

Original Research Article

Utilization of Telang Leaf Meal as Antioxidant Source in Feed of Tilapia (*Oreochromis niloticus*)

ABSTRACT

Aims: This study aimed to determine the effective percentage of telang leaf meal on feed to increase antioxidant levels in tilapia liver feed.

Study design: A total of 12 aquariums with sizes 60 x 40 x 40 cm³ and volume 96 L prepared and filled with 30 L water. Heater installed and raised the temperature slowly until 34°C. Feed in the form of a mixture of telang leaf flour and commercial feed given 3 times a day according to treatment and provision with provisions 3% of body weight of test fish per day. Analysis of antioxidant levels is seen from the content of the enzyme superoxide dismutase as an antioxidant bioindicator in the body of the test fish.

Place and Duration of Study: Breeding of fish is carried out at Hatchery Ciparanje Inland Fisheries Area, Faculty of Fisheries and Marine Science, Universitas Padjadjaran and Dismutase Superoxide Level Analysis was carried out at the Central Laboratory of Universitas Padjadjaran in June and July 2019.

Methodology: This research was carried out by giving high temperatures as a source of stress, during 40 days. Tilapia were given a mixture of telang leaf meal as a source of antioxidants. At the end of research, Test liver fish taken superoxide dismutase levels were observed and analyzed descriptively.

Results: By giving heat stress continuously for 40 days the result is obtained the highest average superoxide dismutase level in treatment B (telang leaf meal 5%) as much -0.92 inhibition and lowest in treatment D (telang leaf meal 15%). A decrease in SOD levels identifies a decrease in antioxidant activity in the body of Tilapia..

Conclusion: ~~Based on the results of research that has been done then conclusions can be drawn that~~ the addition of telang leaf meal is up to 15% in commercial feed by giving heat stress since the beginning of the culture period does not provide an effect of increasing antioxidant levels

Keywords: Antioxidants, Tilapia, Stress, Telang leaf meal

1. INTRODUCTION

Tilapia (*Oreochromis niloticus*) is a species of freshwater fish that is widely cultivated and consumed by the community [1], Because tilapia has the ability to grow fast and high adaptation. Tilapia also has high market demand, with the production volume of tilapia reaching 695 thousand tons in 2012 [2] and increased to 1,280 thousand tons in 2017 [3]. Tilapia production volume increased by 84% for 5 years. This high production can be increased by intensive or super-intensive cultivation.

In intensive aquaculture of tilapia, one challenge is stressful fish. According to [4] the existence of stressed fish is one of the causes of fish susceptible to disease. So a better method is needed for prevention of stress-related illnesses [5].

Prevention of diseases caused by stress can be done with the role of artificial feed given since fry, especially functional feed. Continuity of feed procurement is a very important factor in intensive aquaculture efforts, because the cost of feed is the greatest cost, up to more than 50 percent of total production costs [6].

Functional feed is feed besides having nutritional value, also has the ability to maintain health and control disease, as well as other positive effects on animal nutrition [7]. The use of synthetic antioxidants in feed can be bad that is, it results in the final result if consumed by humans, such as impaired liver function, intestinal mucosa, lungs, where the allowed dose is not allowed $\geq 0,1\%$ [8]. So we need a source of antioxidants from natural ingredients, examples of natural ingredients that can be used as additives, to give a functional effect is the telang plant.

Telang plant (*Clitoria ternatea*) is a plant that has a lot of pharmacological potential, and can be said to be functional feed, because it contains anthocyanin compounds which has high antioxidant activity [9]. Telang plants contain a number of phenols and flavonoids in the leaves and flowers, which plays a role in inhibiting and fighting free radicals as DPPH, hydroxyl radicals, and hydrogen [10]. The content of antioxidant activity can increase the body's ability to ward off disease and minimize stress.

Increased levels of antioxidants in counteracting free radicals can be measured by looking at the enzyme superoxide dismutase, which is widely found in the liver. The superoxide dismutase enzyme converts superoxide free radicals into oxygen and hydrogen peroxide

Testing of antioxidant activity in vivo needs to be done, bearing in mind that there is still little research into antioxidant activity in vivo especially in fish. Testing of antioxidant activity in vivo was carried out by the method of Marklund and Marklund (1974) [11] because the material is easier and easier to obtain, and can be done quickly analysis, cheap instrument, and simple operating protocol [12].

