

## **Original Research Article**

### **Identification of *in-vitro* PEG mediated drought tolerance genotype in rice (*Oryza sativa* L.)**

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#### **ABSTRACT**

Drought is a serious bottleneck in the production of rice globally. For this, an experiment was conducted *in-vitro* on six rice genotypes viz. BRRI dhan-28, Begunbahar, Burikatari, Pashpai, Dular and Begunbichi to investigate the effect polyethylene glycol (PEG) mediated artificial drought on morpho-physiological parameters such as germination percentage, shoot length, root length, fresh weight, dry weight, turgid weight, relative water content, and proline accumulation. Here, different doses of PEG-6000 viz. 0 gL<sup>-1</sup>, 15gL<sup>-1</sup>, 30gL<sup>-1</sup>, 45gL<sup>-1</sup> and 60gL<sup>-1</sup> were used with Murashige and Skoog (MS) medium. The results demonstrated that BRRI Dhan-28, Burikatari and Dular revealed greater performance at control conditions but at the highest degree of water stress conditions only Burikatari showed higher mean value for all parameters studied. Again, Begunbichi followed by BRRI Dhan-28 exhibited the lowest mean value for almost all traits except for proline accumulation. Here, water stress decreased performance of morpho-physiological characters except proline accumulation in rice. The cluster analysis was performed and distributed into three groups where there was a significant variation among the clusters at different water stress conditions. Here, the genotype Burikatari is more diverse giving maximum Euclidian distances in drought treatments. It could be considered as a parent in the hybridization program against Begunbahar, Dular and Paspai. Therefore, considering the mean performances and cluster analysis, Burikatari exhibited greater performances against the highest degree of

drought conditions. This genotype may bear drought-tolerant gene for which could be utilized for further development of drought tolerant variety and gene transfer.

**Key words:** *In vitro*, water stress, PEG (Polyethylene Glycol), rice.

## 1. INTRODUCTION

Rice has been referred to as “Global Grain” and considered as a model cereal crop in the world [1]. It belongs to genus *Oryza* that contains 25 recognized species, of which 23 are wild species and two; *O. sativa* and *O. glaberrima* are cultivated [2]. *O. sativa* is the most widely grown cultivated species. In the year 2017-18, the rice production around the world was estimated at 484.7 million tons and was 0.5 % below the year-earlier record [3] that was cultivated at least 114 countries. Asia is the leader in rice production accounting for about 90% of the world's production where about 75% of rice is consumed by the Asian people [4,5].

Rising temperature as a form of climate change and altered soil moisture is projected to decrease the yield of food crops over the next 50 years [6]. In recent years, drought and salt stress reduces rice production worldwide [7,8]. However, depends on crop growth period and stress intensity, drought reduces the yields by 15 to 50 percent [9]. It has been reported that main constrain to crop yield ~~is for the~~ precarious rainfall or scarcity of the water in soil deep layer due to presence of hardpan that resists accessing water [10, 11]. Rice is one of the most sensitive cultivated species to water stress. So, the farmer is more likely to access ~~the plant breeding brought~~ tolerant genotypes rather than expensive agronomic practices [12].

Plant responses to drought ~~involve and engaging lots of~~ physiological, biochemical and molecular changes [13]. So, it seems impossible to increase crop yield ~~remain in~~ water-deficient condition during crop cultivation [14]. Morphological characters viz., shoot and root length [15],

leaf fresh, turgid, dry weight and relative water content [16] and seed germination and seedling growth [17] and biochemical: proline accumulation [16] are adversely affected by water stress.

*In vitro*, PEG (polyethylene glycol) are known to cause osmotic stress which alters the osmotic potential of the cell and hence these will be as useful selection agents for drought tolerance. It resists both water and mineral upliftment by [root by](#) forming hydrogen bond with water and also decrease the water potential in the culture medium ~~by root~~. Thus, the osmotic agent acts in lowering the water potential in a way similar to soil drying [18,19].

