

# INVIVO ANTIOXIDANT EFFECT OF COCONUT WATER IN WISTAR ALBINO RATS

## ABSTRACT

**Aim:** The present study was designed to investigate the effect of coconut water extract on antioxidant biomolecules in wistar albino rats.

**Methodology:** Twenty five (25) animals randomly selected were divided into five groups of 5 animals per group was used for the study. Group 1 served as the control group while groups 2, 3, 4, and 5 served as the test groups that received 10 ml, 20 ml, 30 ml, and 40 ml of the coconut water extract for 28 days. The rats were sacrificed after 28 days and the blood samples were collected for biochemical analysis,

**Result:** From the result obtained there is a significant increase ( $p < 0.05$ ) between the control group (group 1) and the test group (group 2) that received 20 ml of the coconut water extract for MDA. There is a significant increase ( $p < 0.05$ ) between the control group (group 1) and the test groups (group 2 and 5) that received 10 ml and 50 ml of the coconut water extract for SOD. Also, there is a significant increase ( $p < 0.05$ ) between the control group (group 1) and the test groups (groups 2, 3 and 4) that received 20 ml, 30 ml and 40 ml of the coconut water extract for Catalase. For GSH and Vitamin C, there is a significant increase ( $p < 0.05$ ) between the control group (group 1) and the test groups (groups 2, 3, 4 and 5) that received 10 ml, 20 ml, 30 ml and 40 ml of the coconut water extract.

**Conclusion:** The present investigation showed that the coconut water extract increased antioxidant properties in wistar albino rats and may also be used pharmacologically in the treatment of diseases implicated by free radicals.

**Keywords:** Coconut; antioxidant; Catalase; Malondialdehyde; Superoxide dismutase; Vitamin C, Reduced glutathione.

## 1. INTRODUCTION

For thousands of years, coconut products have held a respected and valuable place in local folk medicine. Coconut water has a host of yet scientifically unproven but traditional uses in cultures all over the world [1]. From ancient times in Africa, reports support the position that about 85% of the world's population rely on coconut fruit in traditional medicine [2]. It is used to conquer irregular or painful menstruation and also taken during pregnancy to give the unborn babies strength and vitality [3]. It is also used to boost semen quality and induce libido [4]. Coconut water contains numerous antioxidant compounds that have the ability to scavenge free radicals in the body [5]. Furthermore, micronutrients such as inorganic ions present in coconut water play a vital role in aiding the human body antioxidant system. Kinetin was shown to act as a strong antioxidant both under in vitro and in vivo from oxidative damage mediated by the Fenton reaction. Kinetin inhibits the formation of 5-oxo-2'-deoxy guanosine, which is a common marker of oxidative damage in DNA. The oxidant properties of kinetin suggested that it may also

present the oxidative damage of unsaturated fatty acids located within the cell membranes [6]. Coconut water or coconut juice is a sweet refreshing drink taken directly from the inner part of coconut fruits (Steiner *et al.*, 2008). It differs from coconut milk, which is the oily white liquid extracted from the grated fresh kernel in most cases, coconut tree plantations more related to garden. As a consequence, the coconut water remains a traditional and water used resource which could thus be considered as an exotic beverage by most people living far from the coconut production area [7]. Coconut water is not only a tropical beverage but also a traditional medicine. A microbiological growth medium and a ceremonial gift [8], and can be processed into vinegar or wine [9]. These various uses are possible thanks to the original biochemical composition of the juice. The particular mineral composition and reasonable total sugar content make coconut water a natural isotonic liquid. The characteristics of coconut water make it an ideal rehydrating and refreshing drink after physical exercise [10].

Consequently, in this present study the *invivo* effects of coconut water were studied in wistar albino rats.

## **2. MATERIALS AND METHODS**

### **2.1 PLANT MATERIAL**

The coconut was purchased from Umuahia market, Abia state, Nigeria and identified by Dr. Garuba Omosun of the Plant Science and Biotechnology Department, Michael Okpara University of Agriculture, Umudike.

### **2.2 EXPERIMENTAL ANIMALS**

Twenty five (25) wistar albino rat weighing (120-140g) from the laboratory animal unit of the department Biochemistry, Michael Okpara university of agriculture umudike was used for this study. They were housed in aluminium cages as five rats per cage and were fed *ad libitum* with standard commercial pelleted growers feed (vital, Nigeria) with free access to clean drinking water. They were kept at normal environmental temperature and natural light/ dark daily cycle. They were maintained in accordance with the recommendation of the guide for the care and use of laboratory animal (DHHS, 1985). They were allowed two weeks to acclimatize before the commencement of the experiment.