2. MATERIAL AND METHODS

2.1 Study area

This research was conducted from June to July 2019 at the Hatchery, Ciparanje Land Fisheries Area, Faculty of Fisheries and Maritime, Universitas Padjadjaran and Central Laboratory of Universitas Padjadjaran. [Need to complete](#)

~~The materials used in this study include: Tilapia, Telang Leaves, Commercial Feed PF 1000, CMC, Pyrogallol. This research was conducted in 40 days by giving heat stress from the beginning. Data on superoxide dismutase levels at the end of maintenance were compared between treatments.~~

2.1 Materials used and Research Implementation

~~The Materials used in this study include Tilapia, Telang Leaves, Commercial Feed PF 1000, CMC, Pyrogallol. This research was conducted in 40 days by giving heat stress from the beginning. Data on superoxide dismutase levels at the end of maintenance were compared between treatments.~~

First is the telang leaves picked and washed clean. Telang leaves that are taken must be overstated due to the shrinkage of fresh telang leaf weights. Drying is done by aerating for 3 days. Telang leaves are then ground into meal.

Test feed was made by mixing commercial feed with telang leaf meal (TLF) according to the treatment concentration. Commercial feed is weighed according to division feed A (control)

Comment [ct1]: Where are general and specific objectives of this study??

100% commercial feed, feed B with 95% commercial feed and 5% TLF, feed C with 90% commercial feed and 10% TLF, feed D with 85% commercial feed and 15% TLF. In the mixing process added CMC as much 1% total feed. Feed mixture is stirred evenly to form a paste. Dough that has been shaped paste is mold then air dried for 1 day.

Tilapia is first given a dowry for 1 day, after that a sampling is done to determine the initial fish weight and determine the dose of feed to be given. Tilapia included 10 fish per aquarium. Furthermore, feed is given 3 times a day at 3% of body weight. Heater is installed as a source of heat stress with initial temperature 26°C at 09.00 WIB, the temperature is slowly raised until it reaches the temperature 34°C at 13.00 WIB, the fish were left in hot stress for 4 hours, after 17.00 WIB the temperature is normalized again. Calculation of the number of individuals, calculation of dead fish, and checking of water quality is done every ten days. Substitution of aquarium water as much as 1/3 is done every two days and 2/3 every week.

2.2 Superoxide Dismutase Analysis

The examination of antioxidant levels was carried out at the end of the study by taking samples of the liver of tilapia fish. Tilapia liver organ is taken by composite namely by mixing three livers samples from each test but still in one treatment. Intake of the liver by dissecting the abdomen. The liver is put into a coolbox and immediately taken to the laboratory to be stored in a cooler at $\pm 4^{\circ}\text{C}$, then testing the levels of antioxidants. Testing levels of antioxidants is done by the method Marklund and Marklund [4974 \[11\]](#) namely measuring SOD activity by utilizing the auto-oxidation process of pyrogallol. This SOD enzyme will be a reference for damage caused by oxidative stress. The value obtained will show the effect of antioxidants in reducing oxidative stress that occurs.

Tilapia liver is taken as much as 1,15 gram and mixed with 13 mL extraction solution containing 0,15 M KCL 0,79 M EDTA (pH 7,4). The solution that has been made is homogenized using a manual homogenizer. Then homogenate is centrifuged at 9000 rpm using Beckman's centrifuge J2-21 with rotors JA 10 at temperature 4°C during 10 minutes. The supernatant is then used to test the activity of the enzyme superoxide dismutase.

Analysis of SOD levels was carried out with the help of pyrogallol solution as free radicals. Pyrogallol was taken 75 μL 10mM mixed with 2850 μL buffer fosfat 50 mM (pH 8,2) then added 75 μL sample. The degree of auto-oxidation of pyrogallol when added to the sample is compared with the standard (with 75 μL extraction solution) by measuring the increase in absorbance value at λ 340 nm during 2 minutes. One unit of enzyme SOD expressed as the number of enzymes needed to inhibit pyrogallol autoxidation reaction by 50% [[132](#)].