Cluster analysis based on Mahalanobis  $D^2$  statistic [20] is the possible quantifier for amounting the degree of genetic variability among the genotypes. The numerous cluster group demonstrated the highest degree of variability present in the materials evaluated. Earlier workers had also reported the presence of substantial genetic diversity in rice [21,22].

Considering the above aspects, one of the best policies would be the development of water stress tolerance genotypes to increase the rice yield in drought-prone area. So, the research hypothesis might be the identification of potential drought tolerance genotypes. This might be achieved through the genetic study of morphophysiological and biochemical traits *in- vitro* conditions using PEG treatment (under drought stress conditions). Therefore, the major objectives of the present research work was (i) to evaluate the result of drought-induced seed germination and seedling growth parameter of rice genotypes, (ii) to mold a quick and effective strategy for rice against drought conditions and (iii) to determine the most drought tolerant the genotype of rice.

## 2. MATERIALS AND METHODS

**Plant materials:** Seeds of six rice genotypes comprised of drought-tolerant landraces (Burikadari, Begunbahar, Dular, Pashpai, Begunbichi) and an elite cultivar BRRI Dhan- 28 were used in the present investigation. These materials were collected from the Genetic Resources and Seed Division of Bangladesh Rice Research Institute (BRRI), Gazipur, Bangladesh.

**Experimental set-up:** During the period of 2017 to December 2017 the experiment was carried out at tissue culture Laboratory of the Genetics and Plant Breeding and Chemistry Laboratory of the Hajee Mohammad Danesh Science Technology University (HSTU) Dinajpur, Bangladesh. Here we used [23] Murashige and Skoog, media for culturing the seeds. Firstly, seeds were sterilized in mercuric chloride for five minutes then sterilized in 70 % ethyl alcohol for three minutes and washed with double distilled water. Afterward, sterilized matured seeds were inoculated into the test tube containing 10ml MS solution with PEG-6000 at the different concentrations such as  $T_0 = 0\text{g L}^{-1}$  (Control condition),  $T_1 = 15\text{g L}^{-1}$ ,  $T_2 = 30\text{g L}^{-1}$ ,  $T_3 = 45\text{g L}^{-1}$ ,  $T_4 = 60\text{g L}^{-1}$  with four replications and lab temperature was controlled at  $25^\circ\text{C}$  with under the correspondence of sixteen hours light period and eight hours dark period.

**Data collection:** The following data were recorded during the experimental period. The procedures of measurement of these data are described here below.

**Percentage of germination:**

$$\text{Percentage of germination} = \frac{\text{Number of seeds germinated}}{\text{Number of seeds inoculum}} \times 100$$

**Shoot length, root length, fresh weight, dry weight, turgid weight of plant, and shoot-root ratio:**

**Comment [S1]:** Mention month

**Comment [S2]:** How many seeds per test tube

**Comment [S3]:** Mention day after inoculation when percent germination was measured.

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**Comment [S4]:** shoot to root ratio data was not found in results section

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Seedling was taken out with the help of forceps at the 17<sup>th</sup> days of inoculation and shoot length, root length and total length were measured in cm by a graduated scale. And also, the fresh, turgid and dry weight was calculated in gram by electrical balance.

**Comment [S5]:** Mention, how plants were dried and how turgid weights were taken.

**Relative water content of leaf:** The relative water content [24] of leaf was determined as follows:

$$\text{Relative water content} = \frac{\text{Fresh weight} - \text{dried weight}}{\text{Fully turgid weight} - \text{dried weight}} \times 100$$

Leaf fresh weight was taken, then leaves were submersed into distilled water in the darkness at 4° C to minimize the respiration losses until they reached the constant weight that is 12 hours, and weighed as turgid weight. After leaf was dried the leaves in the oven for 48 hours at 70° C, dry weight was taken in gram.

**Comment [S6]:** You do not need to write replications as four replications you have already given in experimental set up.

