

[Type text]

Arbuscular Mycorrhizal and Root colonizing Dark Septate Endophytic Fungal Associations in *Urginea indica* and *Urginea wightii* accessions.

ABSTRACT

Urginea indica belongs to Hyacinthaceae family. It is also known as Indian squill, commonly called as wild Onions. The over exploitation and habitat degradation has resulted in the loss of habitat and it has caused genetic depletion and loss of genetic diversity. Immediate Measures have to be undertaken for conservation, to save this economically important medicinal plant. The present investigation is an attempt to highlight the occurrence of arbuscular mycorrhizal (AM) and Dark Septate Endophyte (DSE) fungal associations found in the root bulbs of *Urginea indica*. 8 Accessions of *Urginea* were examined in *Urginea indica kunth* and *Urginea wightii* accessions, collected from various regions of Karnataka and south India. Fungal Symbiosis was present in all the accessions, with 100% root infection. *Urginea* has DSE fungal association and AM Association.

This is the first report on the DSE fungal association in *Urginea indica kunth*. In addition to the prevalence of AM fungal symbiosis, the role of DSE is still in infancy. Though no clear relationship between AM and DSE fungal colonizations was recognized. Our studies suggest the coexistence of AM and DSE fungal colonizations are common in terrestrial habitats especially in the xerophytic environment. DSE-plant symbiosis should not be limited to nutritional uptake but mediates other parameters such as drought resistance, stress tolerance and herbivore resistance. Our results show 100% infection by AM and DSE fungal colonies. Infection levels were high throughout the year, the highest levels occurring in the most nutrient-stressed situations. Their occurrence and variation in different accessions is discussed below.

Key words: Arbuscular Mycorrhiza, Dark Septate Endophytic fungi, Urginea

INTRODUCTION

Urginea indica belongs to Hyacinthaceae family. It is also known as Indian squill, commonly called as wild Onions, (vernacular names Van Pyaz, Kadu erulli). It is a perennial bulbous plant with roots measuring about 8-10 inches in length. (Vishal R. Kamble .2012). It is endemic to India, Africa and Mediterranean regions. (Shiva Kameshwari 2012) The Genus *Urginea* (Syn. *Drimia*). Ethno-medicinally bulbs of *U. indica* has proved to be antiulcerous, antinematodal, antitumorous, anthelmintic, antiarthritic properties and is used to cure skin diseases like warts, abscesses, boils, cardiac diseases, antidote to scorpion sting (M.S. Chittoor 2012). The bulbs contain many compounds that defend cells against free radicals by blocking the development of heart diseases, cancer, dropsy, edema, Dog bites, cuts, wounds, infertility in man and numerous other ailments. Due to these many medicinal properties of *Urginea indica* bulbs have found its place in British and European Pharmacopoeias. (M.N. Shiva kameshwari, G. Paramasivan 2012). As per IUCN criteria, the threat status of *Urginea indica* is VULNERABLE for Chattisgarh and Madhya Pradesh (K.C Joshi, M.S Negi 2010). Hence conservation through germ plasm and awareness is necessary for the sustainable utilization of this medicinally important plant. It has been established that the presence of mycorrhiza (AM) fungi is important for coastal sand dune vegetation. (K R. Beena 2000), (V. Jaiswal 2001) (Vishal kamble 2012).

There are a number of studies carried on VAM Fungi they have shown that vesicular-arbuscular mycorrhizal (VAM) infection can significantly improve the phosphorus nutrition and yield of plants grown in soils of low fertility (Mosse, 1973). Crop plants have been widely studied and relatively little