2.3 Observation Parameters

Analysis of superoxide dismutase levels in tilapia liver using the method [[132](#)].

$$\text{SOD Activity (\%)} = [A-B/A] \times 100\%$$

With :

- A = Reduction of absorbance of 180 minutes with absorbance of 0 minutes of control solution (%) and
- B = Reduction of absorbance of 180 minutes with absorbance of 0 minutes of sample solution (%).

The survival rate of the test fish is carried out using a formula [143] :

$$SR = \frac{N_t}{N_0} \times 100\%$$

With:

SR = Survival Rate

N_t = The number of fish at the end of the research

N₀ = The number of fish at the beginning of the research

Efficiency of feed in test fish is calculated by formula [154] :

$$EP = \frac{W_t + D - W_0}{F} \times 100\%$$

With:

EP = Feed efficiency (%)

W_t = Fish weight at the end of the research (gram)

D = Weight of dead fish during research (gram)

W₀ = Fish weight at the beginning of the research (gram)

F = Amount of feed consumed during research (gram)

Data on survival and efficiency of tilapia feed were analyzed using diversity test with an F test at 5%, and if there are real differences then carried out further tests (Which one??). Test for superoxide dismutase content and water quality were analyzed descriptively.

3. RESULTS AND DISCUSSION

3.1 Superoxide Dismutase Levels

This study analyzes the antioxidant activity of SOD found in tilapia liver which has been given treatment in the form of feeding containing antioxidants and heat stress as a source of oxidative stress. The value of antioxidant activity is indicated by the percentage of inhibition carried out by the SOD enzyme against free radicals formed during oxidative stress, % of the resistance obtained (Fig 1).

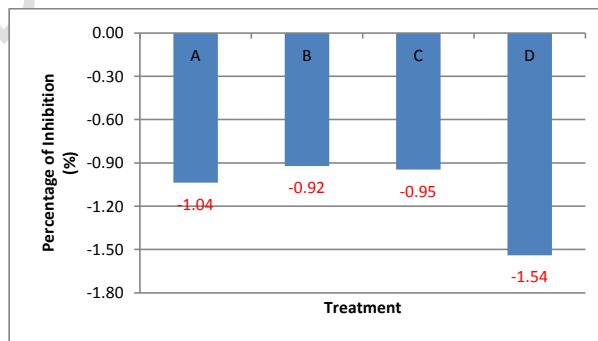


Fig. 1. Percentage graph of dismutase superoxide enzyme inhibition to free radicals

Comment [ct2]: Presented treatment in different cor as in survival rates

Antioxidant activity obtained is negative, this shows no antioxidant activity in the liver of tilapia rather it shows antagonistic activity against pyrogallol or no inhibition process occurs at the end of aquaculture [165]. Oxidative stress due to ambient temperature can cause an increase in free radicals which results in the erosion of antioxidants in the body [176].

Negative value of treatment B (5% TLF) and C (10% TLF) obtained still shows a better status than the control feed, These results are in accordance with the proposed hypothesis. But the increasing concentration of giving TLF show decreased levels of antioxidants where there is the influence of tannins in telang plants which give a bitter taste to the feed [187]. Same is the case with research results [198] that the provision of jaloh leaf meal which has antioxidant levels in tilapia with hot stress will result in decreased antioxidant activity as an increase in the concentration of jaloh leaf meal. This is also reinforced by opinions [2049] that the tannin content has a bitter taste so that the fish don't like it and causes food inhibition in test animals..

Decline in the value of SOD levels in addition caused by the use of TLF is less effective due to tannin content also caused by giving heat stress as a source of oxidative stress, this is in accordance with research conducted by [219] with the result that the SOD enzyme is the most active enzyme in responding to environmental stress conditions. Where the enzyme superoxide dismutase decreases at high temperatures between 30°C - 35°C, however SOD will increase along with a temperature drop of up to 10°C - 15°C.

The results of the treatment were fed with leaves of telang leaf meal shows the same results as the control due to heat stress given since the beginning of aquaculture which results in the erosion of existing antioxidants and those derived from TLF in each feeding. In this case it is advisable not to give heat treatment since the beginning of the fish culture, this is to let the body of a test fish increase antioxidant levels in the body first, before the arrival of free radicals from outside which will erode antioxidant levels.

