1 Niche-proxies of soil from rhizosphere of Weeds from Bodo in Gokana, Rivers State,

2

3

Nigeria.

ABSTRACT

4 Niche-ecology and isolation studies of microbes from the environment have been described 5 as the bedrock and driving-force for bioprocess industry. Ten (10) Cyperus esculentus, Scleria pauciflora., Asystasia gangetica Harungana madagascariensis, Ancistoclaudus 6 7 tectorius, Kyllinga erecta Cinna arundinacea, Brassica chinensis, Cyperus difformis, 8 Kyllinga bulbosa and Brachiaria mutica weeds and soil were obtained from Bodo, Gokana 9 LGA, Sludge farm and Botanical garden of the University of Port Harcourt, Rivers State, Nigeria. The soil was enriched in Mineral Salt Media and Bonny Light Crude Oil, prior to the 10 spread-plating on solidified media. Result of the analysis showed pH of soil samples ranged 11 from 5.26-7.2; Electrical conductivity was 53.4-80.31µS/cm and phosphate 0.74-5.35 mg/kg. 12 13 Levels of Vanadium in pre-impacted rhizosoil obtained from Kyllinga erecta and Cinna arundinaceae was 0.61 and 0.70 mg/kg respectively. Moisture content of soil obtained from 14 15 polluted and pristine environments were 11.75% and 17.82%. Permeability indices were 9.0 describing the soil to have low plasticity. Total heterotrophic bacterial count was within 7.5-16 17 7.77 Log₁₀ Cfu/g, with associated microbial isolates such as with Cyperus esculentus 18 rhizosphere soil was more dominated with others like Achromobacter sp, B. lichenformis, B. anthracis, B. subtilis, B. fumari, Arthrobacter sp, Pseudomonas sp, Ps. aeruginosa, Ps. 19 Florescens present, while fungal isolates were Aspergillus terreus, Trichoderma sp, and 20 Fusarium sp. These findings further supports the rhizosphere of plants as a rich bioresource 21 for biomining of high throughput strains for biotechnological application. 22

23 Keywords: Bioresources, Microbial isolates, Niche-ecology, Rhizosphere, Pristine24 environment,.

25

26 Introduction

27 Oil exploration in Nigeria has remained a live-wire for growth, sustenance and development of Nigeria, these events have also left the nation with a lot of environmental 28 challenges. Issues such as population exploration, increased industrialization and 29 30 urbanization have increased the spate of the problems in modern times (Nwachukwu & Osuagwu, 2019). Oilspill is a term used in the industry to mean the release of crude oil or its 31 fractions into the environment. Over 1020 oil spill incidence have been reported in Nigeria, 32 33 with Niger Delta taking a centre stage of the cases reported in the news media. These cases have caused devastating effects on both fauna and flora of the soil_(REF). The effect of 34 35 pollution on both aquatic and terrestrial ecosystems have different levels of severity to the biota., these Presence of pollutants above recommended threshold in the environment is 36 37 deleterious to soil biota at varied proximal niches as pollutants ean be seen due to the increase are prone to percolation and seepages, the effect of the spill can which have far reaching 38 39 effect to non-target population (Ofoegbu, Momoh and Nwaogazie 2015). These activities 40 Presence of elevated concentrations of pollutants in the soil affects the soil fertility and

Comment [PW1]: Topic does not clearly reflect what is in manuscript. Rephrase it to reflect the aim

Comment [PW2]: Rewrite after all corrections. Check for significant findings to appear here. Mention methodologies.

Comment [PW3]: Avoid such terms in writing scientific paper. The first sentence is hanging as the word remained is past tense and refers to a previous explanation which does not exist in this manuscript. Rephrase the sentence.

Comment [PW4]: Which events are you referring t? growth, sustenance and development or Oil exploration. Rephrase the two sentences to flow

Comment [PW5]: Consider using "population explosion".

Comment [PW6]: Which industry do you refer to? Be general "Oil spill refers to the intentional or nonintentional release of crude oil or its fractions to the environment".

Comment [PW7]: Add REF

Comment [PW8]: What? Do you mean the effects?

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bioavailability of nutrients to plants. These arises from are linked to reduced reduction of
porosity (REF) of the soil to both aeration and moisture, severe effect leads to reduction in
and reduces soil microbial population (REF) in presence of pollutants.

Plants exist as complex microcosm primarily exploited by a variety of living things. 44 The association between plants and microbes within a region have over the years remained 45 poorly explained, vague and mirage to a number of researchers (Santoyo, Moreno-hagelsieb, 46 Orozco-mosqueda, & Glick, 2016). Some peer review articles have identified a feasibility in 47 48 the mutualistic interaction between plants and MO for successive adaptation (REF) and other participating niches (REF). However, plants produce carbohydrates, few of nitrate free 49 compounds, these serves as nutrient for microbes which metabolising nutrients and ease 50 absorption by plants. The synergyies-in the interrelationships between of microbes and plants 51 have been reported to serve as both as biocontrol and growth promotion of plant (Kannan & 52 53 Sureendar, 2009). According to Mendes et al. (2013). reported that a number varied communities of bacteria exist on at the root region of plants and-where they improve seed 54 germination and viability. Ahemad, (2014) agreed to the earlier opinion that several bacterial 55 genera exist at these regions of interaction and create a balance between plants and microbes. 56 Advantages of having bacteria on the root region of the plants includes; the biogeochemical 57 cycling and adsorption, solubilization and, degradation of nutrients as growth factors to 58 plants. Group of bacteria (rhizobacteria) that adhere to the root (rhizobacteria) have been 59 associated with crop yield and resistance to pest and diseases. Rhizosphere is a narrow region 60 around a plant root, controlling both physicochemical and biochemical conditions. They 61 Roots of plants serve as both provide-anchorage systems, play conductive functions, nesting 62 and protective function for soil organisms (Santoyo et al., 2016). This parlance is used to 63 64 refer to organisms that existed and tolerated exudates from plants and played key role to the plant is were described as rhizospheric organisms also known as rhizobacteria. They Soil 65 66 microorganisms are competent in colonizing colonizers of the rhizosphere of plant (REF). This is because the Plant exuduates which are secretions synthesized from plants and contains 67 68 a wide array of organic substances which categorizes an exudate to be an attractant or repellent_(REF). Some are high and low molecular weight which have been described to 69 70 influence plant reproductive health and timing of flowering in plants (Lu et al., 2018). This research was designed to assess the microbiological and physicochemical qualities of 71 different weeds obtained from crude oil polluted soil within Bodo Rivers State, Nigeria. 72

73 Materials and Methods

74 Study area

75 Goi is a community in Bodo, while Bodo is a locality in the heart of Niger Delta southern

76 Nigeria with about 49,000 inhabitants and 35 villages (Obiukwu, 2015). Bodo community is

situated in Gokana, one of the kingdoms that make up Ogoniland and a LGA in Rivers state.

