

1 **Phytochemical Screening, Elemental and Proximate Analysis of *Maerua*** 2 ***angolensis* (Capparaceae) Stem Bark**

3 4 5 *Abstract*

6 *This work was designed to explore the phytochemicals, elemental and proximate analysis of*
7 *Maerua angolensis Stem bark were determined using standard analytical methods. The*
8 *phytochemical screening showed alkaloid (271.30 mg /100g), tannins (340.25 mg /100g),*
9 *flavonoid (176.85 mg /100g), reducing sugar (41.20 mg /100g), glycosides (184.30 mg /100g),*
10 *steroids (112.30 mg /100g), anthraquinones (167.85 mg /100g) and saponin (225.61 mg /100g).*
11 *Also the elemental analysis carried revealed the concentration of Manganese (0.029 mg/kg),*
12 *Copper (0.059 mg/kg), Calcium (0.070 mg/kg), Sodium (7.530 mg/kg), Zinc (0.028 mg/kg),*
13 *Chromium (0.158 mg/kg), Lead (0.007 mg/kg), Iron (0.100 mg/kg) and Magnesium (0.020*
14 *mg/kg). The result of the proximate composition showed that the moisture, fat, crude protein,*
15 *crude fibre, ash, carbohydrate and energy value content of the samples were 3.58±0.04 %,*
16 *6.25±0.09 %, 21.79±0.26 %, 48.51±2.31 %, 13.28±1.86 %, 6.60±1.79 % and 169.81±8.49*
17 *kcal/100g respectively. The P^H value obtained was 5.65±0.09. These indicate that the plant can*
18 *be effective source for drugs. The elemental and proximate analysis shows that it contained*
19 *appreciable amount of nutrients which could be included in diets to supplement human daily*
20 *nutrient needs and animal.*

Comment [U1]: pH not P^H

21
22 **Key words:** Wild, Edible, Plants, Extract, Composition, Drugs and Food

Comment [U2]: Keywords are usually arranged in alphabetical

23 24 25 26 27 28 **INTRODUCTION**

29 Natural products and their reactants have been utilized by man since creation, when plants'
30 leaves, bark or root are mixed with water to make a medicinal portion. A complex mixture of
31 organic natural product is actually extracted for its biologically active components [1].

32 According to FAO, about 1 billion people especially in developing countries depend on edible
33 wild plants in their diets [2]. The use of wild edible plants in different localities provide optimum
34 source of nutrients. Plants serve as indispensable constituents of human diet supplying the body
35 with mineral salts, vitamins and certain hormones precursors, in addition to protein and energy
36 [3]. Also plants serve as a source of medicinal product and shelter to man and his livestock. In
37 the earlier stage man depended on wild food, which is much abundant within his immediate
38 environment, as the population grows, however, sources of food became more difficult to him,
39 which necessitated domestication of many plants [1, 4].

40

41 Food is no doubt the most basic necessity for one to efficiently function in his own ecosystem
42 [5]. Since creation, man has used plant as source of food and drug [6]. The use of medicinal
43 plants as food alternative traces back to ancient human civilization [7, 8]. The useful products
44 obtained from plants directly or indirectly, demonstrate their importance to man.

45 There are about 35,000 species of higher plants that are used for medicinal purposes [9]. Some
46 of these species of plants contain nutrients that have therapeutic properties and are nutritionally
47 important because of their high contains of minerals, essential fatty acids, fibers and proteins
48 [10]. Phytochemicals are chemical compounds that occur naturally in plants. The efficiency of
49 medicinal plants for therapeutic purposes is often based on their organic constituents such as
50 flavonoids, tannins, alkaloids and essential oils [11]. Traditionally the usage of plant in curing
51 illness has deep roots in human history [8]. Ethno pharmacological uses of plants prevail among
52 the various Nigeria communities. Plants continue to play a prominent role in primary health care
53 of about 80% of the world population. [10].

54 Plants are the cheapest and most important available sources of nutrients, supplying the body
55 with mineral salts, vitamins and certain hormone precursors, protein, energy and essential amino
56 acids [12, 13].

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59 *Maerua angolensis* (family Cappariaceae) is a tropical plant that is widespread in the savannah
60 area of tropical Africa to South Africa and Swaziland. It is a tree whose size varies from medium
61 to big and growing up to 10 – 20 meters high. It is commonly found growing in bush and rocky
62 areas but planted on graves in Nupe area of Nigeria. *Maerua angolensis* has a long history of use
63 in traditional medicine to manage various painful conditions in Nigeria and other West African
64 countries [14, 15].

65 The plant is used in indigenous systems of medicine for the treatment of psychosis ecthyma,
66 epilepsy, diarrhea and dysentery. It is also used as sedative and treatment of liver disease. [7].