[Type text]

information has accumulated about the significance of VA mycorrhizal infection in natural and semi-natural wild variety of plant communities. Recently, there has been an increasing awareness on another group of anamorphic Ascomycetous fungi, which also frequently colonize roots of plants growing in various habitats. These fungi termed as dark septate endophyte (DSE) fungi produce dark septate or hyaline hyphae and microsclerotia. The DSE fungi often coexist with different types of mycorrhizal fungi, including the AM fungi. It is therefore essential to understand the interaction of these fungi as they inhabit the same niche within plant roots. (Kumar seerangan and Muthukumar 2014). As part of an experimental study on the ecological role of VAM, a quantitative analysis was carried out to study the infection level in *Urginea indica* and *Urginea wightii* species growing in semi-natural, dry arid and wet soil. The other most studied groups of fungal root endophytes, the so-called Dark Septate Endophytes (DSE), are a polyphyletic aggregate of fungi belonging to Class 4 of non-clavicipitaceous endophytes (Rodriguez R, White 2009) which is broadly defined by the endophytic life strategy and presence of intraradical dark septate hyphae. Dark septate endophytes are an ubiquitous group of hyaline or darkly pigmented, sterile, septate endophytic fungi that colonize living plant organs, especially roots without causing any apparent or negative effects to the host plant (Jumpponen and Trappe 1998). These fungi usually form in root cortical cells clusters of inflated, rounded, thick-walled cells called microsclerotia (Jumpponen and Trappe 1998). There associations has been found in different plant species, suggesting the lack of host specificity (Jumpponen and Trappe 1998). Dark septate endophyte (DSE) fungi often contain melanin, which is helpful under unfavourable or stressful conditions like extreme temperature, drought, etc. (Jumpponen and Trappe 1998). The role played by DSE fungi is currently unresolved, recent studies indicate their potential to function as plant growth promoters both under favourable (Wu and Guo 2008; Andrade-Linares et al. 2011) and unfavourable conditions (Barrow and Osuna 2002; Jumpponen 2001).

MATERIALS AND METHODS

The intensity of vesicular-arbuscular mycorrhizal infection was assessed in over 10 Accessions of *Urginea indica* collected from various parts of Karnataka and across South India.

Urginea indica accessions were collected from Udupi, Sithampundi, Kerala, Shimoga, Karwar, Magadi. *Urginea whitii* 4 accessions collected from Nagarhole, Yediyur, Gulbarga, and Bidadi.

The study was carried in two parts. In the first, a general survey of the infection and the percentage of infection was done number of vegetation and in 3 soil samples. In the second, *Urginea* grown as experimental plant in Sterilized soil grown in green house for a year and the infection levels in roots were analysed for AM and DSE fungal colonization.

Evaluation of AM Fungal colonization

Freshly collected root samples were washed gently and made free from soil particles and cut into small segments of approximately 0.5cm. varying from 5 to 10 pieces, depending on the size of the sample. The roots were fixed in FAA for 24 hours. Roots were then cleared in 10% KOH and autoclaved (heated), once cooled they were acidified with (1N) HCL for 10 to 15 minutes. Later they were stained in Trypan blue (Phillips and Hayman, 1970) the concentration of Trypan blue was reduced to (0.2 % in lacto glycerol) prepared in Lactoglycerol and the stained roots were again heated /autoclaved 15 minutes under 60 pressure (lbs). The stained roots were mounted on a glass on slides and examined under

