

1 **Impact of indole-3-butyric acid (IBA) on the root induction of *Arbutus pavarii***  
2 **Pamp (Strawberry tree) culture in vitro**

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13 **ABSTRACT**

14 The main objective of this study was to clarify the best concentration of the indole-3-butyric acid (IBA)  
15 in order to induce the formation of strong roots of the *Arbutus pavarii* plant. It is an endangered plant  
16 in El-Jabel El-Akhdar- Libya. This study was created to find a protocol for the propagation of *Arbutus*  
17 *pavarii* in vitro. The present paper aimed to explore the role of different concentrations of IBA plant  
18 growth regulator on the rooting. *In vitro* germination three weeks old seedlings were transferred to MS  
19 roots induction medium which supplemented with different concentrations of IBA ( 0, 1, 1.5 and 2 mg  
20 L<sup>-1</sup>). the highest response was obtained when transfer the seedlings to the MS medium half strength and  
21 supplemented with IBA (0.1 mg L<sup>-1</sup> ) . All the growth indicators [rooting percentage (77%) number,  
22 length (7.4 cm) and dry weight (0.0449 g/jar)] that were measured significantly enhanced when using  
23 concentration 0.1 mg L<sup>-1</sup> of IBA.

24 Key words: *Arbutus pavarii*; Germination; Indole-3-butyric acid; Roots dry weight and Sterilization.

25 **INTRODUCTION**

26 *Arbutus pavarii* spreads naturally in El-Jabel El-Akhdar region in the north-eastern part of Libya,  
27 between latitudes 20° 23° east and latitude 32° 33° north [1]. Its presence is concentrated in the  
28 northern and central regions, in the valleys, slopes, mountain slopes and lands with shallow or rocky  
29 calcareous soils. *Arbutus* has about 20 species of trees and has different local names in its spread areas  
30 [2]. Endemic species from around 4% from the total species of Libyan flora, *A. pavarii* Pamp.  
31 (Ericaceae) locally known as “Shmeri” is one of the endemic species in El-Jabel El-Akhdar. IT is  
32 shrubs grows in the El-Jabel El-Akhdar region, mixed with many trees and shrubs such as *Pistacia*  
33 *lentiscus*, *Ceratonia siliqua* L, *Juniperus phoenicea* L, *Quercus coccifera*, *Rhus tripartita*  
34 (Ucria)Grande, *Phillyea media* L., *Ziziphus lotus*(L)Desf,[3 ; 4]. These trees and shrubs are the main  
35 species in the Maquis formation in -Jabel El-Akhdar region, in the form of shrubs dense of branches  
36 which is a small tree or a large evergreen shrub with a smooth reddish brown bark and a multi-  
37 patterned leaves with Serrate or Entire edge. Flowering occurs in late spring and fruits mature in late  
38 summer. The fruit is globose, a many-seeded berry, yellow to orange in colour, turning  
39 red when fully maturity [5]. This plant suffers from increasing degradation due to negative  
40 human activities in many of the El-Jabel El-Akhdar areas and agricultural expansion, urbanization,  
41 overgrazing, charcoal making In addition to the climatic factors of the fluctuation of rainfall and  
42 temperature fluctuations which are accompanied by dry southern winds caused by physiological

43 diseases due to drought, and whereas *A. pavarii* has never been widely cultivated by afforestation of  
44 other species.

45 The need for the continuous improvement of traits in crop species remains an ongoing effort for crop  
46 scientists and farmers. Different plant species have their own set of phenotypes that need to be  
47 improved in order to both add nutritional values and enhance economic gains for humankind. The  
48 increase in food demand worldwide, associated with unequal distribution, and the disequilibrium in the  
49 distribution of wealth has caused an increasingly important pressure on food producers who, in parallel,  
50 have increased their requirements for new technologies that allow greater yields and better quality of  
51 the products that they offer [5]. While at the same time, there has been an increasing consumer led  
52 demand for lower environmental damage and greater sustainability in the food production chain.

53 Strawberry tree is propagated by runners; therefore the health of daughter plant depends on their  
54 mother plants. Strawberry is affected by numerous viruses that greatly reduce the yield [6].