78 The people of Bodo are predominantly farmers and fishermen/women. The community hosts

79 Shell Petroleum Development Company (SPDC) Trans-Niger pipelines which devastated in

80 2008 and 2009 by two large oil spills. The spills affected thousands of hectares of mangroves,

fishing populations and also the livelihoods of occupants of the community. The study

Comment [PW10]: Give reference articles (Rggggg et al.,; Rggggg et al.,)

Comment [PW11]: What do you mean? Use elaborative sentences. Otherwise your audience will not understand what you need to explin Comment [PW12]: Rephrase this sentence

Comment [PW13]: Plant......? Growth, wilting, senescence Complete the sentence to bring meaning

Comment [PW14]: Let definition of a term appear before you use it. "Rhizosphere"

Comment [PW15]: Which parlance do you refer to? There is no flow in the work

Comment [PW16]: Hanging sentence

Comment [PW17]: Why are these weeds a choice? Cyperus esculentus, Scleria pauciflora.,Asystasia gangetica Harungana madagascariensis, Ancistoclaudus tectorius, Kyllinga erecta Cinna arundinacea, Brassica chinensis, Cyperus difformis, Kyllinga bulbosa and Brachiaria mutica

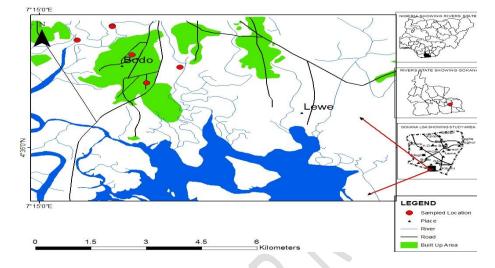
Comment [PW18]: At this point explain what interests you to the selected weeds. How important are the selected weeds? Explain earlier research done on the weeds that your results will be analysed against to show case.

Comment [PW19]: Is community the objective? Consider to mention only locality and activities in the locality. Community is mobile unlike the locality.

Comment [PW20]: Are you investigating a community? Why use community? Instead of human community, I suggest that you mention distribution of the weeds in the different localities as the weeds are of interest to the research rather that human community or inhabitants

- 82 location is known as Bodo creek and situated within the geographical grid of 4^0 37'0" North,
- 7^0 16'0''East. Other comparative plant and rhizosphere soil samples were obtained from a
- 84 pristine location in University of Port Harcourt and Tank sludge treatment farm site at NNPC,
- 85 Alesa, Eleme, Rivers State.

88



87 Figure 1: Geo-map of the sample collection points in Bodo, Gokana-Ogoni, Rivers State

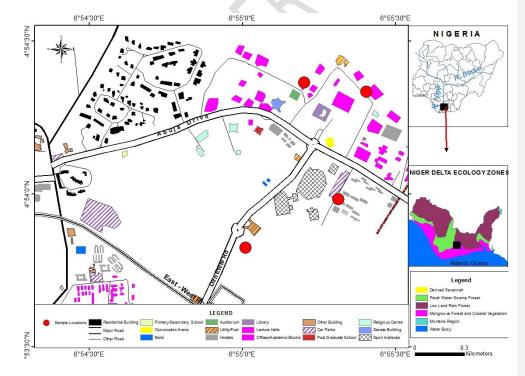


Figure 2: Geo-map of sample collection points in University of Port Harcourt, Rivers StateNigeria

91 Sample collection: Collection of samples

Plant samples were harvested from the polluted soil, wrapped in a sterile container, sealed
and labelled. The soil from the point of collection of plants were obtained using a <u>sterile hand</u>
<u>soil auger to a depth of 0 to 15 cm</u>. Soil samples were labelled with tally on the plants. The
samples were transported using a freezing chest to the laboratory in a freezing chest. The
plants were deposited at the University of Port Harcourt Herbarium for identification.

97 Baseline physicochemical and geotechnical characterization of rhizosphere samples

The physicochemical analysis of rhizosphere soil samples were carried out to, the parameters 98 analysed were analysis pH, alkalinity (APHA2320B), Electrical conductivity (APHA 2510B), 99 100 Salinity (APHA 2520B), Phosphate (APHA 4500PC), Nitrate (ASTM D-3867), Ammonia (APHA 4500), Moisture content (ASTM D2216), Phenol (EPA 604), heavy metals (Ni, Zn, 101 102 V, Fe and Cr). Geotechnical analysis includes soil texture (ASTM D422), Specific Gravity 103 (ASTM D854), Atterberg's limits (liquid, plastic and plasticity limits) (D4318). Plasticity 104 description (ASTM, D2434) while particulate size description was determined using sieve 105 method

106 Enrichment of samples

The soil samples were enriched with Bushnell Haas Media (Lab M) was prepared according 107 to manufacturer's instruction. A measure of 3.2 g of the salt was dissolved in one (1) a Litre 108 109 of distilled water, pH of the media was adjusted using 1.0 M HCl to pH 7.2. About 98 ml of 110 BHM was dispensed in a 250 ml conical flask to create room for adequate headspace, 1 % Bonny Light crude oil (BLCO) was introduced to the media and sterilization was monitored 111 112 <u>done</u> at $121^{\text{p}C}$ for 15 minutes at 15 psi. Upon cooling, the 1^{m} crude polluted pond water samples was added to the sterile set up and with the aid of an orbital shaker incubator 113 (Stuart, Germany S150) the samples were shaken at 170 r.p.m at 37[°]C (Ekwuabu, Chikere, & 114 115 Akaranta, 2016).