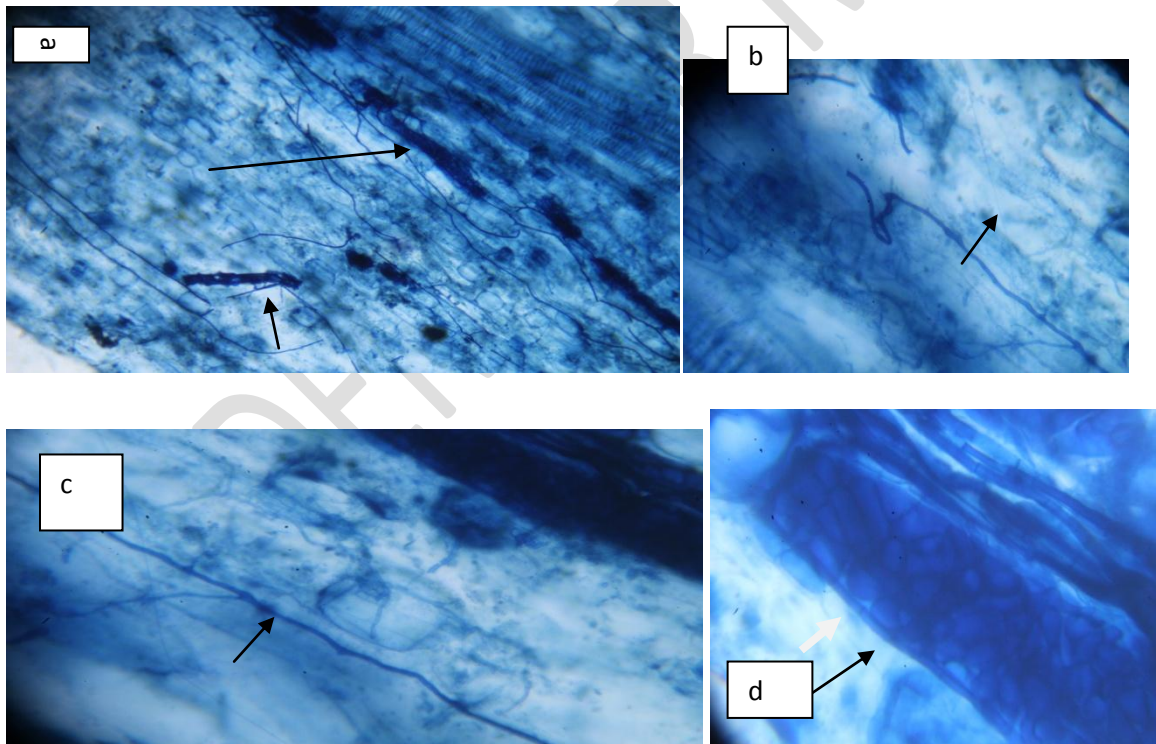
[Type text]

Magnus compound microscope for the AM and DSE fungal structures. The presence of characteristic darkly pigmented or hyaline septate hyphae, and, when present, microsclerotia or moniliform cells were used to characterize DSE fungal colonization. Only root samples possessing arbuscules or arbusculate coils were considered to be arbuscular mycorrhizal.

Root colonization:

The percentage of total root length colonization and root length with different fungal structures for AM fungi (aseptate hyphae, hyphal structures) The percentage of AM infection was estimated by the root-side technique of Nicolson (1955). All infected and uninfected segments were counted. The percentage of infection was calculated using the formula

$$\text{Per cent of mycorrhizal colonization} = \frac{\text{Number of root segments colonized}}{\text{Total number of root segments examined}} \times 100$$



[Type text]