### **2.3 CHEMICALS AND REAGENTS**

Hydrochloric acid (HCl), sulphuric acid solution, phosphate buffer, disodium hydrogen phosphate, potassium permanganate, sodium dihydrogen carbonate ( $\text{NaH}_2\text{CO}_3$ ) ethylene diamine tetracetate (EDTA), sodium citrate, sodium hydroxide, trichloroacetic acid (TCA), Thiobarbituric acid (TBA) were purchased from Sigma-Aldrich (Steinheim, Germany). All other chemicals were of analytical grade.

### **2.4 EXPERIMENTAL DESIGN**

Animals were grouped into five animals each

Group A - vital feed alone (control)

Group B - 10ml coconut water + 150g vital feed

Group C - 20ml coconut water + 150g vital feed

Group D - 30ml coconut water+ 150g vital feed

Group E - 40ml coconut water + 150g vital feed

Experimental period lasted for 28 days, after which animals were fasted overnight and blood obtained through ocular puncture. The rats were later euthanized by cervical dislocation and the liver was dissected out immediately for preparation of liver homogenate. Serum was separated by centrifugation at 895\*g 10min and aspirated with sterile plan sample bottles.

## **2.5 EVALUATION OF THE VARIOUS PARAMETERS STUDIED**

### **2.5.1 DETERMINATION OF REDUCED GLUTATHIONE (GSH)**

Reduced glutathione (GSH) was determined by the method of [11].

### **2.5.2 DETERMINATION OF VITAMIN C**

Determination of ascorbic acid was according to the method proposed by [12].

### **2.5.3 CATALASE ASSAY**

Determination of catalase activity was according to [13].

### **2.5.4 DETERMINATION OF SUPEROXIDE DISMUTASE (SOD)**

This was determined using the method [14].

### **2.5.5 DETERMINATION OF MALONDIALDEHYDE (MDA)**

Lipid peroxidation was determined spectrophotometrically by measuring the level of lipid peroxidation product, malondialdehyde (MDA) as described by [15].

## **2.6 STATISTICAL ANALYSIS**

The data were expressed as Mean  $\pm$  Standard error of Mean (Mean  $\pm$  SEM) and presented as figures. Data were analysed using statistical package for the social sciences (SPSS 22.0). Comparison was made between the test groups and the control groups using One way Anova and  $P < 0.05$  were considered statistically significant.

