Original Research Article

MATHEMATICAL MODELLING OF PREVENTION, CONTROLLING AND TREATMENT OF INFECTIOUS DISEASES DETERMINANT FACTOR OF INFANT MORTALITY WITH MEDICINAL PLANTS GROWN IN NIGERIA, WEST AFRICA.

ABSTRACT

Infant mortality is a challenge for third-world countries like Nigeria where there is little next to non-availability of conventional drugs, and if available, it is costly and out of the reach of the common populace. It is a fact that medicinal plant is a gift from mother nature, but its uses and efficacy have been overlooked over the century because of the conventional drugs but its efficacy is still intact. The purpose of this paper is to formulate the prevention, control, and treatment of various infectious diseases determinant factor of infant mortality among growing infants with medicinal plants being grown in Nigeria, West Africa using the mathematical modeling approach as the research/review scientific point of view. The mathematical model is a system of first-order non-linear ordinary differential equations which are partitioned into five different compartments. Two equilibria states exist, the disease-free equilibrium and endemic equilibrium which are locally asymptotically stable if the basic reproductive number is less than one and unstable if the basic reproductive number is greater than one. Numerical simulations were performed using hypothetical values for the parameters used in the model. The model shows that an increase in the medicinal plants grown or found in the country leads to low disease prevalence among the susceptible infant population considered in this work. Therefore, our medicinal plants become a very alternative for the prevention, control, and treatment of infectious diseases to reduce or prevent infant mortality among infants, especially in rural areas. Also, this will elevate the knowledge from African trade-medical practice and rejuvenate our ethnobotanical properties and characteristics for future uses.

Keywords: Mathematical Modelling, Basic Reproductive Number, Numerical Simulation, Infectious Diseases, Infants, Nigeria.

INTRODUCTION

Medicinal plant has been used for ages in the treatment of disease and it was not known where or when plants first began to be used, but the connection between plants and health has existed for thousands of years (Faleyimu and Oluwalana, 2008). There is limited documentation of medicinal plants used in the treatment of infants' diseases in Nigeria, but several ethnobotanical studies focusing on medicinal plants have been documented all over the world (Cox, 2005; Kumar *et al.*, 2005; Singh and Singh, 2001; Wang *et al.*, 2005).

Medicinal plants possess various arrays chemical substances that support certain physiological and biochemical activities in the human body and they are known as phytochemicals or secondary metabolite. These chemicals are non-nutritive substances used to heal various infectious diseases, as well as provide disease preventive properties (Belman,1983),Botterwecket al,.2000). Pharmacological activity of medicinal plants resides in their secondary metabolites, which are relatively smaller in quantity in contrast to the primary molecules such as carbohydrates, proteins, and lipids. These metabolites has a clue to manufacture new structural types of antimicrobial and antifungal chemicals that are comparatively safe to humans (Ngonoet al, 2003).

Secondary metabolites have different classes with greater antimicrobial properties they are flavonoids (flavones, flavonols, flavanols, isoflavones, anthocyanidins), phenolic acids (hydroxybenzoic, hydroxycinnamic acids), stilbenes, lignans, quinones, tannins, coumarins (simple coumarins, furanocoumarins, pyranocoumarins), terpenoids (sesquiterpene lactones, diterpenes, triterpenes, polyterpenes), alkaloids, glycosides, saponins, lectins, steroids, and polypeptides (Antony,2013)

These compounds have copious mechanisms that underlie antimicrobial activity, e.g., disturbing microbial membranes, weakening cellular metabolism, control biofilm formation, inhibiting bacterial capsule production, attenuating bacterial virulence by controlling quorum-sensing, and

reducing microbial toxin production (Alamet al, 2002). Medicinal plants produce a boundless quantity of secondary metabolites that have great antimicrobial activity (Belman, 1983). These plant-produced low molecular weight antibiotics are classified according to two types, namely phytoanticipins, which are involved in microbial inhibitory actions, and phytoalexins, which are generally anti-oxidative and synthesized de novo by plants in response to microbial infection (Botterwecket al, 2000).

A World Health Organisation (WHO) Expert Group defined Traditional Medicine as the sum total of all knowledge and practices, whether explicable or not, used in diagnosis, prevention and elimination of physical, mental, or social imbalance and relying exclusively on practical experience and observation handed down from generation to generation, whether verbally or in writing In view of the fact that infant diseases are widely observed in Nigeria due to the attitude of mothers to some illness which are supposed to be treated, non-availability of health care practitioners and cost of accessing orthodox mode of treatment, it is paramount to document some ethnobotanicals used in the treatment of infant diseases in Western Nigeria. The following infant diseases were treated with medicinal plant, they are Blood Shortage (Anaemia), Infant Constipation, Infant Convulsion, Infant Cough, Infant Diarrhoea, Infant Dysentery, Jaundice, Helminthic infestation, Malaria, Measles, Small Pox/Chicken Pox, Teething, The medicinal plant used for this treatment include Sorghum bicolor shoots, Mangifera indica and Theobroma cacao, Annanascomosus, sweet orange, Allium cepa, Allium sativum, Zingiber officinale, Xylopia aethiopica, Alstonia boonei, Perquatina nigrescence, Matured unripe pawpaw, Azadirachta indica, Morinda lucida, Momordica charantia, Rauwolfia vomitoria, indigofera, Cajanus cajan and stem of sugar cane. Emphasis on the use of medicinal plants had hitherto been placed on the treatment rather than prevention of infectious diseases. However, there exists in the literature considerable report in recent times on research work on the use of medicinal plants and their constituents in disease prevention.

Infant mortality is the unconditional death of little children from ages 1(one) to 5(Five). Some school of thought describes Infant mortality as the death of an infant before his or her first birthday. The infant mortality rate is the number of infant deaths for every 1,000 live births. In 2013, the leading cause of infant mortality include birth defects (Mathews *et al*, 2002). Other

leading causes of infant mortality include birth asphyxia, pneumonia, congenital malformations, term birth complications such as abnormal presentation of the fetus umbilical cord prolapse, or prolonged labor, (Basics, 2017) neonatal infection, diarrhea, malaria, measles and malnutrition (IM & NH, 2017).

One of the most common preventable causes of infant mortality is smoking during pregnancy (Hall *et al*,.2016)). Lack of prenatal care, alcohol consumption during pregnancy, and drug use also cause complications which may result in infant mortality (Genowska*et al*,.2015). Many environmental factors and seasonal variation may contribute to infant mortality, such as the mother's level of education, environmental conditions, political and medical infrastructure. (Genowska *et al*, 2015). Improving sanitation, access to clean drinking water, immunization against infectious diseases and the use of medicinal plants. A medicinal plant is any plant which, in one or more of its organs, contains substances that can be used for therapeutic purposes or which are precursors for the synthesis of useful drugs. This description makes it possible to distinguish between medicinal plants whose therapeutic properties and constituents have been established scientifically, and plants that are regarded as medicinal but which have not yet been subjected to a thorough scientific study

However, babies born in low to middle income countries in sub-Saharan Africa like Nigeria are at the highest risk of neonatal death. Bacterial infections of the bloodstream, lungs, and the brain's covering (meningitis) may be responsible for 25% of neonatal deaths. Newborns can acquire infections during birth from bacteria that are present in their mother's reproductive tract. The mother may not be aware of the infection. These bacteria can move up the vaginal canal into the amniotic sac surrounding the baby. Maternal blood-borne infection is another route of bacterial infection from mother to baby. Neonatal infection is also more likely with the premature rupture of the membranes (PROM) of the amniotic sac. (Chan *et al*, 2013).