55 Plant tissue culture (PTC) is a set of techniques for the aseptic culture of cells, tissues, organs and their  
56 components like genes and enzymes under defined physical and chemical conditions *in vitro* and  
57 controlled environment. PTC technology also explores conditions that promote cell division and  
58 genetic re-programming in *in vitro* conditions and it is considered an important tool in both basic and  
59 applied studies, as well as in commercial application [7]. PTC techniques have become of major  
60 industrial importance in the area of plant propagation, disease elimination, plant improvement, and  
61 production of secondary metabolites.

62 Growth regulators constitute one of the key and more expensive elements used for *in vitro* propagation.  
63 That is why; they must be optimized or substituted for more efficient and cheaper bio regulators [8].  
64 Indole-3-butyric acid (IBA) is a plant hormone in the auxin family and is an ingredient in many  
65 commercial horticultural plant rooting products.. In plant tissue culture IBA is used to initiate root  
66 formation *in vitro* in a procedure called micropropagation. Micropropagation of plants. the effect of  
67 three different auxins, IBA, IAA and NAA were examined to determine the relative effect of each  
68 auxin on root formation. According to the result for the species, IBA was shown to produce a higher  
69 yield of roots compared to the other auxins.[9] The effect of IBA is in concurrence with other studies  
70 where IBA is the most commonly used auxin for root formation.[10]

71 Therefore, the aim of this study was find a protocol for the propagation *A. pavarii* Pamp and try seed  
72 germination using different concentrations of IBA to obtain seedlings to enable them to re-plant them  
73 in their natural environment.

## 74 MATERIAL AND METHODS

### 75 MATERIAL

76 Fresh Seed *A. pavarii* were collected in December month from the outskirts of the city of Al Bayda –  
77 Libya. Taxonomist at the Department of Botany Herbarium, Faculty of Science, and Omar Al-Mukhtar  
78 University further identified the samples.

### 79 METHODS

80 Preparation of culture media

81 Half basal of Murashige and Skoog 1962 (MS) [11] salts nutrient medium with vitamins, glycine and  
82 supplemented with 30 g L<sup>-1</sup> sucrose, 0.1 g L<sup>-1</sup> myo- inositol (Table 1) was used for *in vitro* seedlings  
83 germination. While full MS nutrient salt medium was used for rooting media. After supplementation  
84 of full MS media with different concentrations (0, 1, 1.5 and 2 mg L<sup>-1</sup>) of indole-3-butyric acid (IBA)  
85 plant growth regulators [for rooting], all cultures pH were adjusted to 5.8 with 1N KOH or 1N HCl,  
86 then with 7 g L<sup>-1</sup> agar prior to autoclaving at 121°C and 1.2 kg cm<sup>-2</sup> for 20 minutes. Culture medium  
87 was dispensed as 50 ml per jar (350 ml) for *in vitro* seedlings germination, and rooting. All types of  
88 culture media were kept for three days under completely darkness for test of contamination.

89 Tissue culture chemicals MS medium and growth regulator indole-3-butyric acid were purchased from  
90 Sigma company.

#### 91 Sterilization and germination

92 Seeds of the *A. pavarii* were washed with running tap water for 30 min. Then they were taken to the  
93 laminar air flow cabinet in which they surface sterilized by dipping in 70% (v/v) ethanol for 2 min and  
94 rinsed with sterilized distilled water. Further, disinfected with 20 % (v/v) of commercial Clorox (5.25%  
95 Cl<sub>2</sub>) solution for 15min (Rabha [12]) and rinsed three times with sterilized distilled water. In complete  
96 aseptic conditions equal number from sterilized seeds represents were inoculated in culture medium  
97 aseptically as six seeds per each. Cultures were maintained under normal condition (16/8 hours  
98 light/dark) at 1500 lux using cool white fluorescent lamps and incubated in a controlled growth  
99 chamber at 26±1°C.