**Determination** Four times weight were obtained for each treatment.

**of pProline content of leafdetermination:** Sulfosalicylic acid was used for the extracted of to extract proline from the leaves, and the filtrated solution was mixed with an equal amount of ninhydrin reagent and glacial acetic acid that is 1.25g ninhydrin, 20ml 6NH<sub>3</sub>PO<sub>4</sub> and 30ml glacial acetic acid and incubated at 100° C for 1hour. All test tubes placed in cold water to cool the sample and toluene mixed with it and vigorously shaken it for complete mixing completely.

**Comment [S7]:** Write concentration of sulfosalicylic acid used in the protocol.

Color was read at 520nm light was passed through the sample using Pharmacia LKB-Novaspaces spectrophotometer to determine the light absorption of toluene. Standard curved was used to measure the concentration of the proline that was expressed as mg/100g of plant parts [25].

**Comment [S8]:** How much toluene? Mention it. Please also check protocol thoroughly for its correctness.

**Cluster analysis:** The statistical software – Agricultural Research (STAR) Version 2.0.1 (2014) was used for estimating of Euclidian distance of coefficients. Euclidean distance matrix

generated from seedling data was used as input data for cluster analysis based on the un-weighted pair-group method of arithmetic average (UPGMA). To estimate the level of relatedness among the genotypes based on the Euclidean genetic distances a UPGMA was drawn.

### 3. RESULTS AND DISCUSSION

The experiment was conducted to analyze morpho-physiological traits of six rice genotypes under control and different water stress conditions. Mean performance and other analyses were done on eight morpho-physiological traits like shoot length, root length, fresh weight, turgid weight, dry weight, relative water content, and germination percentage. The results have been presented and discussed under the following headings:

The analysis of variance for different yield and morpho-physiological characters: The analysis of variance was accomplished to assess the variability pertained for a character among the six rice genotypes (Table 1). For all traits, the analysis of variance showed a significant difference among the genotypes indicated that the genotypes have inherent genetic variation among themselves with respect to the characters studied. The treatment effect was also highly significant for all the studied traits which was also reported earlier. [26,27]. Interaction between genotype and treatment showed highly significant differences for all most all of the traits and the mean squares against the replications found significant for all the characters.

#### ***Genotype x treatment interactions on morpho-physiological characters on rice genotypes:***

The interaction effect of \_genotypes and treatments on eight morpho-physiological traits are presented in Table 2. Significant variations were observed in the different treatments on shoot length, root length, fresh weight, turgid weight, relative water content and percentage of seed germination. Shoot length varied from 7.40 to 21.55cm. Maximum shoot length was recorded in

BRRi dhan-28 (21.55cm), when the seed was inoculated in MS medium supplement with 0 gL<sup>-1</sup> PEG (Figure 1(a)). The lowest shoot length was found in Begunbichi (7.40 cm) followed by ~~BRRi dhan-28~~Pashpai (8.275 cm) when those were treated with 60 gL<sup>-1</sup> PEG (Table 2, Figure 1(b)). Therefore, the interaction effect of varieties and treatments were highly significant for ~~root~~ shoot length. The maximum value of root length was found in BRRi dhan-28 (6.80 cm) followed by Dular Burikatari and the lowest was recorded in Begunbichi (2.28 cm). For drought effect, shoot length, root length, for most of the plants were decreased compared to the control conditions which is a common adverse effect of drought that was similar to previous study [14]. However, reduction of shoot length, root may ~~be occurred due to implementation of~~also occur due to decreased cell division under the stress condition. Moreover, the relative water content was calculated from fresh weight, turgid weight and dry weight, which was varied from 3.82% to 16.75%. Maximum value of relative water content 16.75% was found in Dular followed by Burikatari 15.64% when seeds were inoculated on MS medium with 0gL<sup>-1</sup> PEG and lowest water content was found in Begunbahar 3.82% followed by Begunbichi, BRRi dhan-28 with 5.05% and 5.36% respectively with 60 gL<sup>-1</sup> PEG. The interaction effect of genotypes and treatments differ significantly on relative water content. However, the relative proline varied from