67 *Maerua angolensis* has been extensively used in traditional system of medicine in many
68 countries. It is used in the treatment of pain, however lacking a pharmacological evaluation for
69 its analgesic effect, the leaves, stem bark and roots of *Maerua angolensis* have significant
70 analgesic activity against chemical induced pain model in mice. In a dose dependent manner,
71 however stem bark extract was more potent. [16]. It is used locally for the treatment of
72 psychosis, ecthyma, epilepsy, laxative sedative, dyspepsia neurasthenia, liver disease [8].

73 The pharmacological investigation shows that *Maerua angolensis* is worldwide used for
74 antioxidizing value [17]. The oxidative stress defines itself as being a loss of the balance between
75 oxidizing and antioxidants within a cell [18].

76 Phytochemical screening conducted on the plant of recent showed that the methanol extract of
77 the stem bark contained tannins, saponins, flavonoids, cardiac glycosides and alkaloids [19].

78 As various medicinal plant species are used either in the form of extract or decoction by the local
79 people in different regions, therefore, evaluating their nutritional significance can help to
80 understand the worth of these plants species in different ecological conditions. Some of these
81 medicinal plants serve as both food and medicine [20].

82 Quantifying proximate composition is important in ensuring the requirements of food regulations
83 and commercial specifications for instance moisture content of flesh (of a substance) is a good
84 indicator of its relative content of energy, protein and lipid [21, 22].

85 According to [23] use of wild edible plant in different localities provides optimum source of
86 nutrients. The quantity of nutrient and Phytochemicals varies not only with the species of edible
87 plant but also (for the same Variety) with the location in which they are growing because of the
88 variation of the soil on which they grow and other environmental factors of the location. Hence it
89 is important to evaluate nutrient and phytochemicals of the edible plants in given location, before
90 recommending them as sources of food for the local community.

91 The information on the nutrient and phytochemicals of *Maerua Angolaensis* (capparaceae) stem
92 bark growing in the study area is scanty.

Comment [U3]: angolensis, not Angolensis

93 The objective of the study is to investigate the phytochemicals, elemental and proximate
94 constituents of *Maerua angolensis* (capparaceae) stem bark in the study area. The findings will
95 serve as a guide to dietetics and nutritionist whether or not to recommend it as sources of drugs
96 and food to the local community.

97 **MATERIALS AND METHODS**

98 **Sample Collection and Identification**

99 *Maerua angolensis* plant belongs to the family *Capparaceae* and was collected from the bush
100 area of Muchala, Mubi North Local Government Area of Adamawa State, Nigeria. The plant was

101 identified by a taxonomist in the Department of biological Science, Adamawa State University
102 Mubi and preserved in the Department of Chemistry.

103 **Preparation of Sample**

104 The stem bark of *Maerua angolensis* plant was air dried in Chemistry laboratory 2, Science
105 Complex of the Faculty of Science, Adamawa State University, Mubi. The plant was air dried
106 under shade and was weighed and grounded to get a coarse powder form using sterile mortar and
107 pestle. The powder was stored in an air tight container and was used for successive analysis [24].

Comment [U4]: how many mesh powder particle size

108 **Phytochemical analysis**

109 *Maerua angolensis* stem bark was tested for the presence of bioactive compounds. The
110 phytochemicals of the plant samples were estimated following the procedure adopted by
111 Nwankwo and Ukaegbu-Obi, [25]

112

113 **Test for tannins**

114 200 mg of crude plant extracts was mixed with 2 ml of 2% solution of FeCl₃. Blue-green color
115 was observed which indicates the presence of tannins.

116

117 **Test for flavonoids (alkaline reagent test)**

118 200 mg of extract was mixed with 2 ml of 2% solution of NaOH. An intense yellow colour
119 formed which turned colorless on addition of few drops of diluted acid was observed which
120 indicated the presence of flavonoids.

121 **Test for saponins**

122 200 mg of extract was mixed with 5 ml of distilled water in a test tube and was shaken
123 vigorously. The formation of Stable foam was observed which shows an indication of the
124 presence of saponins.

Comment [U5]: for 10 minutes and add 1 ml HCL 2M, the foam persistent stable

125 **Test for anthraquinones**

126 0.5g of the sample was boiled in 3ml of 1% HCl and filtered. The filtrate was shaken with 5ml
127 benzene and the benzene layer was removed, 10 % NH₄OH was added and pink/violet color in
128 the alkaline phase was observed which indicated the presence of anthraquinones.