Seven out of ten childhood deaths are due to infectious diseases: acute respiratory infection, diarrhea, measles, and malaria. Acute respiratory infection such as pneumonia, bronchitis and bronchiolitis account for 30% of childhood deaths; 95% of pneumonia cases occur in the developing world. Diarrhea is the second-largest cause of childhood mortality in the world, while malaria causes 11% of childhood deaths. Measles is the fifth-largest cause of childhood mortality (Nussbaum, 2011).

Infant mortality rate (IMR) is the number of deaths per 1,000 live births of children under one year of age. The rate for a given region is the number of children dying under one year of age, divided by the number of live births during the year, multiplied by 1,000. (Andrews *et al.*, 2008)

The aim of this paper is to formulate and analyse a mathematical model that extends and complements the ones in the literatures by incorporating medicinal plant class denoted by M(t). Mathematical models are widely used to examine, explain and predict the dynamics of infectious disease transmission and models of specific diseases of global importance have played important role in developing public health strategies for control and prevention of infectious diseases (Anderson & May, 1991).

1. FORMULATION OF THE MATHEMATICAL MODEL

We formulate a non-linear deterministic model for the transmission dynamics of the infectious diseases among the infants. The model subdivides the infant population into five different compartments depending on the epidemiological status of the infants. The compartments are Susceptible class S, Exposed class E, Infected class I, Medicinal plant used class M and the Recovered class R. To indicate this mathematically, we have the following systems of ordinary differential equations as:

$$S'(t) = \Lambda + (1 - P)A + \delta R(t) - \beta S(t)I(t) - (\mu + \eta)S(t); \tag{1}$$

$$\mathbf{E}'(\mathbf{t}) = \beta \mathbf{S}(\mathbf{t})\mathbf{I}(\mathbf{t}) + \eta \mathbf{S}(\mathbf{t}) - (\mu + \sigma)\mathbf{E}(\mathbf{t}); \tag{2}$$

$$I'(t) = \sigma E(t) - (\mu + \tau)I(t); \tag{3}$$

$$\mathbf{M}'(\mathbf{t}) = \mathbf{P}\mathbf{A} + \tau \mathbf{I}(\mathbf{t}) - (\mu + \omega)\mathbf{M}(\mathbf{t}); \tag{4}$$

$$\mathbf{R}'(\mathbf{t}) = \omega \mathbf{M}(\mathbf{t}) - (\mu + \delta)\mathbf{R}(\mathbf{t}). \tag{5}$$

The following assumptions were considered to formulate the above mathematical model.

- 1. All susceptible infants can be exposed and infected through a direct contact with an infected infant in the community.
- 2. Infants are only born into the susceptible class.
- 3. Birth rate is not equal to the death rate.
- 4. Susceptible infants get infected with the infectious diseases at a rate proportional to the susceptible population.
- 5. All parameters used in the mathematical model are non-negative.
- 6. Some infectious diseases are re-infected in the community among the infants.

The Table 1 below shows the definitions and the hypothetical values of the parameters used in the formulation of the mathematical model.

Table 1: Parameter hypothetical values for the model

Parameters	Definition	Hypothetical	Source
		values	
٨	rate of newborn infants into the susceptible infant	100	Estimated
	class		
P	fraction of infants who are given herbal medicine	0.305	Estimated
	plants		
A	number of infants with infectious diseases	1500	Binuyo (2014)
μ	mortality or death rate of the infants in the	0.2	Assumed
	community		
δ	rate at which re-infection of the infectious diseases	0< δ < 1	Binuyo (2012)
	occur		
β	rate of transmission of the infectious diseases in the	0.5	Assumed
	community		
η	rate at which the susceptible infants are exposed to	0.4	Assumed
	infectious diseases		
σ	rate at which the exposed infants become infected	0.1	Binuyo (2012)
	with infectious diseases		
τ	rate at which infected infants are treated, controlled	0.35	Binuyo (2014)
	and prevented from infectious diseases		
ω	rate at which the efficacy of the herbs reduces leading	0.0182	Assumed
	to re-infection of the infectious diseases in the		
	community.		
S_0	initial value of Susceptible infant class with time	1.6568	Estimated
E ₀	initial value of exposed infant class with time	4.4367	Estimated
I_0	initial value of infected infant class with time	0.8066	Estimated

M_0	initial value of medicinal plant used infant class with	5.8356	Estimated
	time		
R_0	initial value of recovered infant class with time	4.0856	Estimated

2. EQUILIBRIUM STATES OF THE MATHEMATICAL MODEL:

The mathematical model (1) – (5) exhibits two states of equilibrium i.e. disease free equilibrium and the endemic equilibrium such that $\frac{dS}{dt} = \frac{dE}{dt} = \frac{dI}{dt} = \frac{dM}{dt} = \frac{dR}{dt} = 0$.

Solving equations (6) - (10) below,

$$\Lambda + (1 - P)A + \delta R(t) - \beta S(t)I(t) - (\mu + \eta)S(t) = 0;$$
 (6)

$$\beta S(t)I(t) + \eta S(t) - (\mu + \sigma)E(t) = 0; \tag{7}$$

$$\sigma \mathbf{E}(\mathbf{t}) - (\mu + \tau)\mathbf{I}(\mathbf{t}) = 0; \tag{8}$$

$$PA + \tau I(t) - (\mu + \omega)M(t) = 0; \tag{9}$$

$$\omega \mathbf{M}(\mathbf{t}) - (\mu + \delta)\mathbf{R}(\mathbf{t}) = \mathbf{0}. \tag{10}$$

We obtain the following results;

(i) Disease free equilibrium (DFE) i.e. in the absence of any infection (I = 0), then the disease free equilibrium points are:

$$(S^*, E^*, I^*, M^*, R^*) = (\frac{\Lambda + (1-P)A}{\mu + n}, 0, 0, 0, 0, 0)$$
 (11)

(ii) The Endemic Equilibrium State (EE) i.e. in the presence of the infection (I \neq 0), we obtain;

$$S^* = \frac{(\mu + \sigma)(\mu + \tau)I^*}{\sigma(\beta I^* + \eta)}, E^* = \frac{(\mu + \tau)I^*}{\sigma}, M^* = \frac{PA + \tau I^*}{(\mu + \omega)}, R^* = \frac{\omega(PA + \tau I^*)}{(\mu + \delta)(\mu + \omega)}$$
(12)