**Table 1. Mean squares (MS) derived from CRD (two Factor) model on morphophysiological and biochemical characters in rice.**

| Characters   | Source of variation with mean square |                    |                      |                                   |                 |
|--------------|--------------------------------------|--------------------|----------------------|-----------------------------------|-----------------|
|              | Genotype<br>(5df)                    | Treatment<br>(4df) | Replication<br>(3df) | Genotype ×<br>Treatment<br>(20df) | Error<br>(87df) |
| Shoot Length | 81.260***                            | 320.27***          | 0.360***             | 5.820***                          | 0.830***        |

|                        |             |             |          |          |          |
|------------------------|-------------|-------------|----------|----------|----------|
| Root Length            | 6.340***    | 18.428***   | 0.253*** | 0.568*** | 0.127*** |
| Fresh Weight           | 0.002***    | 0.005***    | 0.001    | 0.001*** | 0.001    |
| Turgid Weight          | 0.005***    | 0.015***    | 0.001*** | 0.001*   | 0.001*** |
| Dry Weight             | 0.002***    | 0.005***    | 0.001    | 0.001*** | 0.001    |
| Relative Water Content | 60.545***   | 286.488***  | 0.569    | 3.156*** | 0.695    |
| Germination            | 2465.000*** | 4047.300*** | 0.800*** | 932.800* | 0.600*** |
| Proline Content        | 6.710***    | 327.710***  | 0.150*** | 0.57***  | 0.050*** |

Here, \* and \*\*\* indicates significant at 5% and 0.1% levels of probability, respectively and df indicates degrees of freedom.

3.89 mg to 15.28mg. Maximum proline content was observed in BRRI-28 (15.28 mg) with 60g L<sup>-1</sup> of PEG and lowest proline content was observed in Dular (3.89 mg) followed by Begunbichi (3.98 mg) with 0 gL<sup>-1</sup> PEG supplement. It was observed that a significant increase of proline content with the increasing of water stress conditions [28,29]. The highest proline content was found [16] at the 9% PEG supplement on MS medium. The germination percentage was varied 25% to 99.75% with a different concentration level of PEG. The germination percentage showed by Burikateri with 0g L<sup>-1</sup> PEG and the lowest germination percentage was found in Begunbichi followed by BRRI Dhan-28 with 60g L<sup>-1</sup> PEG. According to previous studies [30] water stress decreased the germination percentage.

Cluster analysis: Cluster analysis showed the significances difference among the rice for the rice genotypes that reveled the variability among the genotypes. Cluster analysis was performed for 0g L<sup>-1</sup> PEG (control), 15g L<sup>-1</sup> PEG, 30g L<sup>-1</sup> PEG, 45g L<sup>-1</sup> PEG and 60g L<sup>-1</sup> PEG treatment and Euclidian distance of coefficients were studied for all rice genotypes based on all traits. Dendrogram from UPGMA clustering indicated the grouping of six genotypes of rice into three

clusters. In control conditions ( $0\text{gL}^{-1}$  PEG), Cluster I, II and III, comprised of 2, 1 and 3 genotypes, respectively (Figure 2). Among the three clusters, cluster number II revealed the highest distance by the genotype BRRI Dhan 28 and the lowest distance was exhibited by the cluster III with genotypes Begunbahar, Paspai and Begunbichi. But with the increasing of water stress (increasing PEG amount in MS medium), the cluster arrangement becomes changed. Here, BRRI Dhan 28 revealed the highest distance in ~~cluster 2 of~~  $30\text{gL}^{-1}$  PEG treatment. ~~Moreover, this genotype revealed~~ but moderate distance in  $45\text{gL}^{-1}$  PEG conditions ~~and but~~ lower distance in  $15\text{gL}^{-1}$  PEG and  $60\text{gL}^{-1}$  PEG conditions. Again, the genotypes Begunbahar, Dular and Paspai revealed lowest euclidian distance under all treatment conditions. In contrast, Burikatari exhibited the highest distance in treatments  $15\text{gL}^{-1}$ ,  $45\text{gL}^{-1}$  and  $60\text{gL}^{-1}$  PEG conditions and moderate in  $30\text{gL}^{-1}$  PEG conditions that was similar to previous study. [31]. Therefore, this genotype is more diverse and could be considered as a parent against the genotypes Begunbahar, Dular and Paspai in hybridization program.