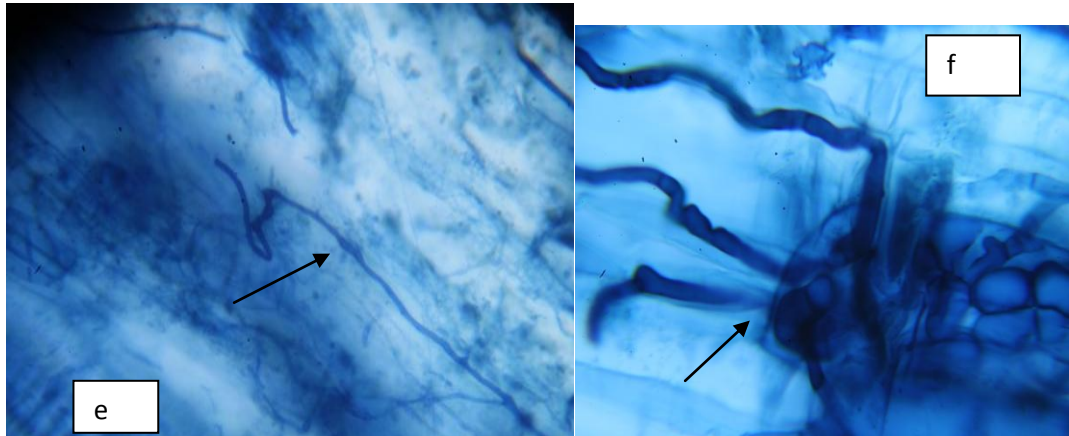
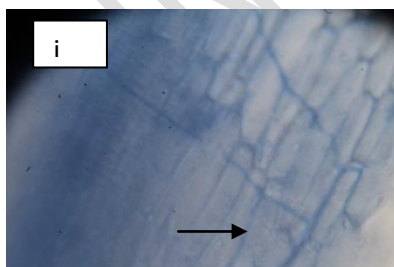
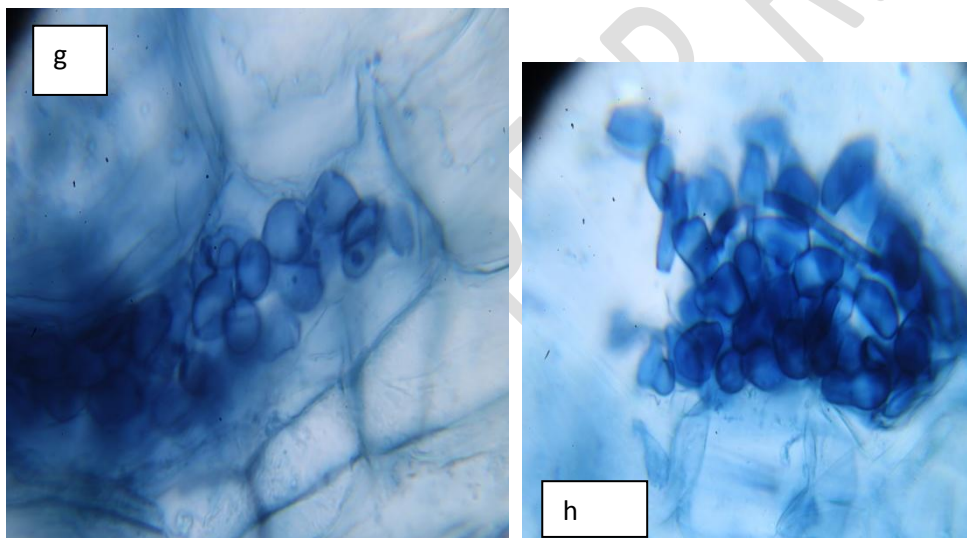


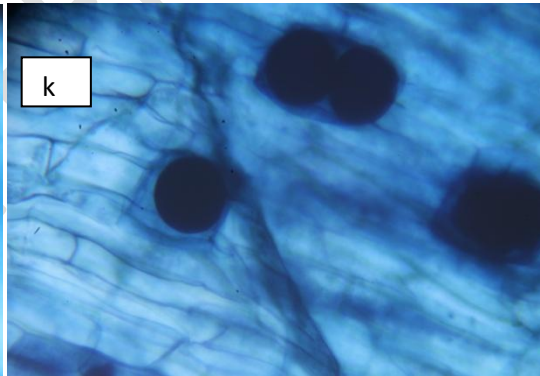
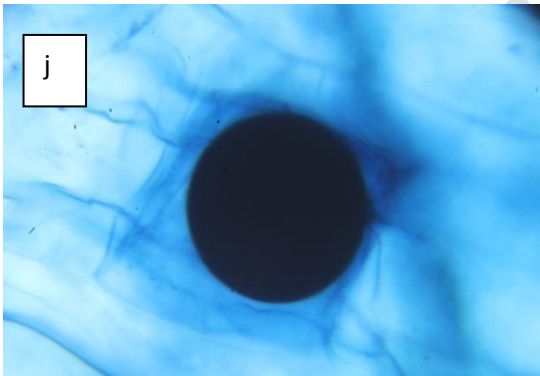
Figure 1: Mycorrhizal fungi, dark septate endophytes and other endophytic fungi inhabiting the same roots of *Urginea indica* accessions and *Urginea wightii* accessions

a) and d) clustered intracellular microsclerotia b) ,c) and e) dark septate hyphae



g) and h) aggregation of vesicles i) the presence of arbuscules

[Type text]