129 **Test for glycosides (Salkowski's test)**

Comment [U6]: This title move to the following page, do not separate from the content

130 200 mg of extract was mixed with 2 ml of chloroform. Then 2 ml of concentrated H₂SO₄ was
131 added carefully and shaken gently. A reddish brown color was observed which indicated the
132 presence of steroidal ring, that is, glycone portion of the glycoside.

133 **Test for alkaloids**

134 200mg of extract was mixed with 10 ml of methanol. To 2ml of the filtrate was added 1% HCl
135 and then steamed. To 1ml of the filtrate was added 6 drops of Wagner reagent. Brownish-red
136 precipitate was observed which indicated the presence of alkaloids.

137 **Test for steroids**

138 To 2ml of acetic anhydride was added 0.5g of the sample followed by an addition of 2ml H₂SO₄.
139 The color changed from violet to blue green indicating the presence of steroid [25]

140

141 **Test for reducing sugar**

142 0.5g of the sample was dissolved in 5ml water, and small amount of Benedict reagent was added.
143 During a water bath, which is usually 4-10 minute, the solution progressed in the colour of blue
144 (with no glucose presence), green, yellow, orange, red and brown indicating high glucose
145 presence.

Comment [U7]: 8 drops not small amount

146 **Test for terpenoids (Salkowski test)**

147 5 ml of each extract was mixed in 2 ml of chloroform, and concentrated H₂SO₄ (3 ml) was
148 carefully added to form a layer. A reddish brown coloration of the interface was formed which
149 was an indication of positive results for the presence of terpenoids [25]

150

151 **Determination of phytochemicals by HPLC**

152 5g of prepared sample was placed into 25cm³ standard volumetric flask and made up to mark
153 over diluent. The solution was refluxed, shaken, centrifuged and decanted. Then filtrate was
154 filtered using the HPLC grade filter paper [26].

Comment [U8]: and then inject to HPLC

155 **Elemental analysis**

156 The dried sample was weighed into a crucible and placed in a muffle furnace at room
157 temperature and the temperature was raised to 550⁰C for three hours to complete ash. The ash
158 was dissolved in hot (10% HNO₃), filtered and diluted to required volume in a standard flask
159 with (0.01M HNO₃). The elements in solution were determined using atomic absorption
160 spectrophotometer (AAS) following the procedure adopted by AOAC [26].

161 **Proximate Analysis of the sample**

162 Proximate Analysis of the samples was carried out following the methods described by Mbaeyi-
163 Nwaoha and Emejulu [27].

164

165 **Statistical Analysis**

166 All determinations were replicated three times and results were reported in mean (\pm) standard
167 deviation.

168

169 **RESULTS**

170 The results of the phytochemical constituents and their quantities are presented in Tables 1 and 2
171 respectively while Table 3 contains the result of the elemental analysis. The results for the
172 proximate compositions are shown in Table 4.

173

174 The results indicates that the plant is rich in crude fibre with the highest percentage followed by
175 crude protein meaning that the plant is rich and a good source of protein. It has appreciable ash
176 content indicating that the plants contain inorganic components. The proximate compositions is
177 in the order Crude Fibre > Crude Protein > Ash Content > Carbohydrate > Crude Lipid >
178 Moisture Content (Table 4). The plant is rich in organic matter with 86.72% indicating that it's a
179 moderate source of energy. Figure 1 Shows the Comparison of the proximate composition of
180 Maerua angolensis with other edible plants.

181

182

183 **Table 1: Result of phytochemical constituents present**

Phytochemical	Sign
Alkaloids	+
Tannins	+
Flavonoids	+
Reducing sugars	+
Steroids	+
Anthraquinones	+
Saponins	+

184 **Keys:** + = Present

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186 **Table 2: Result of the quantitative phytochemical analysis of the sample**

Comment [U9]: Letters in the contents of the table must be smaller than the text

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187 (Composition in mg/100g)

Phytochemical Constituents	Values obtained
Alkaloids	271.30
Tannins	340.25
Flavonoids	176.85
Reducing sugars	41.20
Steroids	112.30
Anthraquinones	167.85
Saponins	225.61

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190 **Table 3: Results of elemental composition of the samples (mg/kg)**

Elements	Concentration
Magnesium (Mg)	2.516 ± 0.020
Iron (Fe)	5.200 ± 0.100
Lead (Pb)	0.146 ± 0.007
Chromium (Cr)	3.233 ± 0.158
Zinc (Zn)	1.256 ± 0.028
Manganese	1.270 ± 0.029
Copper (Cu)	2.243 ± 0.059
Cadmium (Cd)	B.D.L.
Calcium (Ca)	5.066 ± 0.070
Sodium (Na)	163 ± 7.530

191 Keys: B.D.L. = Below Detectable Level

192 The data are mean of three replicates plus standard deviations.

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195 **Table 4 Proximate compositions (% Dry Weight bases) of *Maerua angolensis* stem bark**