**Table 2. Interaction effect of genotypes x treatments on eight morphological and physiological traits in six rice genotypes.**

| Genotype              | Treatment combination | Shoot length (cm) | Root length (cm) | Fresh weight of the plant(gm) | Turgid weight of the plant (g) | Dry weight of the plant (g) | Relative Water Content % of leaf | Proline Content of leaf (gm/100g) | Germination % |
|-----------------------|-----------------------|-------------------|------------------|-------------------------------|--------------------------------|-----------------------------|----------------------------------|-----------------------------------|---------------|
| V1                    | V1T0                  | 21.55A            | 6.80A            | 60.75G                        | 112.25C                        | 53.2H-J                     | 12.68D-F                         | 4.65q                             | 99.50a        |
|                       | V1T1                  | 18.95B            | 5.00D-F          | 50.25K                        | 78.25J                         | 46.80M                      | 10.96G-I                         | 6.90m                             | 74.50b        |
|                       | V1T2                  | 17.55B-D          | 4.55F-H          | 38.50N                        | 58.77Q                         | 36.75O                      | 7.95JK                           | 8.92j                             | 74.25b        |
|                       | V1T3                  | 12.835D-F         | 4.23H-J          | 32.75O                        | 45.82S                         | 31.85P                      | 6.44M-P                          | 11.71f                            | 74.75b        |
|                       | V1T4                  | 8.75HI            | 3.65KL           | 28.50Q                        | 37.25U                         | 28.00R                      | 5.36OP                           | 15.28a                            | 49.50c        |
| V2                    | V2T0                  | 21.23A            | 4.80D-Q          | 76.00B                        | 128.00A                        | 66.35B                      | 15.64AB                          | 4.22q                             | 99.75a        |
|                       | V2T1                  | 18.63BC           | 4.50F-H          | 73.50C                        | 106.75D                        | 67.35B                      | 15.58AB                          | 6.20n                             | 99.75a        |
|                       | V2T2                  | 15.98E-Q          | 4.35GH           | 65.25F                        | 92.50H                         | 61.32EF                     | 12.60EF                          | 8.46k                             | 99.25a        |
|                       | V2T3                  | 13.46IJ           | 3.75I-L          | 67.25E                        | 92.70H                         | 63.92C                      | 11.54F-H                         | 11.36g                            | 99.25a        |
|                       | V2T4                  | 12.75J            | 3.78J-L          | 56.00H                        | 72.00L                         | 54.50H                      | 8.53J                            | 13.84b                            | 99.25a        |
| V3                    | V3T0                  | 18.83BC           | 4.90D-F          | 68.75D                        | 113.12C                        | 62.82CD                     | 11.79E-Q                         | 4.17q                             | 99.25a        |
|                       | V3T1                  | 13.40IJ           | 4.43F-H          | 64.50F                        | 98.45F                         | 60.80F                      | 9.82I                            | 5.63o                             | 99.00a        |
|                       | V3T2                  | 13.25IJ           | 3.50K-M          | 56.00H                        | 81.00I                         | 53.95HI                     | 7.58J-L                          | 8.11l                             | 99.50a        |
|                       | V3T3                  | 10.55K            | 3.40K-M          | 46.75L                        | 62.00O                         | 45.70M                      | 6.43L-P                          | 10.97h                            | 74.75b        |
|                       | V3T4                  | 9.25LM            | 3.30K-M          | 42.75M                        | 55.35R                         | 42.25N                      | 3.82Q                            | 13.47c                            | 74.25b        |