3.2 Survival Rate

Based on observations for 40 days, the percentage of survival of tilapia (Fig 2). The survival rate of tilapia that is kept has a value above 60% and shows a positive response to all feeds given, in accordance with [224] that the minimum survival rate of tilapia for nursery is 60%. This shows a high survival value obtained by feeding fish with a duration of one day for each treatment.

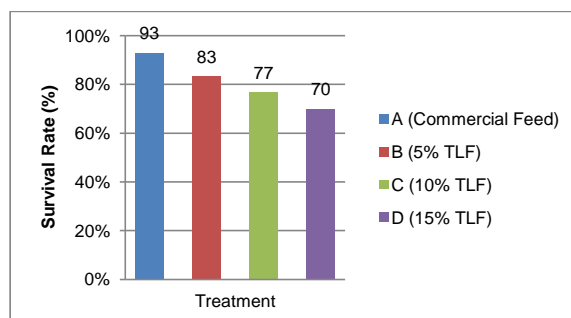


Fig. 2. Graph of final survival rate of tilapia aquaculture

The commercial feed used contains 40% protein, the protein content is used to maintain life especially more absorbed when the ambient temperature is not optimal, metabolism and afterwards used for growth.

Table 1. Average survival of tilapia during maintenance

Treatment	Survival Rate (%)
A (Commercial Feed)	93±0.06 ^b
B (5% TLF)	83±0.06 ^{ab}
C (10% TLF)	77±0.06 ^a
D (15% TLF)	70±0.01 ^a

With : Values followed by the same lowercase indicate no significant difference based on Duncan's Multiple Range Test at a 95% confidence level

Comment [ct3]: This table duplicated the figure 1, choosed the table

The results of analysis of variability in the survival of tilapia have a significantly different effect on the survival rate of tilapia because the calculated F value is greater than the F table (Table 1). The survival rate of tilapia is significantly different from the best treatment, treatment A and B, where this shows that the additional TLF feed gives a negative response to the survival of tilapia during aquaculture.

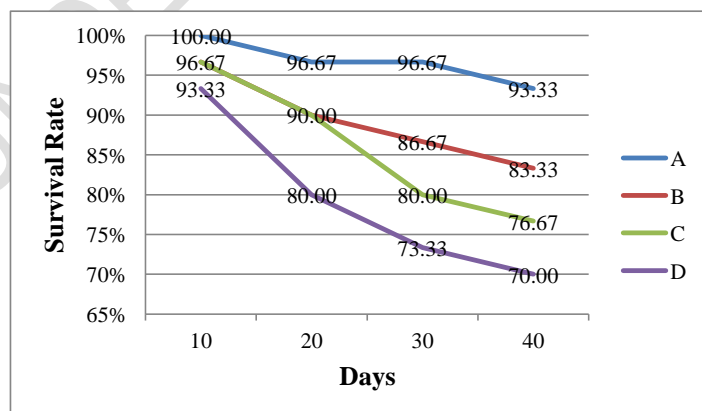


Fig. 3. Graph of survival rate of tilapia

The survival of tilapia has decreased (Fig. 3) with the test fish in treatment A has the highest survival rate compared to other treatments. High survival in treatment A (control) due to tilapia fish use very well to maintain life in extreme environmental conditions, and tilapia that are kept also do not need to adapt to the type of feed provided, because the food used during cultivation is the same as the food used when the adaptation first came and when fish are in fish farmers.

In treatments B and C, the survival of tilapia decreases, because the feed given has been mixed with telang plants so that it affects the response of fish to feed. This can be seen in preliminary research conducted to determine the response of fish to feed that has a different TLF mixture. The result is that with increasing TLF mixture, the fish's response to feed is slower.

Treatment D is the treatment with the lowest survival. This is consistent with the preliminary research described above where treatment D feed had the highest TLF dose, so the fish has a low response. As a result, in addition to adapting to the environment, Test fish must also adapt to new types of feed so that the stress that occurs is higher.

From some of the explanations above, it is found that the effect of stress comes from the high ambient temperature, so the test fish must utilize every feed given to survive. As well as the increasing concentration of TLF in feed, the response of fish is lower because it has to go through the adaptation of new feed that is done since the beginning of aquaculture. This explains why TLF administration of up to 15% is not significantly different. This is supported by research [232] about the effect of high temperatures on the survival of tilapia the result is that temperatures higher than 32 °C will reduce survival to some extent. This is what explains that the higher the TLF concentration content, the smaller the survival rate of fish.