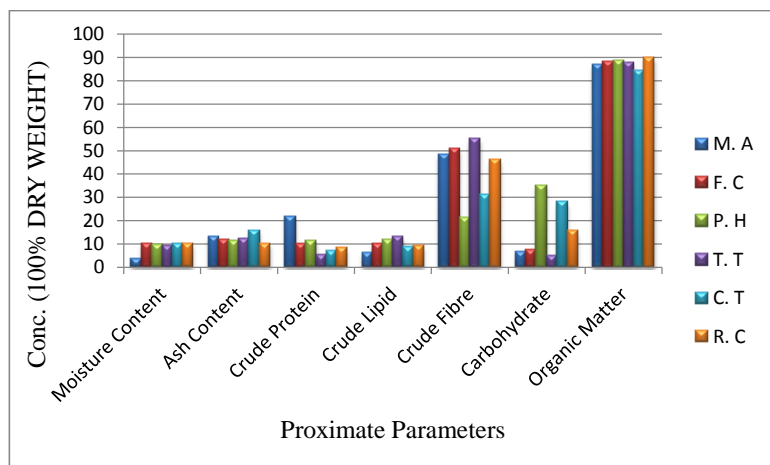
Comment [U11]: This table moves to the next page, don't separate

Parameters

Mean±S.D

Moisture Content	3.58±0.04
Ash Content	13.28±1.86
Crude Protein	21.79±0.26
Crude Lipid	6.25±0.09
Crude Fibre	48.51±2.31
Carbohydrate Content	6.60±1.79
Organic Matter	86.72±1.86
Caloric Value (kcal/100g)	169.81±8.94
<i>p</i> ^H Value	5.65±0.09

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M. A = Maerua angolensis, F. C = *Fagonia cretica* L., P. H = *Peganum harmala* L., T. T = *Tribulus Terrestris* L., C. T = *Chrozophora tinctoria* and R. C = *Ricinus communis* L.,

Fig. 1 Comparison of the proximate composition of Maerua angolensis with some other edible plants

224 **DISCUSSION**

225 Table 1 shows the result of the phytochemical constituents screening which revealed that
226 alkaloids, tannins, flavonoids, reducing sugars, glycosides, steroids, anthraquinones and saponin
227 were presence.

228 The result of phytochemical analysis revealed the compositions of the constituents in the
229 following order: Tannins (340.25) > Alkaloids (271.30) > Saponins (225.61) > Glycosides
230 (184.30) > Flavonoids (176.85) > Anthraquinones (167.85) > Steroids (112.30) > Reducing
231 sugars 41.20 mg/100g (Table 2).

232 The presence of these metabolites indicates the great potentials of the plant as a source of useful
233 phytomedicines. For instance the presence of flavonoids might be responsible for its use as anti-
234 inflammatory recipe in Chinese **follelore** medicine as some flavonoids have anti-inflammatory
235 effects on both acute and chronic inflammation [7].

Comment [U12]: Folklore not follelore

236 Some plants that posse alkaloids are known for decreasing blood pressure and balancing the
237 nervous system in case of mental illness. The presence of tannins could also show that it is an
238 astringent help in wound healing and anti-parasitic. The presence of saponins shows the class of

239 natural products involved and can be used to enhance penetration of micro molecules such as
240 protein through cell membrane. It also indicates the plant potential activity on antimicrobial
241 agents [16].

242 These findings were in agreement with the result reported by Ayo, *et al.* [19] which revealed the
243 presence of reducing sugars, alkaloids, saponins, flavonoids and tannins in the methanolic
244 extract of *Maerua angolensis* leaves.

245 The result of elemental analysis of the samples revealed that the plant contains Mg, Fe, Pb, Cr,
246 Zn, Mn, Cu, and Ca. The most abundant element is Na (163 ± 7.530), followed by the rest in the
247 order Iron (5.200 ± 0.100) > Calcium (5.06 ± 0.70) > Chromium (3.233 ± 0.158) > Magnesium
248 (2.516 ± 0.020) > Copper (2.243 ± 0.059) > Manganese (1.270 ± 0.029) > Zinc (1.256 ± 0.028) >
249 Lead (0.146 ± 0.007) while Cadmium is below detectable level (Table 3).

250 Magnesium is an important mineral element in connection with circulatory diseases such as
251 heart disease [7]. High magnesium concentration is a component of leaf chlorophyll in plants.

252 Copper is an essential trace element in human body and exist as an integral part of copper
253 proteins **cerulosmin** which is concerned with the release of Iron from the cells into the plasma
254 and is involved in energy metabolism [28]. The presence of copper, manganese and zinc
255 indicates that the plant is essential for immune function [29].