[Type text]

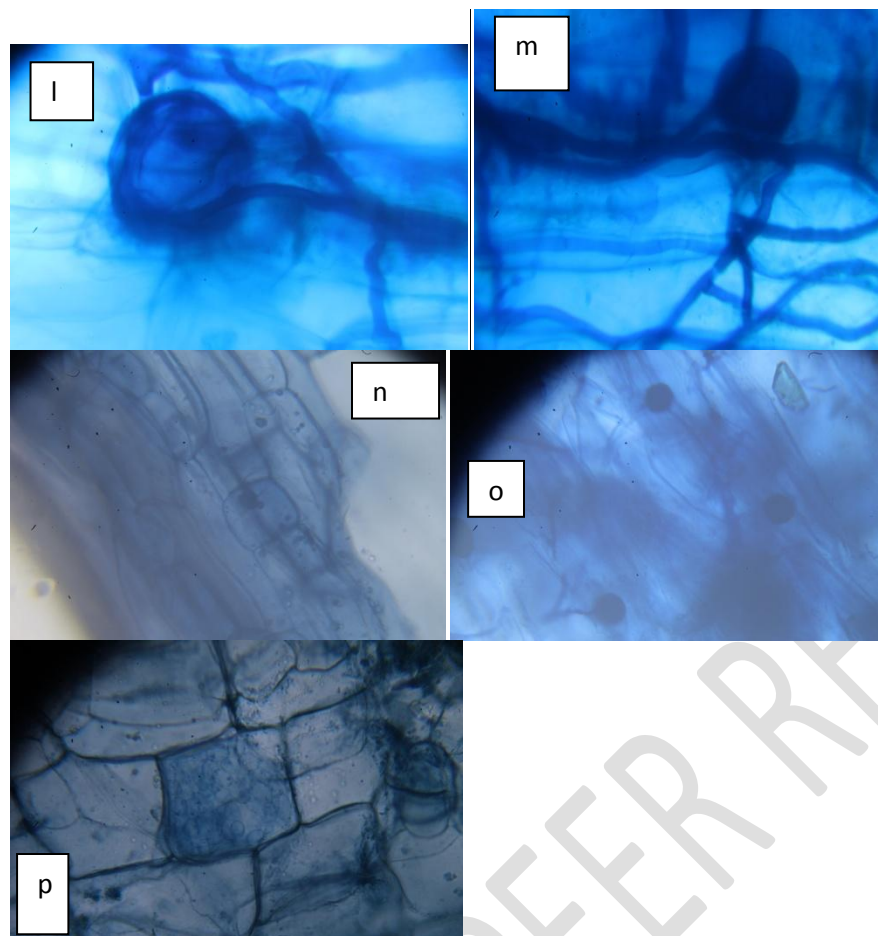


Figure 2)

j) and k) Terminal vesicles. l) and m) intradical hyphae and terminal vesicles. . o) distribution of vesicles P) arbuscules.

RESULTS and Discussion

Occurrence of AM and DSE Fungal Association

Fungal endophytes are defined as mycobionts which live inside living plant tissues, lack localized interfaces or specialized hyphae for nutrient transfer, their development is not synchronized with plant development and the plant does not nutritionally benefit from the symbiosis [Brundrett M.2006]. The present investigation in 10 accessions of *Urgine indica* and *whitii* Accessions. were assessed for AM and DSE fungal association. of this all of them showed 100 % infection of Arbuscular Mycorrhizal and Dark Septate endophytes, the presence of ectomycorrhizae, in few Accessions. The presence of Hyphal structures (hyphae, inter & intra cellular) was dominant, presence of vesicles and spores were present in all the accessions. Infection levels were high, throughout the year, the highest levels of infection occurring in the most nutrient-stressed situations as observed in Thiruchendur Accession.