116 Total fungal and heterotrophic bacterial count.

Twenty-eight grams of nutrient agar was dissolved in one (1) litre, pre-heated and sterilized 117 118 at 121°C for 15 minutes and 15 psi, the media was allowed to cool to room temperature, 62.5 119 g/100_ml of nystatin was seeded to the media to inhibit the growth of fungal contaminants. 120 The samples were dissolved in a pre-sterilized normal saline. Colonies range of 3-300 cfu/ml were counted and adjudged good and results lower than the benchmark were repeated for 121 accuracy. Fungal counts were determined by plating 0.1 ml of the diluted sample on 122 Ssaboraud Ddextrose Aagar fortified with 0.1% lactic acid. Spread plate technique was 123 124 employed and plates were incubated at room temperature $(28^{\circ}C)$ for 7 days. Result for 125 fungal count were determined after B days of incubation (Peekate & Abu, 2017; Abu & Ogiji, 1996). 126

Comment [PW21]: Was there any replication? Mention it at this point

Comment [PW22]: Which is this? Comment [PW23]: Is this the same as the soil sample

Comment [PW24]:

Comment [PW25]: Is this media meant for enrichment of soil? Clarify this statement. If is a modification you did or manufacture instructs to enrich soil?

Comment [PW26]: Check this spacing and apply in while document. I.e. 3.2 g but not 3.2g

Comment [PW27]: Use the correct sign for degree. This is not a degree sign. Apply in all areas as required.

Comment [PW28]: What is the essence of adding this if you already added it earlier? Clarify purpose of adding more crude oil. What will be the final concentration of crude oil in the test.

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Comment [PW29]: Good. Do this in earlier paragraph. See comments

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129 Hydrocarbon utilizing bacterial count

130 Bushnell Haas Mmedium was prepared by dissolving 3.2g/L, fortified with 15 grams of agar, 131 the media was preheated and allowed to cool. About 1.0% One percent of lactic acid was introduced added into the media to inhibit fungal contaminants, the prepared media was 132 autoclaved along with other materials. Vapour phase culturing technique was adopted, pre-133 sterilized Whattman filter paper was placed in the lid of the petri dishes. The plates were 134 135 incubated at 37°C for 48 hours. Hydrocarbon utilizing fungal count was determined by 136 seeding 100 µg/100ml chloramphenicol for inhibition of bacterial contaminants. About 0.1ml of the sample was spread plated after dilution, crude oil impregnated filter papers were placed 137

on the lid of the plates and were incubated at room temperature (Orhorhoro et al., 2018:
Ekwuabu, Chikere, & Akaranta, 2016).

140 Identification of microbial isolates obtained from the study

141 Bacterial isolates were identified using method described by Cheesbrough (2000). These 142 battery procedures were used to ascertain the tentative identity. Isolates of fungi were

143 identified using the method described by Frazier and West Hoff (2000) macroscopic and

144 microscopic Atlas and reference to standard identification keys.

145 Statistical analysis

The data obtained was analysed using statistical package for Social Science version 23.0physicochemical components were analysed using One-way ANOVA. Output data was

148 compared using homogenous subset at p-value < 0.05.

149 RESULT

150 Table 1 describe baseline physicochemical composition of rhizosphere soil obtained from weeds. pH of the soil ranged between 5.26-7.2. The pH of the soil rhizosphere soil obtained 151 152 were, Ancistoclaudus tectorius was 5.26, Brassica chinensis 5.4, C. esculentus was 6.9, Kyllinga erecta was 5.9. The highest pH 7.2 was observed for Asystasia gangetica. 153 154 Temperature of the samples were within 26.3 to 31.6 °C; Brassica chinensis was 31.6 °C while *Cinna arundinacea* was 30 $^{\circ}C$ While the lowest temperatures was recorded for 26.3 $^{\circ}C$ 155 156 for C. esculentus, But Scleria pauciflora and Harungana madagascariensis had a 157 temperature of 27 $^{\circ}C$. Refinery effluent had a conductivity of 5520.9 μ s/cm and *Brassica* 158 chinensis was 400.5_us/cm. Rhizosphere soil obtained from Bodo, Gokana were 80.31 µs/cm and 53.4 µs/cm was reported for Cyperus esculentus and Kyllinga erecta. Phosphate was 159 lowest with soil obtained from Brassica chinensis which was 0.74 mg/kg and 5.4 mg/kg 160 161 phosphate was reported for Cyperus esculentus. Heavy metals like Nickel was below detectable level for most weeds in Bodo, while *Brassica chinensis* had nickel concentration 162 of 1.11 mg/kg. The level of vanadium was 0.71 and 0.61 for Kyllinga erecta and Cinna 163

Comment [PW32]: Avoid this. Be specific.
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| Comment [PW33]: Is it from the weeds or soils surrounding the place of growth of weeds? Check in your method of collection of soil. You used an auger. Clarify and be consistent. |
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Comment [PW35]: Do you mean soils or weeds? Confirm this

arundinacea. The heavy metal (Nickel, Zinc, Vanadium, Lead, Iron and Chromium) werenot detected in the control samples (Table 2).