100 This experiment was carried out to study the effect of indole-3-butyric acid capacity to enhance rooting  
101 on seedling derived *in vitro*. *In vitro* germination three weeks old seedlings (reached about 5 - 6 cm in  
102 height were subjected as a plant materials) which resulted from MS free growth regulators were  
103 transferred to MS roots induction medium (R) which supplemented with different concentrations of  
104 IBA( 0, 1, 1.5 and 2 mg L<sup>-1</sup>) as follow:

R<sub>0</sub>= control ( MS free growth regulators )  
R<sub>1</sub> = MS + 1 mg L<sup>-1</sup> IBA  
R<sub>2</sub>= MS + 1.5 mg L<sup>-1</sup> IBA  
R<sub>3</sub>= MS + 2 mg L<sup>-1</sup> IBA

105 Each treatment was consisted of 6 replicates (jar) and each replicate contained three seedling. Cultures  
106 were incubated in a controlled growth chamber in complete darkness for 3 days at 26±1°C then  
107 transferred to normal condition. After 4 weeks from incubation, the number, length (cm) and dry  
108 weight (g/jar) for root were recorded.

#### 109 Statistical Analysis

110 The data were subjected to two-way analysis of variance (ANOVA) and Duncan's Multiple Range  
111 Test (DMRT) at the 5% level using Microsoft Excel software.

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123 Table 1: Culture media composition (Murashige and Skoog 1962).

Ingredients	Amount (mg/L)
Macronutrients	
NH <sub>4</sub> NO <sub>3</sub>	1650.00
KNO <sub>3</sub>	1900.00
CaCl <sub>2</sub> .2H <sub>2</sub> O	440.00
MgSO <sub>4</sub> . 7H <sub>2</sub> O	370.00
KH <sub>2</sub> PO <sub>4</sub>	170.00
Micronutrients	
KI	0.83
H <sub>3</sub> BO <sub>3</sub>	6.20
MnSO <sub>4</sub> .4H <sub>2</sub> O	22.30
ZnSO <sub>4</sub> .7H <sub>2</sub> O	8.60
Na <sub>2</sub> MoO <sub>4</sub> .2H <sub>2</sub> O	0.25
CuSO <sub>4</sub> .5H <sub>2</sub> O	0.025
CoCl <sub>2</sub>	0.025
Iron stock	
FeSO <sub>4</sub> .7H <sub>2</sub> O	27.80
Na <sub>2</sub> .EDTA.2H <sub>2</sub> O	37.30
Vitamins	
Myo-inositol	100.00
Nicotinic acid	1.00
Pyridoxine HCl	1.00
Thiamine HCl	10.00
Glycine	2.00
Sucrose (g)	30.00
Agar (g)	8.00

124 RESULT AND DISCUSSIONS

125 This experiment was conducted to provide the *in vitro* growing seedlings needed for present study as  
 126 explants source. Indole-3-butyric acid in different concentrations was used to study its effect on the  
 127 roots. The effect of MS medium supplemented with different concentrations of IBA (1, 1.5 and 2 mg L<sup>-1</sup>)  
 128 <sup>1</sup>) on rooting frequency (%), number of roots, root length (cm) and roots dry weight (g/jar) of *Arbutus*

129 seedlings was investigated. Data tabulated in Table 2 clearly showed that MS medium supplemented  
 130 with 1 mg L<sup>-1</sup> IBA (R<sub>1</sub>) gave the maximum value for rooting percentage (70%). In addition, there were  
 131 non-significant differences among R<sub>0</sub>, R<sub>2</sub>, and R<sub>3</sub> and they recorded the minimum value for rooting  
 132 percentage (55%). About number of roots and concerning to seedling *Arbutus*; there were clear  
 133 differences between the treatments used in the study. It was found that MS medium fortified with 1.5  
 134 mg L<sup>-1</sup> IBA(R<sub>2</sub>) in the best number of roots as the number of five is the highest compared to the  
 135 control. On the other hand, the data are also shown in the table 2 the longest root (7.4 cm) recorded  
 136 with R<sub>1</sub> followed by R<sub>2</sub> medium, which recorded 5.4 cm length. Furthermore, there were non-  
 137 significant differences among R<sub>0</sub>, and R<sub>3</sub> media they were shorter in length. The highest dry (0.09 g/jar)  
 138 weight increment was scored on R<sub>1</sub> (0.0449) and R<sub>2</sub> media compared to other used media.