Less optimal use of similar feed was also obtained from research results [243] about the survival of tilapia which is fed with supplementation with jaloh leaf meal, where the survival of the highest test fish is in the treatment B 5%. The increasing concentration of jaloh leaf meal is the lower the survival rate. The results of studies with TLF are different from the results of jaloh leaf meal although both have high tannin content. In studies with TLF. Ambient temperatures have been made high from beginning, will affect the ability to adapt to new feed, resulting in lower survival.

Decreased survival as the temperature increases is found in the research [254] where at a temperature of 30 °C and 32 °C uniform seed size, but the longer the cultivation time the size of tilapia will be different and affect the high mortality rate through the formation of a food hierarchy, and the survival rate at these temperatures is around 79% to 85%.

3.3 Feed Efficiency

Based on observations for 40 days, the value of the efficiency of tilapia feed obtained was found at Fig. 4.

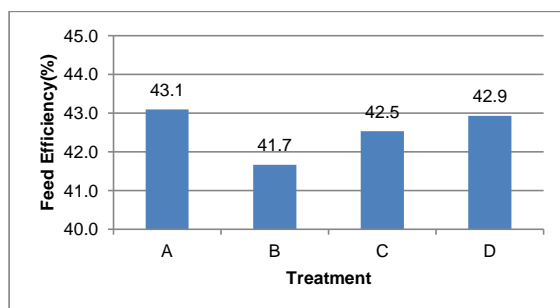


Fig. 4. Feed efficiency graph for each treatment

The low level of feeding efficiency indicates that the feed given is not fully utilized by tilapia for its growth, but is used to defend the body from high ambient temperatures, where temperatures affect the digestibility of fish [243].

Table 2. Average feeding efficiency during aquaculture

Treatment	Feed Efficiency (%)
A (Commercial Feed)	43,1±1,40 ^a
B (5% TLF)	41,7±1,77 ^a
C (10% TLF)	42,5±1,12 ^a
D (15% TLF)	42,9±1,62 ^a

With : Values followed by the same lowercase indicate no significant difference based on Duncan's Multiple Range Test at a 95% confidence level

Comment [ct4]: Translate thes data in english

Comment [ct5]: This table duplicated the figure 1, choosed the table

Based on the results of analysis of variance (Table 2) the value of the obtained feed efficiency was not significantly different between treatments. This shows that feed with the addition of telang leaf meal does not provide a positive response to the efficiency of feeding. Efficiency of feed is closely related to digestibility which illustrates the percentage of nutrients that enter the fish's body and absorbed by the digestive tract of fish. Feed efficiency is influenced by stress caused by the environment which is in accordance with the results of research [265] which shows stress in fish farming will disrupt eating behavior, especially reducing food intake and negative effects on feed conversion efficiency.

Efficiency of tilapia feed at high temperature maintenance it was found that water temperature 32°C significantly reduced feed efficiency, this is due to increasing temperatures and makes the process of eating fish slow [232]. Higher temperatures will accelerate the rate of food that is digested through the intestinal tract, thereby reducing the digestion and assimilation of nutrients.

3.4 Water Quality

Water quality is a very important factor in aquaculture, the water used must meet the optimal needs of the fish. The measured water quality parameter is temperature, *Dissolved Oxygen* (DO), and pH. Water quality measurements carried out 5 times with a span of 10 days for 40

days of maintenance. The results of measurement of water quality parameters can be seen in Table 3.

Table 3. Water quality parameters during aquaculture

Treatment	Max temperature (°C)	Min temperature (°C)	DO (mg/L)	pH
A (Commercial Feed)	32 – 33	26 – 27	4,3 – 7,4	7,03 – 8,17
B (5% TLF)	33	26 – 27	4,0 – 7,3	7,01 – 8,20
C (10% TLF)	32 – 33	26	3,8 – 7,1	6,87 – 8,25
D (15% TLF)	33	26 - 27	3,9 – 7,1	7,01 – 8,25
SNI Standard	30	25	Min 5	6,5 – 8,5