For I* is the positive root of the quadratic equation $AI^{*2} + BI^* + C = 0$ where

$$A = \beta \omega \tau (\mu + \delta)(\mu + \omega)(\mu + \sigma)(\mu + \tau) - \beta \sigma \delta \omega \tau \sigma \omega \tau \tag{13}$$

$$B = \omega \tau (\mu + \delta)(\mu + \omega)(\mu + \eta)(\mu + \sigma)(\mu + \tau) - PA\tau\omega\delta\omega\sigma\beta - \sigma\tau\omega\delta\omega\tau\eta - (\Lambda + A - \omega)(\mu + \delta)(\mu + \omega)(\mu + \delta)(\mu + \delta$$

$$PA)\omega\tau\sigma\beta(\mu+\delta)(\mu+\omega)$$
 (14)

$$C = \eta \omega \tau \sigma (\mu + \delta)(\mu + \omega)(\Lambda + A - PA) + PA \sigma \tau \omega \eta \delta \omega \tag{15}$$

With these values for S*, E*, I*, M* and R*, the positivity and uniqueness of the endemic equilibrium are guaranteed if and only if $R_0 > 1$ where R_0 is the basic reproductive number (Diekmann O. et al, 2000) given in the form;

$$R_0 = \frac{\sigma\beta}{(\mu + \sigma)(\mu + \tau)} \tag{16}$$

In the endemic disease state, the number of infected infants is strictly positive and constant. So, if some of the solutions of the system of equation I(t) approach as time goes to infinity, the number of infective will remain strictly positive for a long time and approximately equal to I(t). Thus, the disease remains in the population and becomes endemic except adequate measures are done to control or prevent the rapid spread of the disease among the infant population. Therefore, in the event of an epidemic, the theoretical determination of condition that can make R_0 less than unity is of great public health interest such that the disease can be greatly reduced or eventually eradicated among the infant population (Hethcote, 2000).

3. NUMERICAL SIMULATIONS OF THE MODEL

Numerical simulations were carried out to graphically explain the long term effects of preventing, controlling and treating on the infectious diseases determinant factor among the infants using the medicinal plants grown in Nigeria. In order to support the analytical results, graphical representations showing the time graphs of different state variables are provided.

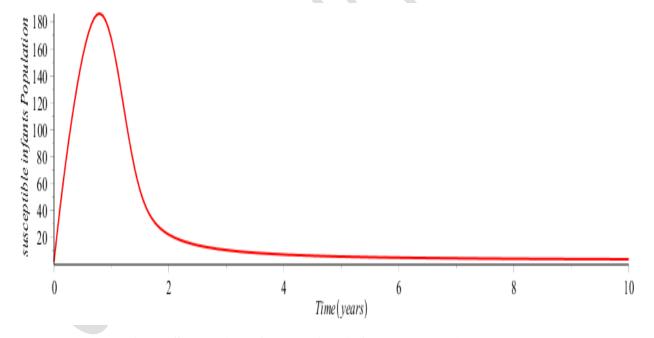


Fig. 1: Simulation of susceptible infant's population

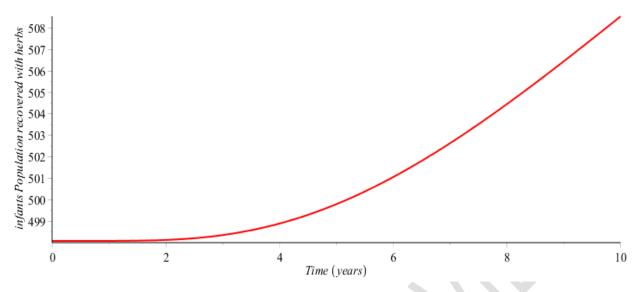


Fig. 2: Simulation of Recovered Infants Population from infectious diseases using Herbs

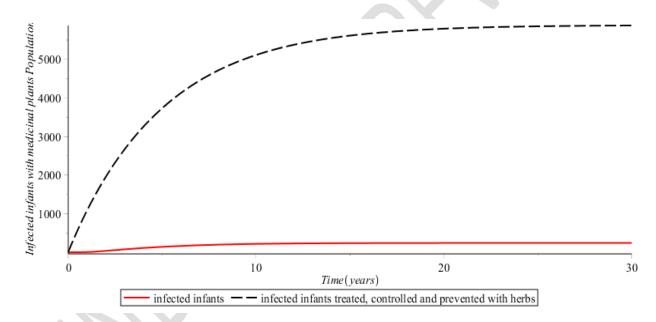


Fig.3: Simulation of infected infants treated, controlled and prevented with medicinal plants

RESULTS

Numerical simulations were carried out to graphically illustrate the long term effect of controlling the dynamics of infectious diseases determinant factor using the medicinal plants (herbs) among the infant mortality. In order to support the analytical results, graphical representations showing the time graphs of different state variables are provided.

Fig. 1 is the diagram showing the dynamics of the susceptible infant population. The susceptible population increases initially as the newborns are introduced into the community and later decreases as time increases. This decrease was possibly because of the high rate of introduction of the medicinal plants in order to control, prevent and treat the infected infants among the susceptible population of the infants.

Fig. 2 shows the graph of the infant population that are recovered from the infectious diseases determinant factor using the medicinal plants. It is observed that as the rate of infectious diseases determinant factor increases, the infant population of the recovered infants shows some rapid increase due to the treatment of the infants with the medicinal plants among the infected infants.

Fig. 3 shows the graph of the combination of the infected infants and the infants treated, prevented and controlled with the medicinal plants. Observe that, as the population of the infected infants increases, the population of the infants that are using medicinal plants are increasing with time.

Tables 2 & 3 show the various medicinal plants that are used to prevent, control and treat infectious diseases determinant factor among the infant which can reduce the infant mortality.

Table 2 ; Selected Profile of plants used in the treatment & prevention of infant Mortality/diseases

S/N	Botanical Names	Family	Common Name	Local Names	Plant Parts Used	Medicinal uses
1.	Sorghum bicolor	Poaceae	Guinea corn	Oka baba (Y)	Shoots, Leaves	Blood tonic, malaria, fever
2.	Mangiferaindica	Anacardiaceae	Mango tree	Mangoro (Y)	Leaves, Barks	Malaria, fever, Anaemia
3.	Theobroma cacao	Sterculiaceae	Cocoa	Koko (Y)	Leaves, Barks	Blood tonic, tooth ache
4.	Annanascomosus	Bromeliaceae	Pineapple	OpeOyinbo (Y)	Fruits	Malaria, Dysentery
5.	Xylopiaaethiopica	Annonaceae	Ethiopian pepper	Eeru (Y)	Fruits	Cough, convulsion, stomach ach
6.	Allium sativum& All	Liliaceae	Garlic	AlubosaAyuu (Y)	Bulbs	Convulsion
7.	Zingiberofficinale	Zingiberaceae	Ginger	Atale (Y)	Rhizomes	Fever, malaria, digestive disorde disease, typhoid.
8.	Alstoniaboonei	Apocynaceae	Stool wood	Ahun (Y)	Leaves, Barks	Fever, Convulsion, Diarrhoea
9.	Perquatinanigrescer	Periplocaceae	African parquatina	Ogbo (Y)	Leaves	Anti-anaemic, dysentery, stomac disorber, skin disease
10.	Azadirachtaindica	Meliaceae	Neem tree	Dongoyaro (H) Igika	Leaves, Stem	Dysentery, Fever
11.	Morindalucida	Rubiaceae	Brime stone tree	Oruwo (Y)	Leaves	Jaundice, fever, malaria
12.	Momordicacharanti	Cucurbitaceae	African cucumber	Ejinrin were (Y)	Leaves	Convulsion, Disorder, measles, o