Comment [U13]: Ceruloplasmin not cerulosmin

256 Lead occurs naturally in the environment. Every one may be exposed to trace amount of lead
257 through air, soil, house hold dust, food, drinking water and various consumer products [30].

258 Sodium has an important role in maintaining the water balance within the cells and in the
259 function of both nerve impulse and muscles. It also helps in the maintenance of normal acid-base
260 balance. An adult need about 3g per day of sodium but modern dietary habits take in 5 – 20 g per
261 day [31].

262 Calcium plays an important role in building and maintaining strong bones and teeth, large part of
263 human blood and extracellular fluids. Approximately 99% of the body calcium is stored in the
264 bones and teeth [32]. The studied plant (*Maerua angolensis*) is essential in building up the level
265 of calcium in the body.

266 Cadmium was not detected in the sample. Cadmium causes kidney and liver problem on long
267 time of its accumulation [33]. *Maerua angolensis* is safe for consumption since these toxic
268 elements are in low concentration or not detected.

269

270 The moisture content of the sample was found to be $3.58 \pm 0.04\%$ (Table 4). The low moisture
271 content would hinder the growth of microorganisms and the storage life would be high [34].

272 Moisture content is among the most vital and mostly used measurement in the processing,

273 preservation and storage of food [35]. The moisture content of sample is lower compared to
274 9.70%, 10.10%, 10.30% 10.30 and 9.20 in *Peganum harmala L.*, *Chrozophora tinctoria L.*,
275 *Ricinus communis L.* in *Fagonia cretica* and *Tribulus Terrestris L.* respectively (Figure 1) [36].

276

277 The ash content of the sample was $13.28 \pm 1.86\%$ on dry matter (DM) bases (Table 4). Ash in
278 food contributes the residue remaining after all the moisture has been removed as well as the
279 organic materials (fat, protein, carbohydrates, vitamins, organic acid etc) have been incinerated
280 at a temperature of about 500°C . Ash content is generally taken to be a measure of the mineral
281 content of the original food [35, 8]. The ash content of the sample is slightly high compared to
282 11.20%, 10.10%, 12.00% and 12.10% in *Peganum harmala L.*, *Ricinus communis L.*, *Fagonia*
283 *cretica* and *Tribulus Terrestris L.* respectively and slightly lower than *Chrozophora tinctoria L.*,
284 (15.70%) figure 4.

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286 Crude fibre in food or plant is an indication of the level of non-digestible carbohydrate and
287 lignin. The crude fibre obtained in the sample was $48.51 \pm 2.31\%$ on DM bases (Table 4). The
288 result obtained showed a good amount of fibre in the stem, an indication that it can serve as a
289 good source of fibre which might aid digestion, help reduce serum cholesterol level, risk of
290 coronary heart disease and hypertension [37]. The crude fibre content was high compared to
291 21.10%, 30.9% and 46.2% in *Peganum harmala L.*, *Chrozophora tinctoria L.*, and *Ricinus*
292 *communis L.* respectively but low compared to 50.8% and 55.20% in *Fagonia cretica* and
293 *Tribulus Terrestris L.* respectively Figure 4. Crude fibre is made up largely of cellulose together
294 with a little lignin which is indigestible in human [35]. The high fibre and protein content is a
295 further confirmation of its use as vegetable. Fibre reduces **tracolonc** pressure which is beneficial
296 in diverticular disease. Plants with high fibre are adequate for better rumination and digestion in
297 ruminant animals [8, 36].

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299 The crude lipid content was $6.25 \pm 0.09\%$ (D M) bases (Table 4). Lipid provides very good
300 sources of energy and aids in transport of fat soluble vitamins, insulates and protects internal
301 tissues and contributes to important cell processes. More so, it is good to add lipid (fat) to most
302 of our diets, because many body functions depend on lipids [35]. The crude lipid content of
303 *Maerua angolensis* stems is low compared to 11.80%, 8.50%, 9.20%, 9.80% and 13.30% in

Comment [U14]: Intracolonic not tracolonc

304 *Peganum harmala L.*, *Chrozophora tinctoria L.*, *Ricinus communis L.*, *Fagonia cretica* and
305 *Tribulus Terrestris L.* respectively (Figure 1). Crude lipid are the principal sources of energy.
306 One gram of lipid provides 9.0 kcal (37.33 kJ) of energy [37] which indicates that 100 g of
307 *Maerua angolensis* stem lipid should provide about 56.25 kcal (233.31 kJ) of energy.