| V4                    | V4T0                  | 19.03B            | 6.25AB           | 118.50A                       | 125.55B                        | 117.10A                     | 16.75A                           | 3.89q                             | 99.25a        |
|                       | V4T1                  | 15.03GH           | 5.88BC           | 67.50DE                       | 102.00E                        | 62.35DE                     | 13.00DE                          | 5.88op                            | 99.00a        |
|                       | V4T2                  | 13.08IJ           | 4.73E-H          | 51.50JK                       | 71.62L                         | 49.07L                      | 10.76I-I                         | 7.99l                             | 99.50a        |
|                       | V4T3                  | 12.33K-M          | 4.58F-H          | 46.75L                        | 60.32P                         | 45.62M                      | 7.67J-L                          | 10.20i                            | 74.75b        |
|                       | V4T4                  | 10.75L-N          | 3.70J-L          | 37.00N                        | 45.25S                         | 36.47O                      | 6.01NOP                          | 12.47e                            | 74.25b        |
| V5                    | V5T0                  | 17.33C-E          | 5.13EF           | 67.50DE                       | 102.25E                        | 61.82D-F                    | 14.03CD                          | 4.01q                             | 99.25a        |
|                       | V5T1                  | 17.78B-D          | 3.65KL           | 64.75F                        | 95.15G                         | 60.65F                      | 11.82E-Q                         | 5.60o                             | 74.25b        |
|                       | V5T2                  | 17.025DE          | 3.50K-M          | 60.00G                        | 81.82I                         | 58.25G                      | 7.30JK-M                         | 7.91l                             | 74.25b        |
|                       | V5T3                  | 12.375IJ          | 3.33K-M          | 52.25J                        | 69.37M                         | 50.95K                      | 7.04K-N                          | 10.42i                            | 49.25c        |
|                       | V5T4                  | 8.250MN           | 2.95M            | 30.50P                        | 37.05U                         | 30.07Q                      | 6.09M-P                          | 13.06d                            | 49.00c        |
| V6                    | V6T0                  | 16.900DE          | 5.40CD           | 60.75G                        | 97.10F                         | 54.27H                      | 15.15BC                          | 3.98q                             | 99.75a        |
|                       | V6T1                  | 15.525FQ          | 4.36G-I          | 56.00H                        | 79.00J                         | 52.60IJ                     | 12.90D-F                         | 6.01n                             | 99.50a        |
|                       | V6T2                  | 13.525H-J         | 3.75J-L          | 54.25I                        | 75.00K                         | 51.87JK                     | 10.286HI                         | 7.07m                             | 74.25b        |
|                       | V6T3                  | 10.25KL           | 3.15LM           | 52.00J                        | 66.12N                         | 50.97JK                     | 6.77K-O                          | 10.21i                            | 49.50c        |
|                       | V6T4                  | 7.40N             | 2.28N            | 30.00PQ                       | 39.82T                         | 29.47QR                     | 5.05PQ                           | 12.35e                            | 25.00d        |
| LSD <sub>(0.05)</sub> |                       | 0.57              | 0.225            | 0.0005                        | 0.0005                         | 8.4052                      | 0.5238                           | 0.1351748                         | 0.497625      |
| CV (%)                |                       | 6.25              | 8.414            | 1.623                         | 1.726                          | 0.523                       | 8.405                            | 1.987608                          | 0.947033      |