13.	Rauwolfiavomitoria	Apocynaceae	Serpent wood	Asofeyeje (Y)	Leaves	Nervous disorder, jaundice, scab
	(Benth.) Swizzlestic					diarrhoea
14.	Theobroma cacao	Sterculiaceae	Cocoa	Koko (Y)	Leaves, Barks	Blood tonic, tooth ache
15.	Vernoniaamygdalind	Asteraceae	Bitter leaf	Ewuro (Y)	Leaves, Stem	Stomach disorder, dysentery, ski infection, malaria
16.	Aframomummelegue K. Schum	Zingeberaceae	Alligator pepper	Ataare (Y)	Fruits	Small Pox, Chicken Pox, Cough
17.	Cajanuscajan(L.) M	Mimosaceae	Pigeon Pea	Otili (Y)	Leaves, Seeds	Small pox, Measles
18.	Carica papaya L.	Caricaceae	Pawpaw	Ibepe (Y)	Leaves	Malaria, Jaundice, Convulsion
19.	Cymbopogoncitrata(Strapt.	Poaceae	Lemon grass	Kookooba (Y)	Leaves	Malaria, Cough
20.	Garcinia kola Hecke	Gutiferae	Bitter Kola	Orogbo (Y)	Nuts	Cough, Catarrh, Jaundice

Source; Fatoba et al,.2018

Table 3: Enumeration of the Medicinal plants recipes for the treatment of various infant diseases

s/n	Blood	Recipes	Modes of Administration and Dosage
	Shortage		
	(Anaemia)		
1)	Blood	Sorghum bicolor shoots, barks of Mangiferaindica and Theobroma	5 ml of the decoction taken orally thrice daily
	Shortage	cacao are boiled together for 30 minutes with 2 litres of water, two	
	(Anaemia)	tins of milk and 4 cubes of sugar are added to the herbal	
		preparation	
2)	Infant	The juice of Annanascomosusand sweet orange are extracted for	The Juice is taken orally; 0-12 months old taken
	Constipation	drinking.	5 ml thrice daily; 1-5years old taken 5 ml five
			times daily.
3)	Infant	Allium cepa, Allium sativumand Zingiberofficinaleare ground	The mixture is applied topically all over the
	Convulsion	together. The ground material is mixed with palm oil.	body of the baby and should be allowed to enter
			the eyes. About 21/2 ml is given orally to the
			affected child
4)	Infant	The fruits of <i>Xylopiaaethiopica</i> are added to fried oil. The fruits are	It is taken Orally by licking.
	Cough	then separated after 8 minutes and sugar is then added to the	
		extract.	
5)	Infant	The leaves of Alstoniabooneiis squeezed to obtain the juice.	The Juice is Orally by using 5 ml of the juice
	Diarrhoea		thrice daily
6)	Dysentery	Leaves of <i>Perquatinanigrescence</i> are squeezed. The juice extracted	5 ml of the juice taken orally every three hours

		is mixed with a pint of salt.		
7)	Jaundice	Matured unripe pawpaw is cut into pieces and soaked in fermented	2.5ml taken orally five times daily	
		maize water (Omiidun) for three days.		
8)	Helminthic	The leaves of Azadirachtaindicais squeezed with water. Lime and	10 ml of the infusion taken orally twice daily	
	infestation	garlic are added to the infusion.		
9)	Malaria	Extract of the leaves of <i>Morindalucida</i> is squeezed out using water	5 ml taken orally thrice daily	
10)	Measles	The leaves of <i>Momordicacharantia</i> is boiled in water	The decocted material is used to bathe every day	
			and night until the measles cure.	
11)	Small	The leaves of Rauwolfiavomitoria, indigofera and	It is taken orally, 15 ml four times daily; the	
	Pox/Chicken	Cajanuscajanare boiled in water for 45minutes.	decoction is also used for bathing	
	Pox			
12)	Teething	The juice is extracted from the stem of sugar cane	Taken orally (5 ml) every three hours.	

Source; Fatoba et al,.2018

DISCUSSION

The purpose of this paper is to formulate the prevention, control and treatment of various infectious diseases determinant factor of infant mortality among growing infants with medicinal plants being grown in Nigeria, West Africa using the mathematical modelling approach as the research/review scientific point of view.

Prevention and chronic disease management are proactive approaches to health care that stresses prevention at different points along the health care continuum. Health promotion and disease prevention strategies focus on keeping people well and preventing diseases from occurring. These strategies are referred to as primary prevention activities. Prevention is categorised into three levels.

Primary Prevention, which seeks to decrease the number of new cases of a disorder or illness. At this level of prevention we have: the health promotion/education, and Specific protective measures (such as immunisation). Secondary Prevention, which seeks to lower the rate of established cases of a disorder or illness in the population (prevalence). This level essentially involves measures that ensure early diagnosis (such as screening) and prompt management. Tertiary Prevention, which seeks to decrease the amount of disability associated with an existing disorder. This level involves the Disability limitation and Rehabilitation

Disease prevention should focus on strategies that reduce the risk of disease, identify risk factors, or detect disease in its early, most treatable stages. Examples of disease prevention activities include well-baby visits, immunisations, calcium and Vitamin D supplements to reduce the risk of osteoporosis, blood pressure and cholesterol assessments during annual health exams, and screening for illnesses such as breast, cervical, colorectal and prostate cancer (Family Health Teams, 2006).(Di Pierro <u>et al.</u>, 2012; Ramakrishna <u>et al.</u>, 2011).