167 Table 1: Baseline Physicochemical composition of rhizosphere soil obtained from plants in

168 Ogoni, Rivers State

| | Rhiz | osoil (pol | luted soil H | Bodo) | | | | | |
|----------------------------|--------------------|---------------------|---------------------|-------------------------------|-----------------------------|-----------------|------------------|----------------------|---|
| Baseline Parameters | Cyperus esculentus | Scleria pauciflora. | Asystasia gangetica | Harungana madagascariensis | Ancistoclaudus tectorius | Kyllinga erecta | Cima arundinacea | Refinery Effluent | Formatted: None, Indent: Left: 0.08", Right: 0.08", Space Before: 0 pt, Line spacing: single, Don't keep with next, Don't keep lines together Formatted Table |
| pH Temperature | 6.9 26. | 6.9 27.0 | 7.20 28.0 | 6.50 27.0 | 5.26 29.0 | 5.90 28.7 | 6.3 30.0 | 6.98 27.2 | 5.4 31.6 |
| (°C) Conductivity | 3 53. | 40.2 | 48.6 | 32.1 | 11.5 | 80.3 | 11.3 | 5520. | Formatted: Highlight 400.5 |
| (µS/cm) | 4 | | | | | | | 9 | |
| Oil and Grease (mg/kg) | 90. 3 | 74.6 | 61.4 | 113.2 | 100.5 | 67.9 | 73.4 | 237.1 | 111.6 |
| Salinity (ppt) | 5 68. 8 | 70.7 | 60.0 | 50.6 | 42.7 | 52.4 | 88.7 | 229.8 0 | 61.5 |
| Alkalinity | 18. 8 | 16.1 | 22.1 | 12.1 | 11.17 | 15.43 | 67.5 | 19.2 | 12.7 |
| Phosphate(mg/ kg) | 8 5.6 | 3.10 | 1.8 | 2.1 | 3.0 | 4.1 | 5.3 | 3.69 | 0.74 |
| Ammonia (mg/kg) | 2.1 4 | 1.30 | 1.0 | 0.50 | 1.10 | 2.0 | 1.95 | 1.98 | 1.32 |
| Phenol (mg/kg) | 90. 3 | 74.6 | 61.4 | 50.4 | 34.5 | 26.5 | 110.8 | 76.9 | 15.3 |
| Hydrogen sulphide(mg/kg | 12. 4 | 11.5 | 11.7 | 20.7 | 31.6 | 19.9 | 43.3 | 51.3 | 20.2 |
|) Nickel (mg/kg) | BD I | BDL | BDL | BDL | BDL | BDL | 0.02 | 0.08 | 1.1 Comment [PW37]: Define this in footer of table |
| Zinc (mg/kg) | 0.1 | 0.10 | 0.32 | 0.6 | 1.5 | 1.7 | 3.34 | 0.12 | 5.8 |
| Vanadium (mg/kg) | 4 0.0 4 | 0.01 | 0.07 | 0.01 | 0.31 | 0.71 | 0.7 | 0.18 | 0.01 |
| (mg/kg) Lead (mg/kg) | BD L | 0.1 | 0.18 | 0.2 | 0.01 | 0.05 | 0.02 | 0.08 | 0.09 |
| Iron (mg/kg) | L 0.2 6 | 0.14 | 0.09 | 0.2 | 0.20 | 0.33 | 0.01 | 0.01 | 0.63 |
| Chromium (mg/kg) | 0.0 8 | 0.04 | 0.02 | 0.01 | 0.05 | 0.021 | 0.04 | 0.08 | 0.02 |
| Sulphates(mg/k | 8 4.8 | 5.8 | 9.4 | 3.3 | 1.9 | 10.8 | 15.4 | 3.0 | 6.45 |
| g) | | | | | | | | | Comment [PW38]: Apply abbreviations Zinc is "Zn" |

171 Table 2: Physicochemical composition of rhizosphere soil obtained from a pristine location

172 University of Port Harcourt

| | | Rhizosphere soil (Uniport) | |
|-----------------------------|----------------------|-------------------------------|-----------------------|
| Baseline Paramete rs | Cyperus difformis | Kyllinga bulbosa | Brachiar ia mutica |
| рН | 6.9 | 5.90 | 7.8 |
| Temperature (°C) | 29.3 | 31.7 | 28.0 |
| Conductivity (µS/cm) | 103.4 | 80.31 | 212.32 |
| Oil and Grease (mg/kg) | 40.3 | 17.9 | 8.4 |
| Salinity (ppt) | 21.8 | 12.4 | 15.7 |
| Alkalinity | 11.8 | 4.7 | 3.8 |
| Phosphate(mg/kg) | 9.6 | 11.1 | 7.7 |
| Ammonia (mg/kg) | 2.14 | 2.0 | 1.95 |
| Phenol (mg/kg) | 4.5 | 3.11 | 5.2 |
| Hydrogen sulphide(mg/kg) | 1.4 | 1.0 | 7.3 |
| Nickel (mg/kg) | BDL | BDL | BDL |
| Zinc (mg/kg) | BDL | BDL | BDL |
| Vanadium (mg/kg) | BDL | BDL | 0.01 |
| Lead (mg/kg) | BDL | 0.05 | BDL |
| Iron (mg/kg) | 1.26 | 1.91 | 1.5 |
| Chromium (mg/kg) | BDL | 0.006 | BDL |
| Sulphates(mg/kg) | 1.06 | 1.8 | 3.1 |
| TPH (mg/kg) | | | |

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Comment [PW39]: Use abbreviations. See earlier comment

173

Table 3 describes the geotechnical evaluation of the soil sample obtained from the from the 174 pristine soil from uniport the herbarium in UNIPORT. The result showed that the soil hadsoil 175 had 82.43 wt %_-sand, 14.19 wt% clay and 2.48% silt while polluted soil had 87.72% silt, 176 9.01% sand and 1.98% clay for the Bodo polluted soil Bodo-Ogoni. Moisture content for the 177 178 pristine soil obtained from Uniport was 17.82% while polluted soil was 11.75%. Permeability description of both pristine and rhizosphere polluted soil were both low and had 179 permeabilities 6.3epermeabilities of 6.3e⁻⁶ and 4.73e⁻⁶ cm/sec_respectively. Organic carbon 180 was high with the polluted soil with 31.85%. Plasticity of the soil was observed to be 7.1 and 181 182 8.9 and were reported to have low plasticity.