Comment [ct6]: Translate thes data in english

Temperature is a very important factor in fish farming that determines the success of tilapia culture, caused by fish are cold-blooded animals [276]. Temperature during aquaculture affects fish growth and survival. During the period of fish aquaculture is at a temperature 26°C - 33°C, which range according to SNI 2009 that is 25°C - 30°C. Increased water temperature up to 33°C carried out by heat stress as a source of oxidative stress when aquaculture, this condition results in low feed efficiency which indicates that the nutrients absorbed by the body are used to adjust to the ambient temperature. In heat stress treatment, the temperature is increased to the maximum limit, during the increase in temperature to the optimal limit that is 30°C, the fish will take food that makes the process of digestion and metabolism also increases.

Dissolved oxygen (DO) is a water quality parameter that shows the solubility of oxygen in a body of water, where dissolved oxygen is influenced by temperature [287]. Dissolved oxygen levels during aquaculture range between 3.9 – 7.4 3.9 – 7.4 mg/L which is in a bad condition which is < 5 mg/L. This is caused by an increase in temperature to exceed the optimal temperature of aquaculture is 30°C, which according to [298] states that oxygen levels will increase the lower the temperature and oxygen levels will decrease the higher the temperature.

The results of measurement of the pH range during tilapia aquaculture 6.87 6.87 – 8.25 8.25, This value is in good condition according to SNI, namely between 6.5 6.5 – 8.5 8.5. The pH value that can interfere with fish life is a pH that is too low (very sour) and pH that is too high (very basa), for the most part fish can adapt to the aquatic environment with a pH around 5 – 9 [3029].

4. CONCLUSION

You have 4 results? Your conclusion should be the answer for your specific objectives; then four conclusions

Based on the results of the research that has been carried out, it can be concluded that the addition of up to 15% of telang leaf meal to commercial feed by giving heat stress since the beginning of the aquaculture period does not have the effect of increasing antioxidant levels.

REFERENCES [ORDER THESE REFERENCES](#)

- [1]. Salsabila M, Suprpto H. Tilapia Enlargement Techniques (*Oreochromis niloticus*) at Pandaan Freshwater Aquaculture Installation Jawa Timur. Journal of Aquaculture and Fish Health. 2018; 7(3).
- [2]. Kementerian Kelautan dan Perikanan. Statistics book 2012 Maritime Affairs and Fisheries: Jakarta; 2013.
- [3]. Kementerian Kelautan dan Perikanan. ne Data on Marine Production and Fisheries in 2017. Jakarta; 2018.
- [4]. Angreni N.P.W, Arthana IW, Wulandari E. Distribution of Pathogenic Bacteria in Tilapia (*Oreochromis niloticus*) on Lake Batur Bali. Current Trends in Aquatic Scscience. 2018; 96-103.
- [5]. Payung CN, Manoppo H. Increased Non-Specific Immune Response and Tilapia Growth (*Oreochromis niloticus*) Through Giving Ginger (*Zingiber officinale*). Journal of Aquaculture. 2015; 3(1): 11-18
- [6]. Rana KJ, Siriwardena S, Hasan MR. Impact of Rising Feed Ingredients Prices on Aquafeeds and Aquaculture Production. FAO Fisheries and Aquaculture Technical Paper. 2009; 63p
- [7]. Magdalena S, Natadiputri GH, Nailufar F and Purwadaria T. Utilization of Natural Materials as Functional Feed. Wartazoa. 2013; 23(1).
- [8]. Panagan AT. "Effect of Addition of Carrot Flour (*Daucus carota* L.) Against Peroxide Numbers and Free Fatty Acids in Bulk Cooking Oil". Journal of Science Research; 2011
- [9]. Vankar, P. S, and J. Srivastava. Evaluation of Anthocyanin Content in Red and Blue Flowers. International Journal of Food Engineering. 2010; 6 (4).
- [10]. Lakshmi CHNDM, Raju BDP, Mahdavi T and Sushma NJ. Identification of Bioactive Compounds by FTIR Analysis and In Vitro Antioxidant Activitiy of *Clitoria ternatea* Leaf and Flower Extracts. Indo American Journal of Pharmaceutical Research. 2014; 4 (9).
- [11]. Campanella L, Bonanni A, Finotti E, Tomassetti. M. Biosensors for determination of total and natural antioxidant capacity of red and white wines: comparison with other spectrophotometric and fluorimetric methods. Biosensors and Bielelectronics. 2004'19: 641–651.
- [12]. Marklund and Marklund G. Involvement of the superoxide anion radical in the autooxidation of pyrogallol and a convenient assay for superoxide dismutase. Eur J Biochem. 1974; 47: 469-474
- [13]. Effendie MI. Biological Methods of Fisheries. Faculty of Fisheries. Institut Pertanian Bogor: 1997.
- [14]. Djajasewaka HY. Fish feed. Penebar Swadaya. Jakarta: 1985.
- [15]. Mir MI, Khan S, Bhat SA, Reshi AA, Shah FA, Balki MH and Manzoor R. Scenario of Genotoxicity in Fishes and Its Impact on Fish Industry. Journal of Environmental Science, Toxicology and Food Technology. 2014; 8 (6): 65-76.
- [16]. Sreejai R and Jaya DS. Studies on the changes in lipid peroxidation and antioxidants in fishes exposed to hydrogen sulfide. Toxicol Int. 2010; 17:7177.
- [17]. Kumar RT, Kumar S, and Anju VS. Phytochemical and Antibacterial Activities of Crude Leaf and Extracts of *Clitoria ternatea* Varieties (*Fabaceae*). Journal of Pharmacognosy and Phytochemistry. 2017; 6 (6): 1104-1108.
- [18]. Sutari VT, Sugito, Aliza D, and Asmarida. Malondialdehyde levels (MDA) in the Tilapia Liver (*Oreochromis niloticus*) who were given Hot Stress and Jaloh Flour Supplementation Feed. Jurnal Medika Veterinaria. 2013; 7 (1).
- [19]. Yunita EA, Nanik HS. and Jafron WH. "Effect of Teklan Leaf Extract (*Eupatorium riparium*) Against mortality and growth of larvae *Aedes aegypti*". BIOMA. 2009; 11 (1)