Prevention and treatment of infant mortality is one of the basic issues in the review, medicinal plants are the magical bullet to prevent and treat basic causes of infant mortality. Medicinal plant can be classified into, primary and secondary prevention strategy which is under the scope of this review work. This is the reason, mechanism of action of this plant must be dealt with, to illustrates the action of medicinal plants on recalcitrant infectious diseases. Some of the mechanisms are as follows, Inhibition of Biofilm Formation, Inhibition of Cell Wall

Construction, Inhibition of Prokaryotic DNA Replication and Inhibition of Energy Production. The details below are as follows

Mechanism of action of action otherwise known as the modus operandi of medicinal plants against infectious diseases should be discuss in the context of this review. If the medicinal plants not reactive against the medicinal plants, then the issue at hand will be in jeopardy. There is need to discuss that activity of it modus operandi against infectious diseases. However, Medicinal plant activity are Promote Cell Wall Disruption and Lysis, Phenolic compounds are a family of aromatic rings consisting of a hydroxyl functional group (-OH) which is alleged to absolute toxicity to microorganisms, although increased reactions of hydroxylation result in microbial cell lysis (Ganesan & Xu (2018). Example are Quercetin, Rutin, Naringenin, sophoraflavanone, Tiliroside, 2, 4, 6-trihydroxy-30-methyl chalcone, etc

Inhibition of Biofilm Formation is one ofthe key features of bacteria developing biofilms are generally 100–1000 times more resistant to antimicrobial drugs while related to their usual planktonic forms (Kahaliw *et al*,.(2017).. Interestingly, numerous researchers have described how flavonoids cause the aggregation of multicellular composites of bacteria and inhibit bacterial growth after aggregation, which indicates that flavonoids are potent antibiofilm compounds. The bioactive flavonoids such as galangin (Dewapriya *et al*,.(2018):, isovitexin (Nath *et al*,.(2018), EGCG and 3-O-octanoyl-epicatechin (Mabona *et al*,.(2013), as well as 5, 7, and 40-trihydroxyflavanol (Prasannabalaji *et al*,.(2012). Induce pseudo multicellular aggregation of *S. aureus* and *S. mutans* [El-Adawi, (2012).

Inhibition of Cell Wall Construction is a another way to eradicated bacteria in infectious diseases. Bacterial cell wall is accountable for osmoregulation, respiration, the transport mechanism, and biosynthesis of lipids. For the execution of these functions, membrane integrity is very important, and its disruption can directly or indirectly cause metabolic dysfunction eventually leads to bacterial death. Catechins (Kariu *et al*,.(2016). attract lipid bilayers of the membrane which involves the following mechanisms [Reygaert,(2014). Catechins form hydrogen bonds, which attract polar head groups of lipids at the membrane edge. Epicatechin

(Spathodea.2016) and epigallocatechin gallate (Muhaisen *et al*,.(2015).alter phospholipids, which can alter structural changes in the cell membrane(Osuntokun *et al*,.2017)..

Inhibition of Prokaryotic DNA Replication is another major tool to prevent infectious diseases. Alkaloids are nitrogenous compounds characterized by their alkaline nature, which aids the inhibition of cell respiration, intercalates with DNA, and inhibits various enzymes involved in replication, transcription, and translation [Zielin *et al.*,2019). Plant-based bioactive compounds such as quercetin (Mozirandi *et al.*,(2019), nobiletin (Zhou *et al.*,2019), myricetin (Mickymaray *et al.*,(2019), tangeritin (Arefin *et al.*,1970) genistein (Vinodhini *et al.*,2016), apigenin (Houlihan *et al.*,(2019), chrysin (Lim *et al.*,2018), kaempferol (Akhalwaya *et al.*,2018), and 3, 6, 7, 30, 40-pentahydroxyflavone (Rawat, *et al.* 2016) have been recognized as noteworthy DNA gyrase inhibitors, which are essential for DNA replication in prokaryotes including *V. harveyi*, *B. subtilis*, *M. smegmatis*, *M. tuberculosis*, and *E. coli* (Vijayakumar, *et al.*,2018).

Inhibition of Energy Production should be discussed to toprevent and treat infectious diseases. Energy production or ATP synthesis is the supreme vital requirement for the existence and development of bacteria as these chemicals are the main source of living systems. The treatment of flavonoids such as isobavachalcone (Durairaj&Dorai,(2010) and 6 prenylapigenin (Bhattacharjee, et al,.2010) with *S. aureus* cause membrane depolarization, resulting in bacterial cell wall lysis [Yasukawa et al,.1998). Similarly, licochalcones inhibited oxygen consumption in M. luteus, interruping the electron transport system eventually killing the bacteria [Antony &Singh(2011). It has been described that flavonoids such as baicalein (Banothuet al,.2017), morin (Mickymarayet al,.2016), silibinin (Chatterjeeet al,.2011), quercetin (Mozirandet al,.2019), isoquercetin (Gazianoet al,.2018), quercitrin (Nefzi& Abdallah,.2016), and silymarin (Chahalet al,.1980) can constrain the F1FO ATPase system of E. coli and result in the obstruction of ATP synthesis.(Walker, et al,.2000)

Inhibition of Bacterial Toxins, It is noteworthy that catechins and other flavonoids can cause bacterial cell wall destruction, resulting in an inability to discharge toxins(Shah, *et al*, 2000). Catechins (Kariu, *et al*, 2016), pinocembrin, kaempferol, EGCG (Prasannabalaji et al, 2012), gallocatechingallate (Muhaisen *et al*, 2015), kaempferol-3-O-rutinoside (Pandian *et al*, 2006),

genistein (Vinodhini *et al*,.2016), quercetin glycoside (Arulmozhi, *et al*,.2018),and proantho cyanidins (Mubarack *et al*,.2012) are suggested to neutralize bacterial toxic factors initiating from *V. cholerae*, *E. coli*, *S. aureus*, *V. vulnificus*, *B. anthracis*, *N. gonorrhoeae*, *and C. botulinum*.(Choi, *et al*,.2007)

Bacterial hyaluronidases are enzymes formed by both Gram-positive and Gram-negative bacteria and directly interact with host tissues, causing the permeability of connective tissues and reducing the viscosity of body fluids due to hyaluronidase-mediated degradation.(Ahmed *et al.*,2016) Flavonoids such as myricetin (Mickymaray,&Aboody (2019) and quercetin (Abe 1974) have been identified as hyaluronic acid lyase inhibitors in Streptococcus equisimilis and Streptococcus agalactiae (Hertel, *et al.*,2006).

CONCLUSION

We conclude that effective control, prevention and treatment of infectious diseases determinant factor among infants by medicinal plants have greater benefits and reduces the rapid progression of infection in the community. It was observed that the mathematical model produced an asymptotically stable population such that the infectious diseases determinant factor among the infants die out from the infant population as time increases when adequate measures (medicinal herbs in large quantity) are encouraged, administered and used for the infants in appropriate quantity.

REFERENCES:

- 1) Binuyo, Adeyemi O. (2015). A deterministic model for the transmission dynamics of infectious diseases among infants. Elixir Appl. Math. 78 (2015) 29761-29764.
- 2) Binuyo, AO and Komolafe O. (2012). Stability Analysis of an SEIRC epidemic model for an infectious disease. Elixir Appl. Math. 42, pps. 6062 6064.
- 3) Faleyimu OI and Oluwalana SA (2008). Medicinal Value of Forest Plant Seeds in Ogun State, Nigeria. *World Journal of Biological Research*, **1**(2): 1-6.
- 4) Cox PA (2005). The seven pillars of ethnomedical wisdom. Ethnobotany, 17:24-34.