Roots were studied at different lengths about an average of 10 segments (0.5cms). It was observed that there was extensive hyphae formation in 0.5-2 cms of the roots. Hartings Net was observed 0.2-0.4 the presence of Vesicles, clustered vesicles and occurrence of spores was seen. predominant occurrence of spores was seen beyond 3.5 cms of the root length from the soil. It was also observed that the fungus grows throughout the cortex, but not the endodermis and the stele. The fungus penetrates from

[Type text]

cell to cell forming a new coil. The Intracellular hyphae were usually found in the intermediate layers of the cortical cells of parenchyma.

The unresolved ecophysiological significance of Dark Septate Endophytes (DSE) may be in part due to existence of morphologically indistinguishable cryptic species in them (Tereza Lukešová et al 2015). The main objective in this study was to study and report the presence of AM fungus of Dark septate mycorrhizae.

Conclusion:

The beneficial effect of indigenous AM fungi on the nutrition of agricultural plants depends on the abundance and type of fungi present in the soil. (Abbott and Robson, 1982) The presence of any mycorrhizal associations found the roots of vascular plants plays an important role on sustainable agriculture its management. But the potential for employing AM fungi and the role of Dark septate mycorrhizae in agriculture and protection of its habitat requires more attention. In the present study indigenous AM fungi and dark septate mycorrhizal association present in different 8 accessions were studied. From our studies we would like to conclude that DSE are prevalent in various habitat, not much understanding has been achieved on DSE fungus, it cannot be overlooked as it has been stated to be multifunctional, such as drought resistance, environmental tolerance. The production of melanin tissues may deter mammals and other pathogen root infections. The dynamics of this plant community, its host response under natural conditions may be difficult to determine in a simple and controlled preliminary experiment. Utricularia being a xerophyte, subjected to unfavourable, arid and dry environment, it can be clearly drawn to conclusion that the presence of the fungal associations is necessary to sustain itself. Exactly how it does, associations are symbiotic or not, still investigation has to be done, if so the role of each component is still yet to be confirmed. Our experiment suggests that DSE are abundant and their ecological significance, in relation to AM fungi and other ectomycorrhizae has to be significantly understood. This is a one small step before a giant leap.

Acknowledgments

The author conveys sincere thanks to UGC for providing funding through Rajiv Gandhi National Fellowship. Department of Botany, Bangalore University, Bangalore.

References

1. M. Brundrett, "Mycorrhizal associations and other means of nutrition of vascular plants: understanding the global diversity of host plants by resolving conflicting information and developing reliable means of diagnosis," *Plant and Soil*, vol. 320, no. 1-2, pp. 37–77, 2009.
2. W. Kai and Z. Zhiwei, "Occurrence of arbuscular mycorrhizas and dark septate endophytes in hydrophytes from lakes and streams in southwest China," *International Review of Hydrobiology*, vol. 91, no. 1, pp. 29–37, 2006.
3. K. P. Radhika and B. F. Rodrigues, "Arbuscular mycorrhizae in association with aquatic and marshy plant species in Goa, India," *Aquatic Botany*, vol. 86, no. 3, pp. 291–294, 2007.
4. J. F. de Marins, R. Carrenho, and S. M. Thomaz, "Occurrence and coexistence of arbuscular mycorrhizal fungi and dark septate fungi in aquatic macrophytes in a tropical river-floodplain system," *Aquatic Botany*, vol. 91, no. 1, pp. 13–19, 2009.
5. P. Kohout, Z. Šýkrová, M. Čtvrtíková et al., "Surprising spectra of root-associated fungi in submerged aquatic plants," *FEMS Microbiology Ecology*, vol. 80, no. 1, pp. 216–235, 2012.
6. S. P. Miller and R. R. Sharitz, "Manipulation of flooding and arbuscular mycorrhiza formation influences growth and nutrition of two semiaquatic grass species," *Functional Ecology*,