- [20]. Zhang J, Jiang F, Yang P, Li J, Yan G and Hu L. Responses of canola (*Brassica napus* L.) cultivars under contrasting temperature regimes during early seedling growth stage as revealed by multiple physiological criteria. *Acta Physiol. Plant.* 2015;37 (7).
- [21]. Mulyani YS, Yulisman and Fitriani M. Growth and Efficiency of Tilapia Feed (*Oreochromis niloticus*) Periodically Fasted. *Jurnal Akuakultur Rawa Indonesia.* 2014; 2(1):1-12
- [22]. Pandit NP and Nakamura M. Effect of High Temperature on Survival, Growth and Feed Conversion Ratio of Nile Tilapia (*Oreochromis niloticus*). *Out Nature.* 2010; 8: 219-224.
- [23]. Yanti Z, Muchlisin ZA and Sugito. Tilapia Fish Growth and Survival (*Oreochromis niloticus*) in Several Jaloh Flour Concentrations (*Salix tetrasperma*) in the Feed. *Journal of Fisheries and Aquatic Sciences.* 2013; 2(1): 16-19.
- [24]. Drummond CD, Murgas LDS and Vicentini B. Growth and Survival of Tilapia (*Oreochromis niloticus*) Submitted to Different Temperatures During the Process of Sex Reversal. *Cienc. agrotec.* 2009; 33(3): 895-902
- [25]. Leal E, Fernandez B, Guillot R, Riot D, Miguel J and Reverter C. Stress Induced Effects on Feeding Behavior and Growth Performance of The Sea Bass (*Dicentrarchus labrax*): A Self Feeding Approach. *J Comp Physiol B.* 2011; 181 (8): 1035-44
- [26]. Aliza D, Winaruddin and Sipahutar LW. Effects of Increased Water Temperature on Behavior, Anatomical Pathology, and Tilapia Gills Pathology (*Oreochromis niloticus*). *Jurnal Medika Veterinaria.* 2013; 853-1943.
- [27]. Nugroho, A. Water Quality Bioindicator. Penerbit Universitas Trisakti, Jakarta: 2006
- [28]. Odum, E.P. Fundamental of Ecology. W.B. Saunder Com. Philadelphia : 1971.
- [29]. Effendi, H. Water Quality Study. Kanisius. Yogyakarta: 2003.