- 5) Diekmann O and Heesterbeek JAP. (2000). Mathematical Epidemiology of infectious diseases: model building, analysis and interpretation. Chichester: Wiley
- 6) Hethcote, HW (2000). The mathematics of infectious disease. SIAM Review, volume 42, number 4, pps. 599-653
- 7) Kumar A, Tewari DD, Sharma R, and Pandey VO (2005). Practices of folk phytoveterinery in Devipatan division, Uttar Pradesh. *India Journal Natcon*, **17**(1), 153-161.
- 8) Singh NK and Singh DP (2001). Ethnobotanical survey of Balrampur. Flora- fauna, **7**(2), 59-66
- 9) Wang, Yu-hua, Pei, SJ, and Xu JC (2005). Sustainable management of medicinal plant resources in China: Literature review and implications. *Resources Science*, **24**(4):81-88.
- 10) Bharati S, Rishi P, Koul A. *Azadirachta indica* exhibits chemopreventive action against hepatic cancer: Studies on associated histopathological and ultrastructural changes. Microsc Res Tech. 2012;75(5):586–595.
- 11) NgonoNgane A, Biyiti L, Bouchet P, Nkengfack A, AmvamZollo PH. Antifungal activities of *Piper guineense* of Cameroun. Fitoter. 2003;4(5):464–468.
- 12) Antony ML, Singh SV. Molecular mechanisms and targets of cancer chemoprevention by garlic-derived bioactive compound diallyltrisulfide. Indian J Exp Biol. 2011;49(11):805–816.
- 13) Alam A, Khan N, Sharma S, Saleem M, Sultana S. Chemopreventive effect of *Vitisvinifera* extract on 12-*O*-tetradecanoyl-13-phorbol acetate-induced cutaneous oxidative stress and tumor promotion in murine skin. Pharmacological Research. 2002;46(6):557–564.
- 14) Belman S. Onion and garlic oils inhibit tumor promotion. Carcinogenesis. 1983;4(8):1063–1065. 1983. [PubMed] [Google Scholar]
- 15) Botterweck AAM, Verhagen H, Goldbohm RA, Kleinjans J, Van den Brandt PA. Intake of butylated hydroxyanisole and butylated hydroxytoluene and stomach cancer risk: results from analyses in the Netherlands cohort study. Food ChemToxicol. 2000;38:599–605.
- 16) Mathews TJ, MacDorman MF, Menacker F (January 2002). "Infant mortality statistics from the 1999 period linked birth/infant death data set"(PDF). *National Vital Statistics Reports*. **50** (4): 1–28. doi:10.1037/e558952006-001.
- 17) Labor and Delivery Complications -- the Basics". WebMD. Retrieved 2017-03-16.

- 18) Infant Mortality & Newborn Health". Women and Children First. Retrieved 2017-04-25
- 19) .Hall ES, Venkatesh M, Greenberg JM (November 2016). "A population study of first and subsequent pregnancy smoking behaviors in Ohio". *Journal of Perinatology*. **36** (11):94895 3. doi:10.1038/jp.2016.119.
- 20) Genowska A, Jamiołkowski J, Szafraniec K, Stepaniak U, Szpak A, Pająk A (July 2015). "Environmental and socio-economic determinants of infant mortality in Poland: an ecological study". *Environmental Health*. **14**: 61. doi:10.1186/s12940-015-0048-1.
- 21) .Chan GJ, Lee AC, Baqui AH, Tan J, Black RE (August 2013). "Risk of early-onset neonatal infection with maternal infection or colonization: a global systematic review and meta-analysis". *PLOS Medicine*. **10** (8): e1001502.doi:10.1371/j ournal.pme .1001502. PMC 3747 995
- 22).Nussbaum M (2011). *Creating Capabilities*. The Belknap Press of Harvard University Press. ISBN 978-0-674-05054-9.
- 23) Andrews KM, Brouillette DB, Brouillette RT (2008). "Mortality, Infant". *Encyclopedia of Infant and Early Childhood Development*. Elsevier. pp. 343–359. doi:10.1016/B978-012370877-9.00084-0. ISBN 9780123708779.
- Norton M (April 2005). "New evidence on birth spacing: promising findings for improving newborn, infant, child, and maternal health". *International Journal of Gynaecology and Obstetrics*. 89 Suppl 1: S1-6. doi:10.1016/j.ijgo. 2004.12
- ²⁵⁾ <u>Infant Mortality"</u>. *Maternal and Infant Health | Reproductive Health*. Centers for Disease Control and Prevention. Retrieved 2017-03-07.
- 26) Brown KH, Black RE, Guillermo LR and Hilary CK (1989). Infant-Feeding Practices and their Relationship with Diarrheal and Other Diseases in Huascar (Lima), Peru. *Paediatrics*, **83**(1):31-41
- 27) Jegede AS (2002). The Yoruba Cultural Construction of Health and Illness. *Nordic Journal of African Studies*, 11(3): 322-335.
- 28) Espo M (2002). Infant Mortality and its underlying Determinants in Rural Malawi (Unpublished). University of Tampere Medical School
- 29) UNICEF (2010). Levels and Trends in Child Mortality, Report 2010. UNICEF, New York.

- 30) Vijayakumar, R.; Aboody, M.; AlFonaisan, M.; Turaiki, W.; Mickymaray, S.; Mariappan, P.; Alsagaby, S.;Sandle, T(2012). Determination of Minimum inhibitory concentrations of Common Biocides to Multidrug-Resistant Gram-negative bacteria. Appl. Med. Res. **2016**, 2, 56
- 31) Mesike CG and Mojekwu JN (2012). Environmental Determinants of Child Mortality in Nigeria. *Journal of Sustainable Development*, **5**(1):65-75.
- 32) Fatoba P.O, S. B. Adeyemi, A. A. Adewole and M. T. Fatoba(2018), Medicinal Plants Used in the Treatment of Infant Diseases in South Western Nigeria, Nigerian Journal of Basic and Applied Science (June, 2018), 26(1): 14-22 DOI: http://dx.doi.org/10.4314/njbas.v26i1.2
- 33) Family Health Teams, author. Guide to Health Promotion and Disease Prevention. Ontario: Canada: 2006.
- 34) Pierro F, Rapacioli G, Ferrara T, Togni S. Use of a standardized extract from *Echinacea angustifolia* (Polinaceae) for the prevention of respiratory tract infections'. Altern Med Rev. 2012;17(1):36–41.
- 35) Ramakrishna Y, Goda H, Baliga MS, Munsh AK. Decreasing cariogenic bacteria with a natural alternative prevention therapy using phytochemistry (plant extracts) J ClinPaediatr Dent. 2011;36(1):55–63
- 36) Ganesan, K.; Xu, B(2018). A critical review on phytochemical profile and health promoting e ects of mung bean(Vignaradiata). Food Sci. Hum. Wellness **2018**, 7, 11–33.
- 37) Kahaliw, W.; Ase_a, A.; Abebe, M.; Teferi, M.; Engidawork, E(**2017**). Evaluation of the antimycobacterial activity of crude extracts and solvent fractions of selected Ethiopian medicinal plants. BMC Complement. Altern. Med. **2017**, 17.
- 38) Dewapriya, P.; Khalil, Z.G.; Prasad, P.; Salim, A.A.; Cruz-Morales, P.; Marcellin, E.; Capon, R.J.Talaropeptides A-D(2018): Structure and Biosynthesis of Extensively N-methylated Linear Peptides From an Australian Marine Tunicate-Derived Talaromyces sp. Front. Chem. **2018**, 6.