[Type text]

- vol. 14, no. 6, pp. 738–748, 2000.
7. K. J. Stevens, S. W. Spender, and R. L. Peterson, "Phosphorus, arbuscular mycorrhizal fungi and performance of the wetland plant *Lythrum salicaria* L. under inundated conditions," *Mycorrhiza*, vol. 12, no. 6, pp. 277–283, 2002.
 8. S. Dickson, "The *Arum-Paris* continuum of mycorrhizal symbioses," *New Phytologist*, vol. 163, no. 1, pp. 187–200, 2004.
 9. S. Dickson, F. A. Smith, and S. E. Smith, "Structural differences in arbuscular mycorrhizal symbioses: more than 100 years after Gallaud, where next?" *Mycorrhiza*, vol. 17, no. 5, pp. 375–393, 2007.
 10. Y. Wang, Q. Qiu, Z. Yang, Z. Hu, N. F.-Y. Tam, and G. Xin, "Arbuscular mycorrhizal fungi in two mangroves in South China," *Plant and Soil*, vol. 331, no. 1, pp. 181–191, 2010.
 11. T. Muthukumar and K. Udaiyan, "Arbuscular mycorrhizas of plants growing in the Western Ghats region, Southern India," *Mycorrhiza*, vol. 15, pp. 297–313, 2005. S. Dharmarajan, K. Kannan, and C. Lakshminarasimhan,
 12. "Vesicular-arbuscular (VA) mycorrhizal status of some aquatic and marshy plants," *Acta Botanica Indica*, vol. 21, pp. 161–171, 1993.
 13. T. Muthukumar and K. Prabha, "Arbuscular mycorrhizal and septate endophyte fungal associations in lycophytes and ferns of south India," *Symbiosis*, vol. 59, no. 1, pp. 15–33, 2013.
 14. T. Muthukumar and K. Udaiyan, "Seasonality of vesicular-arbuscular mycorrhizae in sedges in a semi-arid tropical grassland," *Acta Oecologica*, vol. 23, no. 5, pp. 337–347, 2002. T. Muthukumar, K. Udaiyan, and P. Shanmughavel, "Mycorrhiza in sedges—an overview," *Mycorrhiza*, vol. 14, no. 2, pp. 65–77, 2004.
 15. M. Brundrett and B. Kendrick, "The roots and mycorrhizas of herbaceous woodland plants. II. Structural aspects of morphology," *New Phytologist*, vol. 114, no. 3, pp. 469–479, 1990.
 16. J. W. Gerdemann, "Vesicular-arbuscular mycorrhizae formed on maize and tuliptree by *Endogone fasciculata*," *Mycologia*, vol. 57, pp. 562–575, 1965.
 17. W. K. Cornwell, B. L. Bedford, and C. T. Chapin, "Occurrence of arbuscular mycorrhizal fungi in a phosphorus-poor wetland and mycorrhizal response to phosphorus fertilization," *American Journal of Botany*, vol. 88, no. 10, pp. 1824–1829, 2001.
 18. S. D. Turner, J. P. Amon, R. M. Schneble, and C. F. Friese, "Mycorrhizal fungi associated with plants in ground-water fed wetlands," *Wetlands*, vol. 20, no. 1, pp. 200–204, 2000.
 19. T. Muthukumar and K. Prabha, "Arbuscular mycorrhizal and septate endophyte fungal associations in lycophytes and ferns of south India," *Symbiosis*, vol. 59, no. 1, pp. 15–33, 2013.
 20. T. Muthukumar and K. Udaiyan, "Seasonality of vesicular-arbuscular mycorrhizae in sedges in a semi-arid tropical grassland," *Acta Oecologica*, vol. 23, no. 5, pp. 337–347, 2002.
 21. T. Muthukumar, K. Udaiyan, and P. Shanmughavel, "Mycorrhiza in sedges—an overview," *Mycorrhiza*, vol. 14, no. 2, pp. 65–77, 2004.
 22. M. Brundrett and B. Kendrick, "The roots and mycorrhizas of herbaceous woodland plants. II. Structural aspects of morphology," *New Phytologist*, vol. 114, no. 3, pp. 469–479, 1990.
 23. D. E. Evans, "Aerenchyma formation," *New Phytologist*, vol. 161, no. 1, pp. 35–49, 2004.
 24. J. W. Gerdemann, "Vesicular-arbuscular mycorrhizae formed on maize and tuliptree by *Endogone fasciculata*," *Mycologia*, vol. 57, pp. 562–575, 1965.
 25. M. Yamato, "Morphological types of arbuscular mycorrhizal fungi in roots of weeds on vacant land," *Mycorrhiza*, vol. 14, no. 2, pp. 127–131, 2004.
 26. T. R. Cavagnaro, L.-L. Gao, F. A. Smith, and S. E. Smith, "Morphology of arbuscular mycorrhizas is influenced by fungal identity," *New Phytologist*, vol. 151, no. 2, pp. 469–475, 2001.
 27. S. E. Smith, F. A. Smith, and I. Jakobsen, "Functional diversity in arbuscular mycorrhizal (AM) symbioses: the contribution of the mycorrhizal P uptake pathway is not correlated with mycorrhizal responses in growth or total P uptake," *New Phytologist*, vol. 162, no. 2, pp. 511–524, 2004.
 28. J. W. Gerdemann, and T. H. Nicolson, "Spores of mycorrhizal *Endogone* species extracted from soil by wet sieving and decanting," *Trans. Br. Mycol. Soc.*, 46: 1963, 235–244.