308

309 The crude protein of the sample was $21.79 \pm 0.26\%$ DM bases (Table 4). The recommended
310 dietary allowance (RDA) for protein is 56g for individual weighing 70kg and 46g for adult
311 weighing 50kg, children may consume 2kg/day [35, 36]. The plant is a moderate source of
312 protein. According to Akpabio and Ikpe, [35], proteins from plant sources have lower quality but
313 their combination with many other sources of protein such as animal protein may result in
314 adequate nutritional value. The crude protein content of the sample was high compared to
315 11.20%, 6.90%, 8.40%, 9.80 9.80% and 5.20% in *Peganum harmala L.*, *Chrozophora tinctoria*
316 *L.*, *Ricinus communis L.*, *Fagonia cretica* and *Tribulus Terrestris L.* respectively (Figure 1)

317

318 The carbohydrate content was $6.60 \pm 1.79\%$ (Table 4). The plant is a low source of carbohydrate
319 when compared with the Recommended Dietary Allowance (RDA) of 130g [34, 35] but can be
320 used as supplement in an extreme condition of carbohydrate requirement. The carbohydrate
321 content of the samples were low compared to 35.00%, 27.90%, 15.80%, 7.20% in *Peganum*
322 *harmala L.*, *Chrozophora tinctoria L.*, *Ricinus communis L.* and *Fagonia cretica* respectively but
323 high compared to 5.00% in *Tribulus Terrestris L.*(Figure 1). This relatively low carbohydrate
324 content makes it suitable to be eaten when one wants to lose weight [36].

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327 The caloric value of the samples were 169.81 ± 8.49 kcal/100g. An average person requires 2000-
328 3000 kcal per day [16]. The total energy value was low compared to 418.10 kcal/100g, 420.20
329 kcal/100g, 422.80 kcal/100g, 440.70 kcal/100g, and 380.60 kcal/100 g in *Peganum harmala L.*,
330 *Ricinus communis L.*, *Fagonia cretica L.*, *Tribulus Terrestris L.* and *Chrozophora tinctoria L.*
331 respectively (figure 1). The plant can contribute to the caloric requirement of the body. The low
332 calorific value of *Maerua angolensis* stem is an indication that it can be recommended to
333 individuals suffering from overweight and obesity [36].

334

Comment [U15]: Too much ??? child needs approximately 18 – 20 grams protein per day

335 The organic matter content of the samples was obtained to be 86.72±1.86% on dry matter bases
336 (Table 4) indicating a high level of organic components compared to the inorganic composition
337 with a value about 13.28%.

338

339 The *pH* value of the samples was 5.65±0.09 indicating that the plant is weakly acidic in nature
340 probably as a result of the soil P^H where the plant's habitation. Based on this finding patient with
341 hyper acidic problem could be advice not to consume the plant in excess.

342

343 CONCLUSION

344 The result of this study indicated that the plant contain some major phytochemicals that inhibits
345 the growth of micro-organism thereby proving very effective source of drugs. This means the
346 plants could be used for remedy of dysentery, diarrhea, typhoid, fever and treatment of
347 hypertension.

348 Furthermore the result of the elemental analysis showed appreciable amount of minerals content
349 in the plant. This indicates that the plant could be a source of minerals in diet as well as drugs in
350 pharmaceutical industries.

351 The result of the proximate composition showed the nutritive value of the plant, which indicates
352 that the *Maerua angolensis* analyzed have a great potential as sources of food particularly
353 considering their proximate composition. The ash content signifies that the plant is a potential
354 plant to supply the body with important minerals.
355 Therefore, *Maerua angolensis* plant could contribute significantly to the nutrient requirement of
356 both men and animals.