139 There have been many studies devoted to the cultivation of strawberry trees and the exact propagation  
 140 has been reported about 30 years ago [13]. In recent years, many research groups have been involved in  
 141 establishing reliable regeneration protocols for agronomical important *A. pavarii*, because it would be a  
 142 primary step to facilitate gene introduction and improvement of the crop. Our aim in the study was to  
 143 investigate the effect of IBA hormone on root induction in vitro. Through our study of the effect of  
 144 IBA on the root induction we found that it significantly enhanced the number and length and dry  
 145 weight in strawberry seedling. In vitro plant regeneration of Strawberry from different parts, has been  
 146 reported by seeds, leaves, petioles [14], stem [15], stipules [16], and roots [17]. The results in Table 2  
 147 showed that the growth and formation of roots were very low in the treatment of control ( MS free  
 148 growth regulators ) compared to all other treatments. These findings are somewhat similar to those  
 149 previously reported by Ashraf [18] .Regarding the effect of IBA on the root response, the results  
 150 indicated that the IBA with (1 and 1.5 mg/L) showed the highest roots response compared to all other  
 151 treatments. These results do not exactly match what he concluded [19]. Our results agreed with Gautam  
 152 [20] indicated that the highest root induction frequency obtained was 95.23% on MS medium with IBA  
 153 at 1.0 mg/l . Mereti [21] found that the highest percentages of rooting were achieved in MS medium  
 154 contained 10 µM IBA (92%) and 10 µM IAA (82%). Additionally, by increasing the concentration of  
 155 IBA the height of root was decreased. Haddadi et al [22] reported that the presence of NAA strength  
 156 the rooting percentage and root number but the medium without any Auxin had the lower number of  
 157 root. However, the highest root development was observed in the control treatment. Here it was  
 158 concluded that the root phenotype (number and length) was diverse as influenced by different Auxin  
 159 treatments. All different concentrations of IBA ( 0, 1, 1.5 and 2 mg L<sup>-1</sup>) induced the root induction in  
 160 strawberry and significantly differences were observed among treatments in number and length of  
 161 regenerated seedling.

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164 Table 2: Effect of culture media composition with different concentrations of IBA on rooting  
 165 percentage number of roots, root length and root dry weight of *A. pavarii* after 4 weeks of culturing  
 166 and incubation at normal condition.

Parameters	rooting	No. of	Root length	Dry weight
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Treatments	percentage (%)	roots	( cm)	(g/jar)
R <sub>0</sub> = control ( MS free growth regulators )	55 <sup>b</sup>	2 <sup>c</sup>	3.2 <sup>c</sup>	0.008 <sup>d</sup>
R <sub>1</sub> = MS + 1 mg L <sup>-1</sup> IBA	70 <sup>a</sup>	3 <sup>b</sup>	7.4 <sup>a</sup>	0.0449 <sup>a</sup>
R <sub>2</sub> = MS + 1.5 mg L <sup>-1</sup> IBA	55 <sup>b</sup>	5 <sup>a</sup>	5.4 <sup>b</sup>	0.0303 <sup>b</sup>
R <sub>3</sub> = MS + 2 mg L <sup>-1</sup> IBA	55 <sup>b</sup>	3 <sup>b</sup>	3.2 <sup>c</sup>	0.0183 <sup>c</sup>

167 Means having the same letters in a column were not significantly different at p<0.05

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## 171 CONCLUSION

172 In vitro regeneration of Strawberry tree (*Arbutus pavarii*) is  
 173 a requirement for genetic transformation, which involves induction and development to the whole  
 174 plant. Several studies have shown that IBA the effects of root induction in plants. That's what agreed  
 175 with our results in this study demonstrated that IBA (1mg L<sup>-1</sup>) was an effective concentration among  
 176 different other concentrations on root induction.

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