- 39) Nath, D.; Banerjee, P.; Shaw, M.; Mukhopadhyay, M.K. Bottle Gourd (LagenariaSiceraria)(2018). In Fruit and Vegetable Phytochemicals: Chemistry and Human Health, 2nd ed.; JohnWiley& Sons, Ltd: Hoboken, NJ, USA, 2018; Volume II, pp. 909–920
- 40) Prasannabalaji, N.; Muralitharan, G.; Sivanandan, R.N.; Kumaran, S.; Pugazhvendan, S.R(2012). Antibacterial activities of some Indian traditional plant extracts. Asian Pac. J. Trop. Dis. **2012**, 2, S291–S295
- 41) Mabona, U.; Viljoen, A.; Shikanga, E.; Marston, A.; Van Vuuren, S(2013). Antimicrobial activity of southern African medicinal plants with dermatological relevance: From an ethnopharmacological screening approach, to combination studies and the isolation of a bioactive compound. J. Ethnopharmacol. **2013**, 148, 45–55.
- 42) Oludare temitope Osuntokun, AO, Oluduro, TO Idowu & AO Omotuyi,(2017)Assessment of Nephrotoxicity, Anti-inflammatory and Antioxidant properties of Epigallocatechin, Epi catechin and Stigmasterol phytosterol (synergy) Derived from ethyl acetate stem bark extract of *Spondias mombin* on Wister Rats Using Molecular method of analysis, *Journal of Molecular Microbiology*, Vol.1 No.1:103, Pp 1-11
- 43) BenevidesBahiense, J.; Marques, F.M.; Figueira, M.M.; Vargas, T.S.; Kondratyuk, T.P.; Endringer, D.C.; Scherer, R.; Fronza, M. Potential anti-inflammatory, antioxidant and antimicrobial activities of Sambucusaustralis. Pharm. Biol. **2017**, 55, 991–997.
- 44) El-Adawi, H(2012). Inhibitory effect of grape seed extract (GSE) on cariogenic bacteria. J. Med. Plants Res. **2012**, 6.
- 45) Kariu, T.; Nakao, R.; Ikeda, T.; Nakashima, K.; Potempa, J.; Imamura, T(2016). Inhibition of gingipains and Porphyromonas gingivalisg rowth and biofilm formation by prenyl flavonoids. J. Periodontal Res. **2016**, 52, 89–96.
- 46) Reygaert, W.C(2014). The antimicrobial possibilities of green tea. Front. Microbiol. **2014**, 5.
- 47) SpathodeacampanulataBeauv(2016). In SpringerReference; Springer: Berlin, Germany, 2016.
- 48) Muhaisen, H.M.H.; Ab–Mous, M.M.; Ddeeb, F.A.; Rtemi, A.A.; Taba, O.M.; Parveen, M(2015). Antimicrobial agents from selected medicinal plants in Libya. Chin. J. Integr. Med. **2015**, 22, 177–184.

- 49) Zielin´ ska, S.; Wójciak-Kosior, M.; Dzia gwa-Becker, M.; Glen´ sk, M.; Sowa, I.; Fijałkowski, K.; Rura´ nska-Smutnicka, D.; Matkowski, A.; Junka, A(2019). The Activity of Isoquinoline Alkaloids and Extracts from Chelidonium majus against Pathogenic Bacteria and Candida sp. Toxins **2019**, 11, 406.
- 50) Mozirandi, W.; Tagwireyi, D.; Mukanganyama, S(2019). Evaluation of antimicrobial activity of chondrilla sterol isolated from Vernonia adoensis (Asteraceae). BMC Complement. Altern. Med. **2019**, 19.
- 51) Zhou, J.-X.; Braun, M.; Wetterauer, P.; Wetterauer, B.; Wink, M(2019). Antioxidant, Cytotoxic, and Antimicrobial Activities of Glycyrrhizaglabra L., Paeonialactiflora Pall., and Eriobotrya japonica (Thunb.) Lindl. Extracts. Medicines **2019**, 6, 43
- 52) Mickymaray, S.; Al Aboody, M.S(2019). In Vitro Antioxidant and Bactericidal E_cacy of 15 Common Spices: Novel Therapeutics for Urinary Tract Infections? Medicina **2019**, 55, 289
- 53) Arefin, M.K.; Rahman, M.M.; Uddin, M.Z.; Hassan, M.A. Angiosperm flora of Satchari National Park, Habiganj, Bangladesh. Bangladesh J. Plant. Taxon. **1970**, 18, 117–140.
- 54) Vinodhini, R.; Moorthy, K.; Suresh, M(2016). Incidence and virulence traits of Candida dubliniensis isolated from clinically suspected patients. Asian J. Pharm. Clin. Res. **2016**, 9, 77.
- 55) Houlihan, A.J.; Conlin, P.; Chee-Sanford, J.C(2019). Water-soluble exudates from seeds of Kochia scoparia exhibit antifungal activity against Colletotrichumgraminicola. PLoS ONE **2019**, 14, e0218104
- 56) Lim, S.S.; Selvaraj, A.; Ng, Z.Y.; Palanisamy, M.; Mickmaray, S.; Cheong, P.C.H.; Lim, R.L.H(2018). Isolation of actinomycetes with antibacterial activity against multi-drug resistant bacteria. Malays. J. Microbiol. **2018**.
- 57) Akhalwaya, S.; van Vuuren, S.; Patel, M(2018). An in vitro investigation of indigenous South African medicinal plants used to treat oral infections. J. Ethnopharmacol. **2018**, 210, 359–371.