[Type text]

30. K.C. Joshi, M.S. Negi and A.D. Tiple, Achanakmar-Amarkantak Biosphere Reserve. Biosphere Reserve Information Series (BRIS), 2(1-2), (Tropical Forest Research Institute Jabalpur, 2010, pp. 1-158).
31. K.R. Beena, N.S. Raviraja and K.R. Sridhar, Association of arbuscular mycorrhizal fungi with *Launaea sarmentosa* on maritime sand dunes of west coast of India, *Kavaka* 25: 1997, 53-60.
32. K.R. Beena, N.S. Raviraja, A.B. Arun and K.R. Sridhar, Diversity of arbuscular mycorrhizal fungi on the coastal sand dunes of west coast of India, *Current Science* 79: 2000, 1459-1466.
33. M. Brundrett, L. Melville, L. Peterson, Practical Methods in Mycorrhizal Research. Mycologue Publications, (University of Guelph, Guelph, Ontario, 1994).
34. M.N. Shiva Kameshwari, A.B. Lakshman and G. Paramasivam, 2012, Biosystematics studies on medicinal plant *Urginea indica* Kunth. liliaceae - A review. *Int. J. of Pharm. & Life Sci.*, 3 (1): 2012, 1394-1406.
35. M.N. Shiva Kameshwari, Biosystematics studies on *Urginea indica* Kunth. Liliaceae. (Abs) Nat. Conf. on Forest Biodiversity Resources: Exploitation Conservation & Management, 21-22 March 2006 CBFS, Madurai Kamaraj University: Madurai, 24-25.
36. M.S. Chittoor, A.J. Roger Binny, S. Yadlapalli, A. Cheruku, C. Dandu, and Y. Nimmanapalli, Anthelmintic and antimicrobial studies of *Drimia indica* (Roxb.) Jessop. bulb aqueous extracts *Journal of Pharmacy Research*, 5(5): 2012, 3677-3686.
37. M.V. Gokhale, S.S. Shaikh, N.S. Chavan, Floral survey of wet coastal and associated ecosystems of Maharashtra, *Indian Journal of Geo Marine Sciences*, 40 (5): 2011, 725-730.