| Parameters | Pristine soil (Uniport) | Pristine soil (Goi, Bodo) | Rhizosphere Polluted soil | different types which has significant influence on rhizosphere biota. What was the objective of testing the weeds in different kinds of soils? If they were similar weeds subjected to different kinds of soils it would be better. This compromises the whole |
|-----------------------------------|----------------------------|------------------------------|------------------------------|--|
| Sand (wt %) | 82.43 | <mark>26</mark> | 9.01 | results as is already known to be different. |
| Clay (wt %) | 14.19 | 40.7 | 1.98 | Formatted: Highlight |
| | | | | Formatted: Highlight |
| Silt (wt %) | <mark>2.48</mark> | <mark>39.79</mark> | <mark>87.72</mark> | Formatted: Highlight |
| Soil type | Loamy sand | Clay Loam | Silt loam | Formatted: Highlight |
| Moisture content (%) | 17.82 | 11.52 | 11.75 | |
| Permeability (cm/sec) | 2.23E-03 | 6.30E-06 | 4.73E-06 | |
| Permeability description | Moderate | Low | Low | |
| Organic carbon (%C) | 31.85 | 3.81 | 129.63 | |
| Fotal Organic Carbon (mg/kg | 11.48 | 22.79 | 300.09 | |
| Total Hydrocarbon content (mg/kg) | 5.8 | 9.1 | 95 | |
| Liquid limit (%) | 27.41 | 20.52 | 18.73 | |
| Plastic limit (%) | 18.51 | 17.93 | 13.35 | |
| Plasticity index | 8.91 | 9.0 | 7.1 | |
| Plasticity description | Low | Low | Low | |
| Particulate size distribution | Silt(0.02 mm) | Silt(0.02 mm) | Silt(0.02 mm) | |
| | Clay (0.002mm) | Clay (0.002mm) | Clay (0.002mm) | |
| | Sand (2.00mm) | Sand (2.00mm) | Sand (2.00mm) | |

184 Table 3: Geotechnical qualities of soil samples obtained during the study.

185

Figure 3 describes the microbial population monitoring of the soil samples obtained from the 186 187 rhizosphere of weeds. The study revealed that Total heterotrophic count for count for the control samples were significantly (p < 0.05-) different from the soil obtained from the 188 rhizosphere of plants. The results revealed that 7.5 Log₁₀Cfu/g to 7.77 Log₁₀Cfu/g, for Rz4, 189 A. tectorius had a 5.11 Log₁₀Cfu/g while A. gangetica had a TAHC of 6.38 Log₁₀Cfu/g. 190 Kyllinga bulbosa had a population of 6.7 Log₁₀Cfu/g for TFC, 4.76 Log₁₀Cfu/g HUFC 191 192 while HUBC was 5.16 Log₁₀Cfu/g. Soil samples obtained from S. pauciflora, had HUB and 193 THBC of 5.79 Log₁₀Cfu/g and 6.04 Log₁₀Cfu/g. Crude oil polluted water had 6.9 Log₁₀Cfu/g, for THBC, 5.56 Log₁₀Cfu/g, while the total fungal count was 5.36 Log₁₀Cfu/g 194

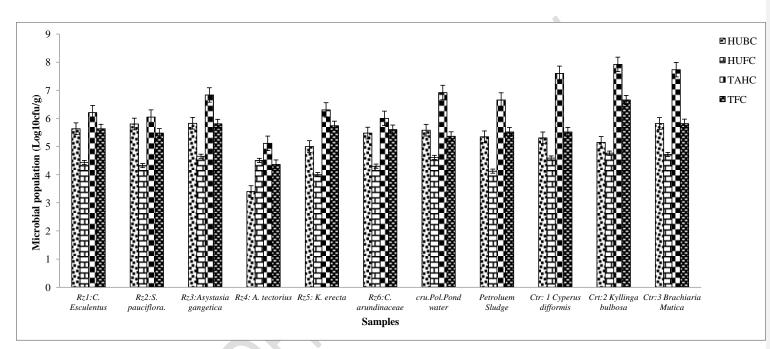
195 Table 4 represents microbial evaluation of the soil samples obtained from weeds of Bodo 196 polluted soil, Cyperus esculentus rhizosphere soil was dominated with Achromobacter sp, B. lichenformis, B. anthracis, B. subtilis, B. fumari, Arthrobacter sp, Pseudomonas sp, Ps. 197 198 aeruginosa, Ps. florescens, Fungal isolates associated with plant samples were presented in Table 5. Organisms revealed includes Aspergillus terreus, Trichoderma sp, and Fusarium sp. 199 200 S. pauciflora had Micrococcus sp, B. cereus, B. subtilis and Pseudomonas sp while A. niger, 201 Mucor sp, Fusarium sp and Penicillium sp are the fungi isolated. Refinery wastewater was 202 reported to harbour Salinococcus sp. Staphylococcus sp. and Micrococcus sp. while fungal 203 isolates are Monilia sp. and Prunius sp. (Tables 5, 6 and 7).

Comment [PW41]: This is definitely different as concern the soil types.

Comment [PW40]: These soils are definitely of

Comment [PW42]: You definitely needed separate control for each soil type.

UNDER PERMIT



²⁰⁸ Legend: HUBC= Hydrocarbon utilizing bacterial count; HUFC= Hydrocarbon Utilizing Fungal count, TAHC= Total heterotrophic

211 pristine environment

212

<sup>Bacterial count; TFC= Total Fungal Count
Figure 3: Average microbial population of Rhizosphere soil, pond water, petroleum sludge obtained from plants pre-exposed to crude oil and</sup>