Here, V1= BRRI dhan-28, V2= Burikatari, V3= Begunbahar, V4= Dular, V5= Pashpai, V6= Begunbichi

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#### 4. CONCLUSION

From this study, it ~~was disclosed~~can be concluded that moisture stress ~~imposed in all genotypes~~ causes differential responses ~~of in rice~~ genotypes ~~to impose water stress condition~~thus may indicate ~~the differential~~ drought tolerance ability of rice genotypes. Based on the findings of the present investigation, it was found that the genotype Burikarari showed the best performance in control conditions followed by BRRI Dhan 28 and Dular based on parameters. But with the increasing the degree of drought stress, the percentage of the morpho-physiological characters were less affected ~~by in~~ Burikatari. On the contrary, BRRI Dhan-28 were affected significantly higher followed by Begunbichi based on parameters. So, these findings suggested that the genotype Burikatari could be considered as more tolerant than the other genotypes against drought conditions.

**Comment [S9]:** Mention, based on which parameters ?

**Comment [S10]:** Mention which parameters?

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#### List of figures:

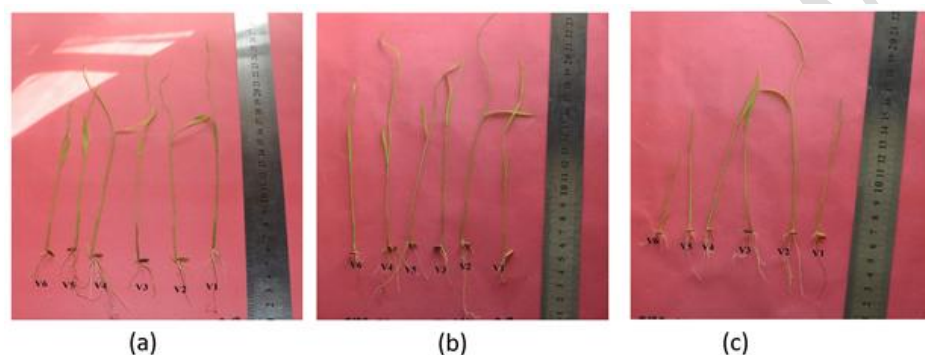


Figure 1. Effect of different water stress (a.  $0\text{gL}^{-1}$  PEG treatment, b.  $30\text{gL}^{-1}$  PEG treatment and c.  $60\text{gL}^{-1}$  PEG treatment) at *in vitro* conditions on six rice genotypes (V1= BRRI dhan-28, V2= Burikatari, V3= Begunbahar, V4= Dular, V5= Pashpai, V6= Begunbichi) after seventeen days of sowing.

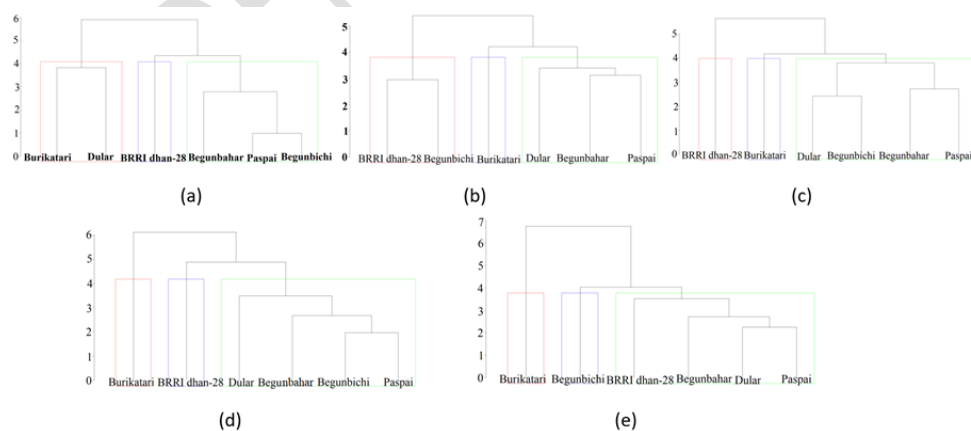


Figure 2. Dendrogram from UPGMA clustering for six rice genotypes using Euclidean genetic distance based on all traits measured in different stress water conditions (a.  $0\text{gL}^{-1}$  PEG treatment, b.  $15\text{gL}^{-1}$  PEG treatment, c.  $30\text{gL}^{-1}$  PEG treatment, d.  $45\text{gL}^{-1}$  PEG treatment and e.  $60\text{gL}^{-1}$  PEG treatment).

UNDER PEER REVIEW