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Comment [U16]: pH not P^H

Comment [U17]: microorganism not micro-organism

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Reference

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367

- 368 1. Kawo A.H., Abdullahi B.A., Gaiya, Z.A. Halilu, A., Dabai, M. and Dakare, M.A. (2009).
369 Preliminary Phytochemical Screening, Proximate and Elemental Composition of Moringa
370 Oleifera Lam Seed Powder. *Bayero Journal of Pure and Applied Sciences*, Vol. 2(1), 96-
371 100.
- 372 2. Agbo O. J., Shomkegh S.A. and Mbakwe R. (2013). Local Perception And Proximate
373 Analysis Of Some Edible Forest Plants Around University Of Agriculture Wildlife Park,
374 Benue State, Nigeria. *Journal of Research in Forestry, Wildlife And Environmental*, Vol.
375 5(1); 10-22.
- 376 3. Tairo Vendeline Emmanuel, Jesse T. Njoka, Lukhoba W. Catherine and Herbert V.M. Lyaruu.
377 (2011). Nutritive and Anti-nutritive Qualities of Mostly Preferred Edible Woody Plants in
378 Selected Drylands of Iringa District, Tanzania. *Pakistan Journal of Nutrition*, Vol. 10(8);
379 786-791.
- 380 5. Umar K. J., Hassan L. G. and Garba H. J. (2005). Proximate and Mineral Composition of M.
381 Mirista. *Chem. Class J.*, 81-84.
- 382 6. Rosemary I U and Donatus E O (2012). An Evaluation of the Phytochemical and Nutrient
383 Composition of the Seeds and Stem Bark of Detarium senegalense Gmelin. *Journal of*
384 *Natural Sciences Research*, Vol. 2(5), 107-111.
- 385 7. Hussain J, Najeeb U R, Abdul L K, Liaqat A, Ahmed A, Zabta K S, Hidayat H and Tania S
386 R (2013). Proximate Based Comparative Assessment of Five Medicinal Plants to Meet
387 the Challenges of Malnutrition. *European Journal of Medicinal Plants*, Vol. 3(3); 444-
388 453.
- 389 8. Nwauzoma A. B. and Dawari S L. . (2013). Study on the phytochemical properties and
390 proximate analysis of Piper Umbellatum (LINN) from Nigeria. *American Journal of*
391 *Research Communication*. Vol. 1(7); 164-177.
- 392 9. Boroomand N, Grouh MSH (2012) Macro element Nutrition (NPK) of Medicinal Plant. A
393 review. *J. Med plant Red* .Vol. 6(12).2249-2255.
- 394 10. Barker V. and Nsekuye N. (1995). *traditional Vertinary practice in Africa*. German
395 Technical Cooperation. Pp 45-57.
- 396 11. Sharma RK, Agrawal M. Marshall FM (2009) heavy metals in vegetables collected from
397 production and marked sites of tropical urban area of India *food chem. Toxicol* Vol.
398 47(3); 583-59.

- 399 12. Andzouana M and Mombouli J B (2012). Proximate, Mineral and Phytochemical Analysis of
400 the Leaves of *H. myriantha* and *Urera trinervis*. *Pakistan Journal of Biological Sciences*,
401 *Vol. 15*; 536-541. DOI: 10.3923/pjbs.2012.536.541.
- 402 13. Frances I O, Thomas M O and Gabriel I O (2013). Effect of processing methods on the
403 chemical composition of *Vitex doniana* leaf and leaf products. *Food Science & Nutrition*
404 *published by Wiley Periodicals, Inc., 1*, DOI: 10.1002/fsn3.31.
- 405 14. Wild H. (1960). *Flora Zambesiaca (F. Z.) (Vol. 1)*. Pp25-27.
- 406 15. Burkri, H.M (1985). The useful Plants of Tropical Africa, 2nd ed. Royal Botanical Garden. Kew,
407 Richmond, Surrey UK pp 334-335 content of some wild vegetable leaves consumed in
408 eastern anotocia, *acta agric sci and B-soil plants sci* .Vol. 53(3):129-137.
- 409
410
- 411 16. Iliya H A, Boakye E, Adongo W D, Ampadu F A, Woode E (2014). Antinociceptive activity
412 of various solvent extracts of *Maerua angolensis* DC stem bark in rodents. *The Journal of*
413 *Phytopharmacology*, Vol. 3(1);1-8.
- 414 17. Meda N.T. (2013). Antioxidant activity of phenolic and flavonoid fractions of *Cleome*
415 *gynandra* and *Maerua angolensis* of Burkina Faso. *Journal of Applied Pharmaceutical*
416 *Science*, Vol. 3(02); 036-042.
- 417 18. Mohammed A., Tanko Y., Okasha M. A., Sadiq Y., Isa A. I. (2008). Effect of aqueous
418 methanolic stem bark of *Maerua angolensis* (Capparidaceae) extract on blood glucose
419 levels of streptozocin-induced diabetic wistar rats. *Research Journal of Pharmacology*,
420 *I(CCCC)*, 1- 4.
- 421 19. Ayo R. G. ,Audu O. T., Amupitan J. O. and Uwaiya E (2013). Phytochemical screening and
422 antimicrobial activity of three plants used in traditional medicine in Northern Nigeria.
423 *Journal of Medicinal Plants Research*, Vol. 7(5); 191-197.
- 424 20. Friday E T. and Omale J (2010). Proximate and nutrient composition of *Euphorbia*
425 *hetrophylla*: A medicinal plant from Anyigba, Nigeria. *Journal of Medicinal Plants*
426 *Research*, Vol. 4(14); 1428-1431. DOI: 10.5897/JMPR10.295.
- 427
428 21. AOAC (2005) Official methods of Analysis of Association of Analytical Chemists. AOAC
429 International, 18th ed; Horowitz, W.(ed) vol 1 & 2, AOAC *International Maryland, USA*
430 pp 774-784.
- 431
432
433