| | ΓO | Ċ | ы Н | | | 2 | | | | | Ver | 101 | | | | ~ | ~ | | | | | | | | | Probable |
|-----------------|------------------|---------------------------|----------------------|---------|---------|----------|----------|-----|-------|------|--------|-----|----|----|---------|---------|---------|---------|-----------|--------|----------|---------|-----------|----------|-----------|----------------------|
| ISOLATE CODE | GRAM MORPHOLO | GRAM GRAM MORDITOLO | MUKPHULU CATALASE | OXIDASE | CITRATE | MOTILITY | Glycerol | GAS | SLANT | BUTT | STARCH | | MR | VP | GluCOSE | LACTOSE | SUCROSE | Maltose | Arabinose | xylose | Mannitol | Salicin | Trehalose | Sorbitol | Galactose | isolates |
| 1 | Rod | + | + | + | - | + | + | - | Κ | А | + | - | - | - | Ag | - | - | + | A/g | - | A/g | - | А | A/g | | Arthrobacter sp. |
| 2 | Rod | + | + | + | + | - | + | - | Κ | Κ | + | + | + | - | A/g | - | - | - | A/g | А | - | А | - | - | А | B. anthracis |
| 3 | Rod | + | + | + | + | - | - | - | А | Α | + | - | + | - | A/g | - | А | A/g | - | A/g | - | - | А | - | А | B. subtilis |
| 4 | Rod | - | - | - | + | + | A/g | - | А | А | + | - | + | - | Ag | Ag | Ag | A/g | - | А | А | A/g | - | - | А | Pseudomonas sp. |
| 5 | Cocci | + | + | + | + | - | - | - | Κ | А | + | - | + | - | A/g | A/g | A/g | - | - | - | - | A/g | - | - | - | Salinococcus sp. |
| 6 | Rod | + | + | - | + | + | - | + | Κ | А | - | - | - | - | A/g | - | - | | - | - | Α | - | - | - | А | Achromobacter sp |
| 7 | Rod | - | + | + | + | + | - | + | А | Α | - | - | - | + | A/g | A/g | A/g | А | А | A/g | A/g | А | A/g | A/g | A/g | Ps. Fluorescence |
| 8 | Rod | - | + | - | - | + | - | - | А | Κ | + | - | + | - | A/g | Ag | A/g | - | - | Α | - | - | A/g | - | А | Pseudomonas sp |
| 9 | Rod | + | + | - | - | + | - | - | А | А | + | - | - | - | A/g | A/g | A/g | А | А | A/g | А | А | - | А | - | Bacillus. sp. |
| 10 | Rod | + | + | + | - | - | - | + | А | А | + | + | + | - | A/g | - | Ag | - | А | А | - | A/g | А | - | А | Bacillus cereus |
| 11 | Rod | + | - | + | + | - | - | - | Κ | А | - | - | + | - | A/g | A/g | A/g | - | - | А | - | - | А | - | А | Clostridium sp |
| 12 | Rod | - | + | + | - | + | - | + | А | А | + | + | + | - | A/g | A/g | Ag | A/g | A/g | A/g | А | А | A/g | А | A/g | E. coli |
| 13 | Rod | + | + | - | - | + | - | + | Κ | А | - | - | + | + | А | - | А | А | - | - | - | А | А | - | А | Bacillus thuriiensis |
| 14 | Cocci | + | + | + | - | + | - | - | А | Κ | - | - | - | - | A/g | - | A/g | А | А | - | - | А | - | А | А | Staphylococcus sp. |
| 15 | Cocci | + | + | + | - | - | - | - | Κ | А | + | + | - | - | А | - | А | - | - | - | - | А | | А | - | Micrococcus sp |
| 16 | Rod | + | + | + | - | - | А | - | Κ | Κ | - | - | - | - | А | - | А | - | А | А | - | А | - | - | А | Paenibacillus sp |
| 17 | Rod | + | - | - | + | - | - | + | K | Κ | - | - | - | + | А | - | А | - | A/g | А | - | А | А | - | A/g | B. lugardi |
| 18 | Rod | - | | - | + | + | + | + | А | А | + | + | - | - | A/g | А | A/g | - | - | - | - | А | - | - | - | Klebsiella sp. |

213 Table 4 Biochemical characteristics of bacterial Isolates from both pristine and impacted rhizosphere soil

214 Key:+=positive; -=Negative, A=Acid formation; K= Alkaline; A/g= Acid formation and gas production; A= Acid formation alone

215 MR=Methyl Red, VP= Vogues Poskauer test

- 216
- 217
- 218
- 219

| Rz1:Cyperus esculentus | Rz2:Scleria pauciflora. | Rz3:Asystasia gangetica | Rz4:Harungana madagascariensis. | Rz5:Ancistoclaudus erectus. | Rz6: Cinna arundinaceae | Rz7: Kyllinga erecta |
|--------------------------------------|----------------------------|------------------------------------|------------------------------------|-----------------------------|---------------------------------|---|
| Achromobacter sp. B. lichenformis | Micrococcus sp. | Acinetobacter sp. | B. lugardi | Klebsiella sp. | Pseudomonas sp. | B. thuringiensis |
| B. anthracis | B. cereus. | B. thuringiensis | B. subtilis | Pseudomonas sp. | Ps. aeruginosa. | B. subtilis |
| B. subtilis | B. subtilis | Paenibacillus sp. | B. thuringiensis | Achromobacter sp. | Klebsiella sp. | |
| B. fumari | Pseudomonas sp. | Micrococcus sp. | Achromobacter sp. | | | |
| Arthrobacter sp. | | | Pseudomonas sp. | Q | | |
| Pseudomonas sp. | | | | | | |
| Ps. florescence | | | | | | |
| Ps. aeruginosa | | | | | | |
| Salinococcus sp. | | | | | | |
| Rz8:Cyperus esculentus | Rz9:Kyllinga bulbosa | Rz10:Brachiaria Mutica | | Petroleum Sludge | Petroleum polluted pond | Produce water |
| esementus | ontoosu | | | B. subtilis | water | Salinococcus sp. |
| B. thuringiensis | Staphylococcus sp. | B. cereus. B. subtilis | | B. cereus. | Ps. florescence | Staphylococcus sp |
| Ps. florescence | Micrococcus sp. | \sim | | | U | |
| Ps. aeruginosa | Alcaligenes sp. | Citrobacter sp. | | Paenibacillus sp. | Ps. aeruginosa Achromobacter | <i>Micrococcus</i> sp. <i>Klebsiella</i> sp. |
| | Flavobacterium sp. | Alcaligenes sp. Pseudomonas sp. | | | sp. | Kiebsiena sp. |
| Arthrobacter sp. | | Proudomonas en | | | | |

Table 5: Bacterial isolates associated with rhizosphere of weeds obtained during the study