- 58) Rawat, S.; Jugran, A.K.; Bahukhandi, A.; Bahuguna, A.; Bhatt, I.D.; Rawal, R.S.; Dhar, U(2016). Anti-oxidant and anti-microbial properties of some ethno-therapeutically important medicinal plants of Indian Himalayan Region. 3 Biotech. **2016**, 6, 154
- 59) Vijayakumar, R.; Sandle, T.; Al-Aboody, M.S.; AlFonaisan, M.K.; Alturaiki, W.; Mickymaray, S.; Premanathan, M.; Alsagaby, S.A(2018). Distribution of biocide resistant genes and biocides susceptibility in multidrug-resistant Klebsiellapneumoniae, Pseudomonas aeruginosa and Acinetobacterbaumannii A first report from the Kingdom of Saudi Arabia. J. Infect. Public Health **2018**, 11, 812–816.
- 60) Durairaj, B.; Dorai, A(2010). Antiplatelet activity of white and pink NelumbonuciferaGaertn flowers. Braz. J. Pharm. Sci. **2010**, 46, 579–583.
- 61) Bhattacharjee, I.; Chatterjee, S.K.; Chandra, G(2010). Isolation and identification of antibacterial components in seed extracts of Argemonemexicana L. (Papaveraceae). Asian Pac. J. Trop. Med. **2010**, 3, 547–551.
- 62) Yasukawa K, Demitrijevic SM, Evans FJ, Kawabata S, TakidoM(1998). Inhibitory effect of Prunus Cortex extract and its component, octacosylferulate, on tumor promotion by 12-*O*-tetradecanoylphorbol-13-acetate in two-stage carcinogenesis in mouse skin. Phytotherapy Research. 1998;12(4):261–265.
- 63) Antony ML, Singh SV(2011). Molecular mechanisms and targets of cancer chemoprevention by garlic-derived bioactive compound diallyltrisulfide. Indian J Exp Biol. 2011;49(11):805–816.
- 64) Banothu, V.; Neelagiri, C.; Adepally, U.; Lingam, J.; Bommareddy, K(2017). Phytochemical screening and evaluation of in vitro antioxidant and antimicrobial activities of the indigenous medicinal plant Albiziaodoratissima. Pharm. Biol. **2017**, 55, 1155–1161.
- 65) Mickymaray, S.; Al Aboody, M.S.; Rath, P.K.; Annamalai, P.; Nooruddin, T(2016). Screening and antibacterial effcacy of selected Indian medicinal plants. Asian Pac. J. Trop. Biomed. **2016**, 6, 185–191

- 66) Chatterjee, S.K.; Bhattacharjee, I.; Chandra, G(2011). Isolation and identification of bioactive antibacterial components in leaf extracts of Vangueriaspinosa (Rubiaceae). Asian Pac. J. Trop. Med. **2011**, 4, 35–40.
- 67) Mozirandi, W.; Tagwireyi, D.; Mukanganyama, S(2019). Evaluation of antimicrobial activity of chondrillasterol isolated from Vernoniaadoensis (Asteraceae). BMC Complement. Altern. Med. **2019**, 19.
- 68) Gaziano, R.; Campione, E.; Iacovelli, F.; Marino, D.; Pica, F.; Di Francesco, P.; Aquaro, S.; Menichini, F.; Falconi, M.; Bianchi, L(2018). Antifungal activity of Cardiospermumhalicacabum L. (Sapindaceae) against Trichophytonrubrum occurs through molecular interaction with fungal Hsp90. Drug Des. Dev. 2018, 12, 2185–2193
- 69) Nefzi, A.; Ben Abdallah, R.A(2016). Antifungal activity of aqueous and organic extracts from Withaniasomnifera L. against Fusariumoxysporum f. sp. Radicislycopersici. J. Microb. Biochem. Technol. **2016**, 8. [CrossRef
- 70) Chahal, S.S.; Matthews, H.R.; Bradbury, E.M.(1980) Acetylation of histone H4 and its role in chromatin structure and function. Nature **1980**, 287, 76–79
- 71) Walker, E.H.; Pacold, M.E.; Perisic, O.; Stephens, L.; Hawkins, P.T.; Wymann, M.P.; Williams, R.L(2000) Structure determinants of phosphoinositide 3-kinase inhibition by wortmannin, LY294002, quercetin, myricetin and staurosporine. Mol. Cell **2000**, 6, 909–919
- 72) Shah, S.; Stapleton, P.D.; Taylor, P.W(2007). The polyphenol (□)-epicatechingallate disrupts the secretion of virulence-related proteins by Staphylococcus aureus. Lett. Appl. Microbiol. **2007**, 46, 181–185
- 73) Kariu, T.; Nakao, R.; Ikeda, T.; Nakashima, K.; Potempa, J.; Imamura, T(2016). Inhibition of gingipainsAndPorphyromonasgingivalisgrowth and biofilm formation by prenyl flavonoids. J. Periodontal Res. **2016**, 52, 89–96.
- 74) Prasannabalaji, N.; Muralitharan, G.; Sivanandan, R.N.; Kumaran, S.; Pugazhvendan, S.R(2012). Antibacterial activities of some Indian traditional plant extracts. Asian Pac. J. Trop. Dis. **2012**, 2, S291–S295.

- 75) Muhaisen, H.M.H.; Ab–Mous, M.M.; Ddeeb, F.A.; Rtemi, A.A.; Taba, O.M.; Parveen, M(2015). Antimicrobial agents from selected medicinal plants in Libya. Chin. J. Integr. Med. **2015**, 22, 177–184.
- 76) Pandian, M.R.; Banu, G.S.; Kumar, G. A(2006) study of the antimicrobial activity of Alangiumsalviifolium. Indian J. Pharm. **2006**, 38, 203.
- 77) Vinodhini, R.; Moorthy, K.; Suresh, M(2016). Incidence and virulence traits of Candida dubliniensis isolated from clinically suspected patients. Asian J. Pharm. Clin. Res. **2016**, 9, 77
- 78) Arulmozhi, P.; Vijayakumar, S.; Kumar, T.(2018) Phytochemical analysis and antimicrobial activity of some medicinal plants against selected pathogenic microorganisms. Microb. Pathog. **2018**, 123, 219–226.
- 79) Mubarack, H.; Doss, A.; Vijayasanthi, M.; Venkataswamy, R(2012). Antimicrobial drug susceptibility of Staphylococcus aureus from subclinical bovine mastitis in Coimbatore, Tamilnadu, South India. Vet. World **2012**, 5, 352
- 80) Choi, O.; Yahiro, K.; Morinaga, N.; Miyazaki, M.; Noda, M(2007). Inhibitory e_ects of various plant poly phenols on the toxicity of Staphylococcal _-toxin. Microb. Pathog. **2007**, 42, 215–224.
- 81) Ahmed, S.I.; Hayat, M.Q.; Tahir, M.; Mansoor, Q.; Ismail, M.; Keck, K.; Bates, R.B(2016). Pharmacologically active flavonoids from the anticancer, antioxidant and antimicrobial extracts of Cassia angustifoliaVahl. Bmc Complement. Altern. Med. **2016**, 16. [
- 82) Mickymaray, S.; Al Aboody, M.S(2019). In Vitro Antioxidant and Bactericidal E_cacy of 15 Common Spices: Novel Therapeutics for Urinary Tract Infections? Medicina **2019**, 55, 289.
- 83) Abe H. Catalase. In method of enzymatic analysis. New York: Academic Press; 1974. pp. 673–684.
- 84) Hertel, W.; Peschel, G.; Ozegowski, J.-H.; Müller, P.-J.(2006) Inhibitory Effects of Triterpenes and Flavonoids on the Enzymatic Activity of Hyaluronic Acid-Splitting Enzymes. Arch. Pharm. **2006**, 339, 313–318