- 434 22. Sutharshiny S. and Sivashanthini K. (2011). Proximate Composition of Three Species of
435 Scomberoides Fish from Sri Lankan Waters. *Asian Journal of Clinical Nutrition* Vol. 3;
436 103-111, doi:10.3923/ajcn.2011.103.111.
- 437 23. FAO/WHO (2011). Codex Alimentarius Commission/FAO/WHO food standards,
438 “Standard for named vegetable oils”, CODEX-STAN 210 Ed. FAO/WHO.
439
- 440 24. Kubmarawa D., Andenyang I. F. H. and Magomya A. M.(2009). Proximate Composition and
441 Amino Acid profile of two non-conventional leafy vegetables (*Hibiscus cannabinus* and
442 *Haemastaphis bacteri*). *African Journal of Food Science*, Vol. 3(9); 233-236
- 443 25. Nwankwo I. U. and Ukaegbu-Obi K. M. (2014). Preliminary phytochemical screening and
444 antibacterial activity of two Nigerian medicinal plants (*Ficus asperifolia* and *Terminalis*
445 *catappa*). *Journal of Medicinal Plant and Herbal Therapy Research*, Vol. 2, 1-5.
- 446 26. AOAC (2000). *Official Methods of Analysis of the Association of Official’s Analytical*
447 *Chemists*,
448 (17th edn.) Arlington, Virginia. pp. 96-105.
449
- 450 27. Mbaeyi-Nwaoha I.E. and Emejulu V.N. (2013). Evaluation of Phytochemical Composition
451 and Antimicrobial Activity of Sweet Potato (*Ipomoea batatas*) Leaf. *Pakistan Journal of*
452 *Nutrition*, Vol. 12(6); 575-586.
- 453 28. Indrayan, AK, Sharma, SD, Durgapal, N Kumar and Kumar , M (2001). “Determination of
454 nutritional value and analysis of mineral element for some medicinal valued plants
455 from Uttaranchal” *Current sc* 89(7): 1252-1257.
456
- 457 29. Timothy N (2019). Variation of Heavy Metal Concentration in Soil and Plant with Distance
458 Away from the Edge of the Road and Depth at which the Soil Samples were taken
459 Along Song – Yola Highway Adamawa State Nigeria *International Journal of*
460 *Ecology and Development Research* Vol. 5(1); 053-061.
461
- 462 30. Shivery D and Sofora A (2009) *medical plants and traditional medicine in Africa spectrum*
463 *book ltd Ibadan, Nigeria*, pp.289.
464
- 465 31. Njidda A.A. ,Olatunji E. A. and Garba M. G. (2013). In Sacco and In Vitro Organic Matter
466 Degradability (OMD) Of Selected Semi Arid Browse Forages. *IOSR Journal of*
467 *Agriculture and Veterinary Science*, 3(2), e-ISSN: 2319-2380, p-ISSN: 2319-2372, 9-16.
- 468 32. Holleman G, Arnold F, Wibora, G and Egon W W (1988) “Nutrition” *lehrbuch der*
469 *anorganischen Cemie (in German) (91-100ed) Walter de Gruster* issue 3. Pp 931- 943,
470 11-609511-3.
471

472

473 33. Grabley S. and Thiericke R. (1999). *Drug discovery from Nature*. London: Springer pp29-35.

474 34. Bamishaiye E.I., Olayemi F.F., Awagu E.F and Bamshaiye O.M. (2011). Proximate and
475 Phytochemical Composition of *Moringa oleifera* Leaves at Three Stages of Maturation.
476 *Advance Journal of Food Science and Technology*, Vol. 3(4); 233-237.

477 35. Akpabio U. D. and Ikpe E. E. (2013). Proximate composition and nutrient analysis of
478 *Aneilema aequinoctiale* leaves. *Asian Journal of Plant Science and Research*, Vol. 3(2);
479 55-61.

480 36. Mann A. and Otori A. A. (2014). Determination of chemical composition, minerals and
481 antinutritional factors of two wild seeds from Nupeland, North Central Nigeria. *American*
482 *Journal of Chemistry and Application*, 1(1), 20-26, <http://www.aascit.org/journal/ajca>.

483 37. Adinortey M.B., Sarfo J.K., Quayson E.T., Weremfo A., Adinortey C.A., Ekloh W. and
484 Ocran J. (2012). Phytochemical Screening, Proximate and Mineral Composition of
485 *Launaea taraxacifolia* Leaves. *Research Journal of Medicinal Plant*, Vol. 6; 171-179.
486 DOI: 10.3923/rjmp.2012.171.179.

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488

489

490

491