Table 6: Fungal microflora obtained from rhizosphere region of plants pre-exposed to pollution

| Sample source | Total fungal flora | Probable Identity | Hydrocarbon utilizing fungal flora | Probable identity |
|--|---|---|--|---|
| Rz1: C. esculentus | a) Suded- army green, grey rough reverse side; b) Whitish-suede dense mycelia. Brown reverse side c) Fluffy-white with a ring and raised centre | a)——Aspergillus terreus b) Trichoderma sp. c)Fusarium sp. | a)Wooly-white hairlike mycelia b) Green rough surface, brown reverse side | a) Fusarium sp. b) Aspergillus flavus |
| Rz2: S. pauciflora | with salt crystals a) White mycelia with a black tips covering at the centre b) Dull-leaf green surface with venation c) Fluffy-white with a ring and raised centre with salt crystals | a)Aspergillus niger b)Penicillium sp. b)Fusarium sp. | a)Smooth green surface fungi b) Fluffy-white with a ring and raised centre with salt crystals b)White fluffy, no | Penicillium sp. Fusarium sp. Mucor sp. |
| Rz3: Asystasia gangetica Rz4: A tectorius | a)Creamy smooth growth and rough depressed centrea)Whitish flat mycelia with a circular ring.b)white dense mycelia and spots of liquid crystals | a) <i>Candida</i> sp. a) <i>Prunius</i> sp. b) <i>Monilia</i> sp. | colour at the reverse side a)Creamy smooth fungi. Rough flat – bacterial-like growth | a) <i>Candida</i> sp. a)Rhodotorula sp. |
| Rz 5: K. erecta | a)Tiny brown-raised mycelia with a cream, rough reverse side | Cladosporium sp. | a) Tiny brown- raised mycelia with a cream, rough | a) Mucor sp. |
| Rz6: C arundinacea | b) white Hair-like growthwhite dense mycelia and spots of liquid crystalsFluffy-white with a ring and | Mucor sp. Monilia sp. Fusarium sp. | reverse sidea) Fluffy-whitewith a ring andraised centre withsalt crystals.b) Creamy smooth | b) Fusarium sp. b) Candida sp. |

225 Table 7: Fungal microflora of rhizobacterial flora of plants on pristine soil.

| Sample source | Total fungal flora | Probable Identity | Hydrocarbon utilizing fungal flora | Probable identity |
|---------------|---|--|---|---|
| Ctr1: | a)Suded- army green, grey rough reverse side; b)Tiny brown- raised mycelia with a cream, rough reverse side c)Fluffy-white | a) <i>Penicillium</i> sp b) <i>Cladosporium</i> sp. c) <i>Fusarium</i> sp | a)Fluffy-whitewith a ring andraised centrewith salt crystalsb)White myceliawith a black tipscovering at thecentre. | a)Fusarium sp b) <i>Aspergillu</i> <i>niger</i> |
| | with a ring and raised centre with salt crystals d)White mycelia with a black tips covering at the centre | d)Aspergillus niger | | |
| Ctr2: | Bright leaf- green round colony, with | Penicillium sp. | Smooth, raised, mucoid growth | <i>Rhodotorula</i> sp |
| | veneations Round raised white Hair-like growth | <i>Mucor</i> sp | Round raised white Hair-like growth | <i>Mucor</i> sp |
| Ctr3 | white dense mycelia and spots of liquid | a) <i>Monilia</i> sp. | No growth | No growth |
| | crystals Whitish flat mycelia with a circular ring. | b)Prunius | | |
| | | | | |
| | | | | |
| | | | | |

230 Discussion

Phytodiversity of polluted environment is reflective of the history of devastation on the
ecosystem, loss in biodiversity, geotechnical and physicochemical qualities. Adaptation ofto
plants have has been identified as a measure of adaptability to tolerate pollutant. Orhorhoro et
al., (2018) identified *Schoenopletus senegalensis*, *Fuirena umbellate* and *Cyperus tuberosis*Edwin-wosu, (2016) reported a vast number of plant species in pristine environment in Rivers
State. These separate accounts agrees with the report of the present study; *Cyperus esculentus, Scleria pauciflora, Asystasia gangetica, Harungana madagascariensis*,

238 Ancistoclaudus tectorius, Kyllinga erecta, Cinna arundinacea, and Brassica chinensis.

Physicochemical attributes of the soil samples obtained from rhizosphere regions of weed 239 serves as eco-indicators of niches and could further describe the quality of bio-activities 240 241 withinactivities within the region of the soil. pH of the rhizosphere soil during the study was 242 observed to be slightly acidic and temperature mesophilic, B. chinensis had a pH 5.4 and a temperature of 31.6[°]C, samples obtained from C. esculentus had a pH 6.9 and 26.3[°]C. These 243 244 findings corroborated the earlier report of Wang et al.(2013) that the temperature of pristine soil should be lower than that of the polluted soil had pH and temperature of 6.9 and 29.3 $^{\circ}$ C 245 246 respectively. However Ofoegbu (2015) reported a pH 6.37 and 28 °C in their separate 247 investigation. This is because crude fraction could conduct heat and energy. The presence of long-chain and persistent hydrocarbon fraction as well could also have a low degradation 248 process and impact on the pH of the environment. Alkalophilic and mesophilic environments 249 250 could encourage the synthesis of enzymes and bioavailability of nutrients (Olowomofe, Oluyege, & Sowole, 2017). Electrical conductivity (E.C) is a measure of residual ions, 251 radicals and polarity. In this study, it ranged from 11.32- 80.3 µS/cm. Which tallied with the 252 253 report of Ekwuabu et al. (2016) in whose report E.C was 12.0 µS/cm for a pre-impacted soil. The polarity could impact the porosity of the soil, thereby retarding the flow of nutrients and 254 255 water. Rhizodeposites affects the quality of conduction and ease the passage or flow of nutrients, the variation could arise from the deposits and leaching activity caused by the 256 257 pollutant. One of the limiting nutrients that retards growth is phosphorus, as it ranged from 258 0.74-5.6 mg/kg. Phosphorus and phosphates aid absorption of nitrates in microbiome. They 259 could be easily washed off by run-offs and seepages. It could also be affected by seasonal 260 variations (Wang et al., 2013), although they reported 13.9 mg/kg, this was in agreement with 261 the position of this study.