### **3. RESULTS AND DISCUSSION**

**Effects of Coconut water extract on Malondialdehyde (MDA)**

UNDER PEER REVIEW

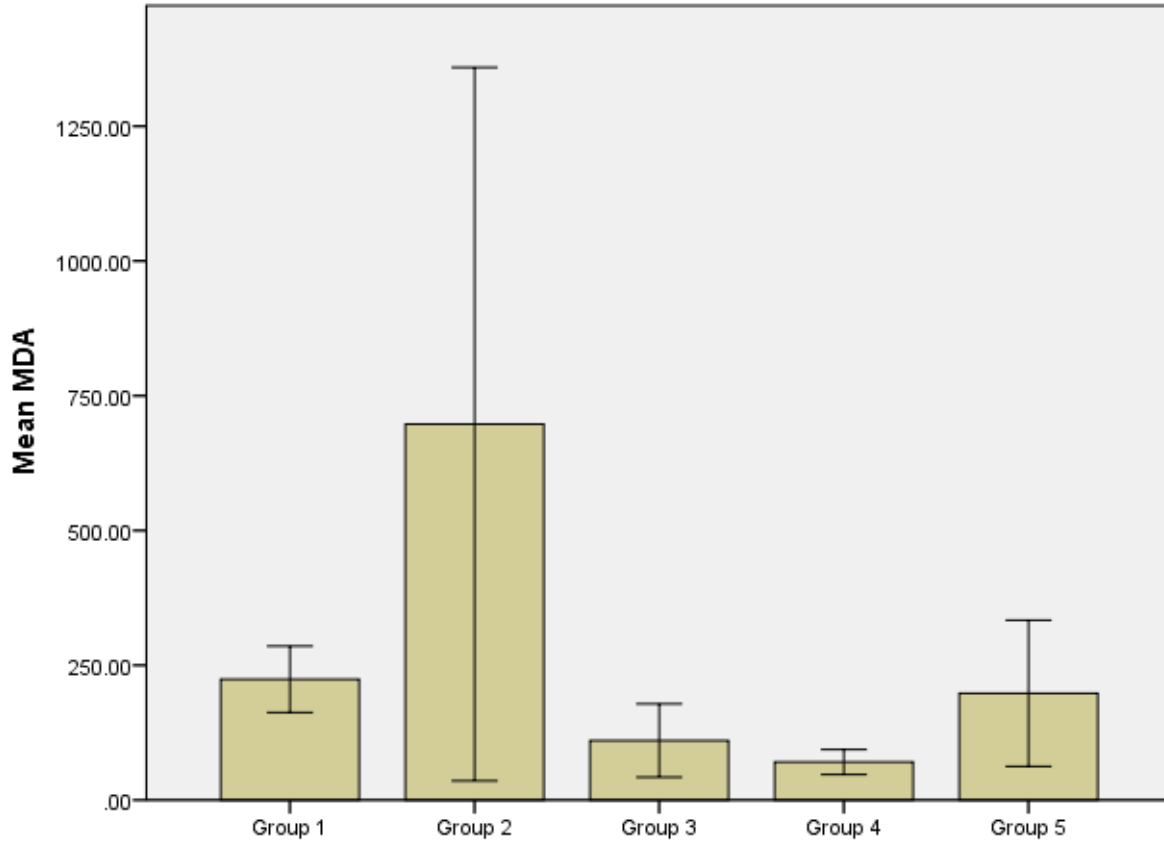


Fig. 1 Mean comparison of the control group (group 1) and the test groups (group 2, 3, 4 and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract for MDA.

The result of the mean comparison of the control group (group 1) and the test groups (Group 3, 4, 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract showed that there is a significant increase ( $p < 0.05$ ) only between the control group and the test group (Group 2) that received 20ml of the coconut water extract.

### **Effects of Coconut water extract on Superoxide dismutase (SOD)**

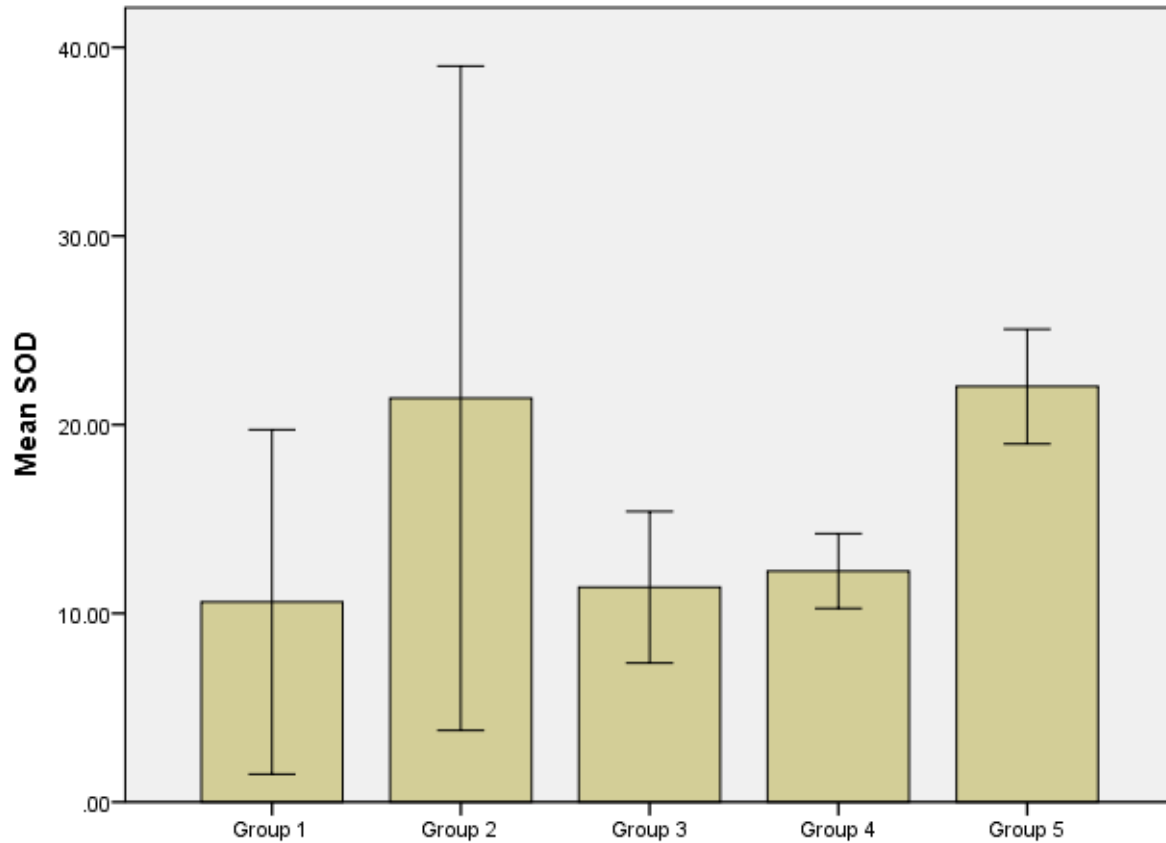


Fig. 2 Mean comparison of the control group (group 1) and the test groups (Group 2, 3, 4 and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract for SOD.

