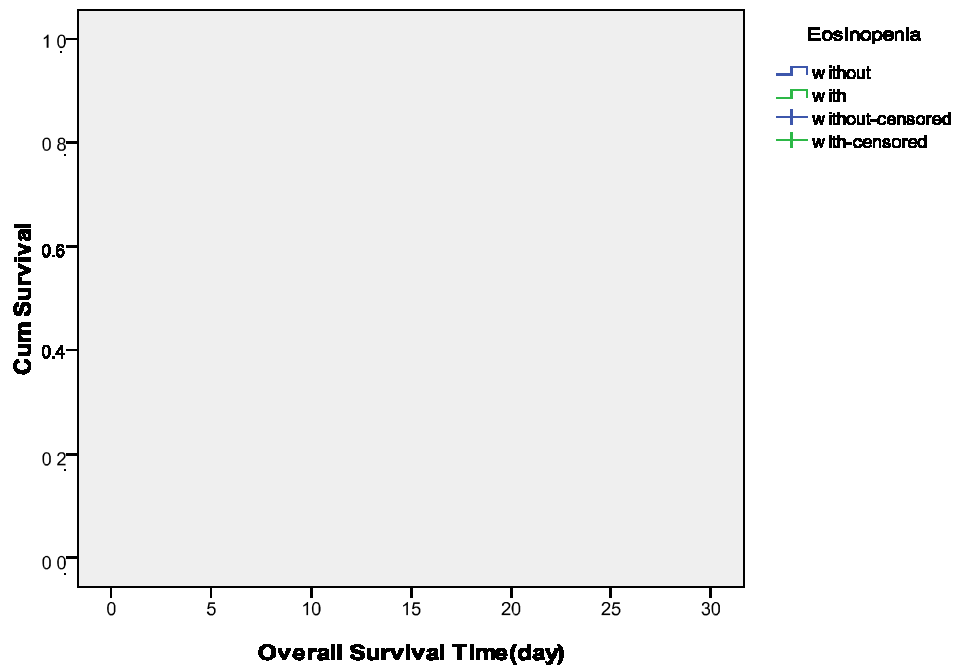


Fig.1. Survival curve for patients with eosinopenia and patients without eosinopenia(log rank test,  $P<0.01$ ). The blue solid line indicates the survival curve for patients without eosinopenia, and the green dotted line indicates the survival curve for patients with eosinopenia.



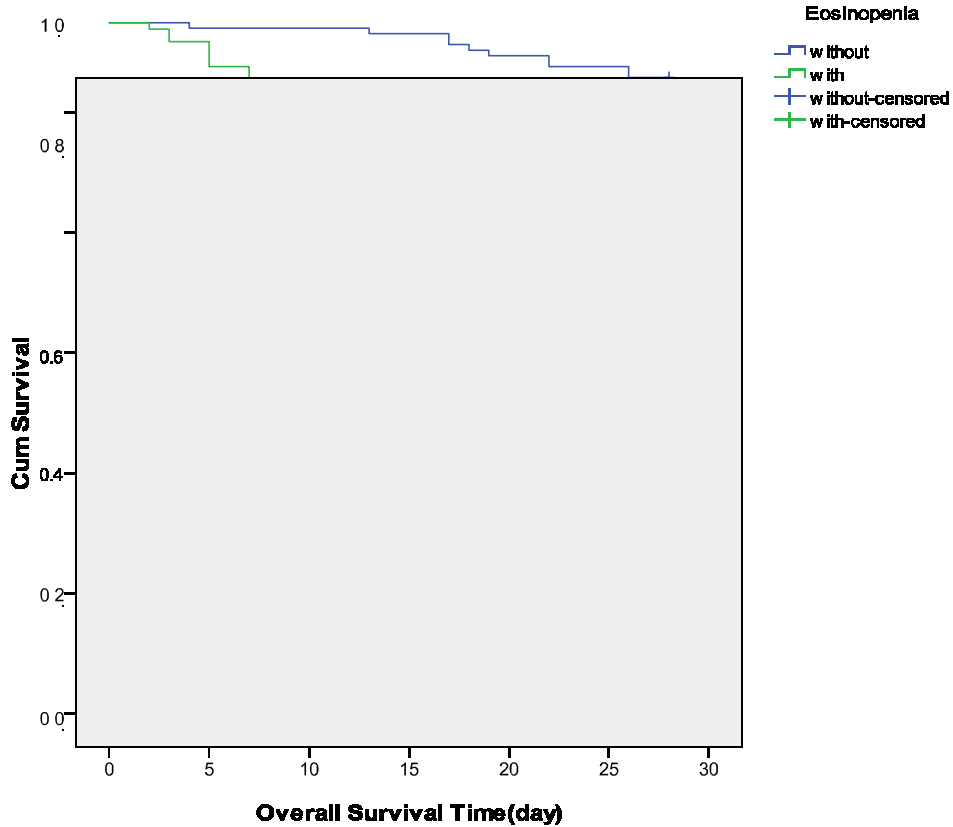


Fig.2. For patients who developed *Staphylococcus aureus* bloodstream infection after 48 hours in hospital, survival curve for patients with eosinopenia and patients without eosinopenia(log rank test,  $P<0.01$ ). The blue solid line indicates the survival curve for patients without eosinopenia, and the green dotted line indicates the survival curve for patients with eosinopenia.