262 Incidence of oil spill in Niger Delta have caused devastating damages to arable lands in the region. Microbiological qualities of the soil revealed a steady decline from the microbial 263 264 indices, the range revealed that 7.5 $Log_{10}Cfu/g$ to 7.77 $Log_{10}Cfu/g$. The result for Rz4 A. 265 tectorius had a 5.11 Log₁₀Cfu/g while samples obtained from A. gangetica had a TAHC of 266 6.38 Log₁₀Cfu/g. Kyllinga bulbosa rhizosphere soil had a population of 6.7 Log₁₀Cfu/g for TFC, 4.76 Log₁₀Cfu/g HUFC while HUBC was 5.16 Log₁₀Cfu/g, while Soil samples 267 268 obtained from S. pauciflora, had HUB and THBC of 5.79 Log₁₀Cfu/g and 6.04 Log₁₀Cfu/g. 269 Crude oil polluted water had 6.9 Log₁₀Cfu/g, for THBC, 5.56 Log₁₀Cfu/g, while the total 270 fungal count was 5.36 Log₁₀Cfu/g, Ekwuabu et al.(2016) reported THB of 7.89 Log₁₀Cfu/g. 271 Furthermore Olowomofe et al. (2017) reported 5.3-7.9 Log₁₀Cfu/g for polluted soil in Bodo, **Comment [PW43]:** Do you mean adoption or what? Check meaning of this sentence.

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Ogoni. The level of microbial interaction could be used as predictive component pollutionmonitoring and control.

274 Bacterial diversity inof the soil obtained from rhizosphere region of the weeds were 275 documented from the study. The result suggestsuggests the dominance of Bacillus sp and Pseudomonas sp from the soil with presence of Achromobacter sp, B. lichenformis, B. 276 anthracis, B. subtilis, B. fumari, Arthrobacter sp, Pseudomonas sp, Ps. aeruginosa, Ps. 277 Florescens. Fungal isolates associated with samples were Aspergillus terreus, Trichoderma 278 279 sp, and Fusarium sp. The result corroborates with the report of Olowomofe et al. (2017) where they isolated bacterial isolates from tarsand with more of *Pseudomonas* sp. and 280 Bacillus sp. This corroborates with the report of Yrjälä, Keskinen, Åkerman, Fortelius, & 281 Sipilä, (2010) whose study revealed the preponderance of Bacillus sp. at the rhizosphere of 282 283 weeds. This further agrees with the report of Tesar et al., (2002) who opined showed that the 284 dominance of Gram negative microbes and few spore formers may be observed from crude oil polluted soil. The report of Omotayo et al., (2014) supports the fact that there is a level of 285 286 sociomicrobial interaction of microbes in different environmental media, plays a key feature 287 in the distribution of soil microbiota. Furthermore, Orhorhoro et al. (2018) corroborated the earlier studies and they were able to associate Arthrobacter sp., Bacillus pumilus, B. 288 sphaericus and Serratia marcescens. This study revealed the presence of Pseudomonas sp. 289 290 Corynebacterium sp., Bacillus sp. Bacterioides sp. Staphylococcus sp. Klebsiella sp. and 291 Kingella sp. However, Daane et al. (2001) reported the presence of Flavobacterium, 292 Pseudomonas putida and Mycobacterium sp. Ukaegbu-Obi and Mbakwem-Aniebo, (2014) 293 reported the dominance of Flavobacterium sp and Pseudomonas sp. Van Hamme and Ward, (2001) suggested that these organisms have a selective resistance to oil interfaces, thereby 294 295 secreting an organic acid that aids degradation of hydrocarbon. The findings of this study also 296 corroboratescorroborate the report of Ukaegbu-Obi and Mbakwem-Aniebo (2014) who 297 reported the presence of Acinetobacter, Bacillus, Pseudomonas, Alcaligens and Micrococcus 298 as rhizophytes. The percentage occurrence of any group of bacterial isolate describes the 299 nature of the environment. The study revealed the predominance of Bacillus sp. and Pseudomonas sp. These bacterial isolates have been associated with degradation and 300 301 tolerance petroleum hydrocarbon fractions. (Ekwuabu et al., 2016; Olowomofe et al., 2017; 302 Yrjälä et al., 2010)

303 Conclusion and Recommendation

304 Niches within rhizosphere of plants affected by exudates and exogenous secretions from plant microbe-interaction. The quality exudates either as an attractant or repellent affects microbial 305 306 indicators of niches. Rhizosphere soil from *Cyperus esculentus* had a higher species diversity. 307 Bacterial load was observed to decline. pH of most soil samples were slightly acidic and 308 hence encouraged a narrow range of fungal isolates. Geotechnical considerations suggest total organic carbon, plasticity and porosity of the soil samples were low and were affected 309 310 by the pollutant. Pseudomonas and Bacillus sp were the most dominant bacterial isolates 311 while Aspergillus sp., Fusarium sp. and Penicillium sp.were the most dominant microbes at 312 the rhizosphere region of weeds.

Comment [PW44]: Opined? Is it an opinion? Dic he conduct research to make it factual? Check application of word opined to support research findings

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Comment [PW45]: Ckeck application of word "However"

Comment [PW46]: Rephrase to bring meaning

Comment [PW47]: Did this study test the types of plant exudates? No. pollution was the key target. So conclude on impact of pollution but not exudates. Or link the exudates to crude oil. Is there any synergistic effect or are there suppressant effects?

Comment [PW48]: It is not in order to conclude this because the soil type effect also contributes to this difference. You need to address this anomaly.

Comment [PW49]: Where? Hanging sentence

Comment [PW50]: Too general. Give the specific sites with low pH

314 Recommendation

Rhizobiology and niche-indices of impacted soil could represent a whole new perspective in
biomining of high throughput strains for biotechnological development in Nigeria. Weeds in
Bodo, Ogoniland could harbour countless families of functional and degradative bacterial
communities which could be veritable roles in the clean-up of the pollutants in the Niger
Delta.

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Comment [PW51]: Recommend on your findings but not what you think

313

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- 377

Comment [PW52]: Confirm all references to be in text and the reference section