The result of the mean comparison of the control group and the test groups (Group 2, 3, 4,5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract showed that there is a significant increase ( $p < 0.05$ ) between the control group and the test groups that received 10 and 50 ml of the extract.

### Effects of Coconut water extract on Catalase

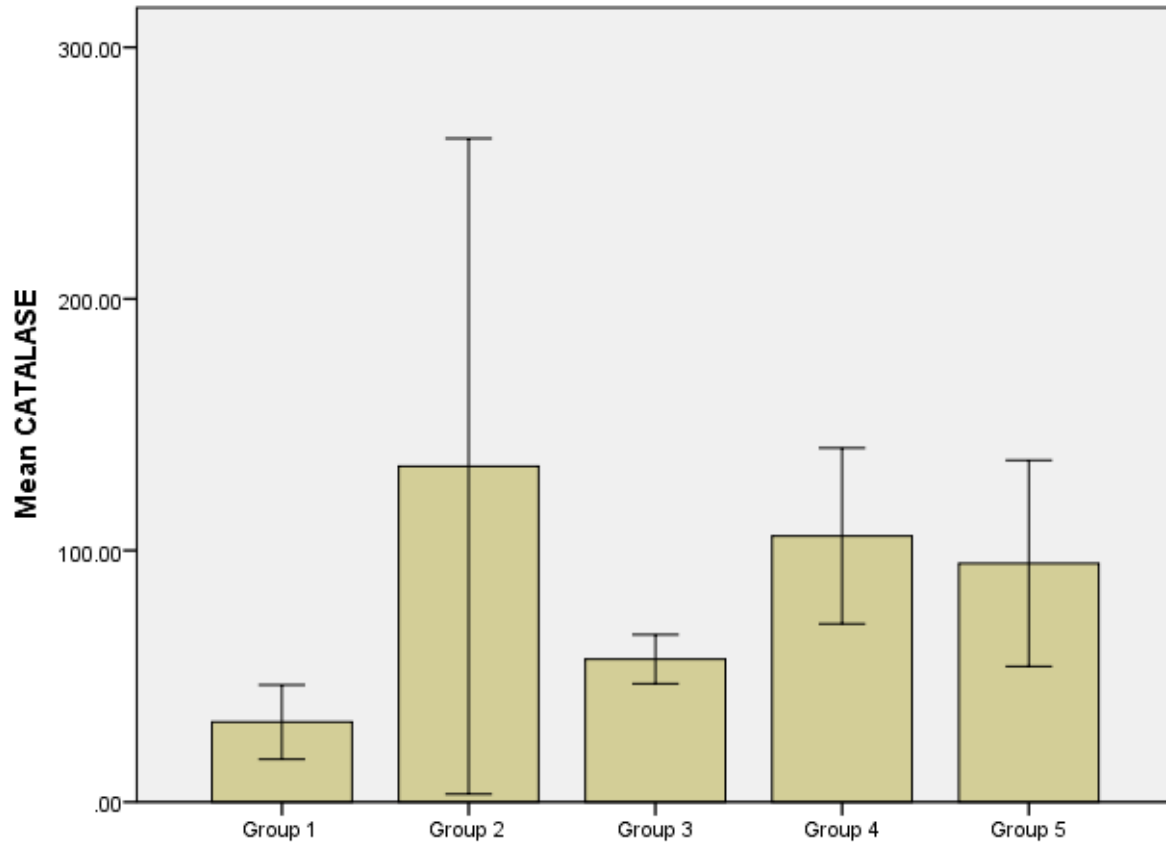


Fig. 3 Mean comparison of the control group (Group 1) and the test groups (Group 2, 3,4, and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract for Catalase (CAT).

The result of the mean comparison of the control group (group 1) and the test groups (Group 2, 3, 4, 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract showed that there is a significant increase ( $p < 0.05$ ) between the control group and the test groups (Group 3, 4, and 5).

**Effects of Coconut water extract on reduced glutathione (GSH)**

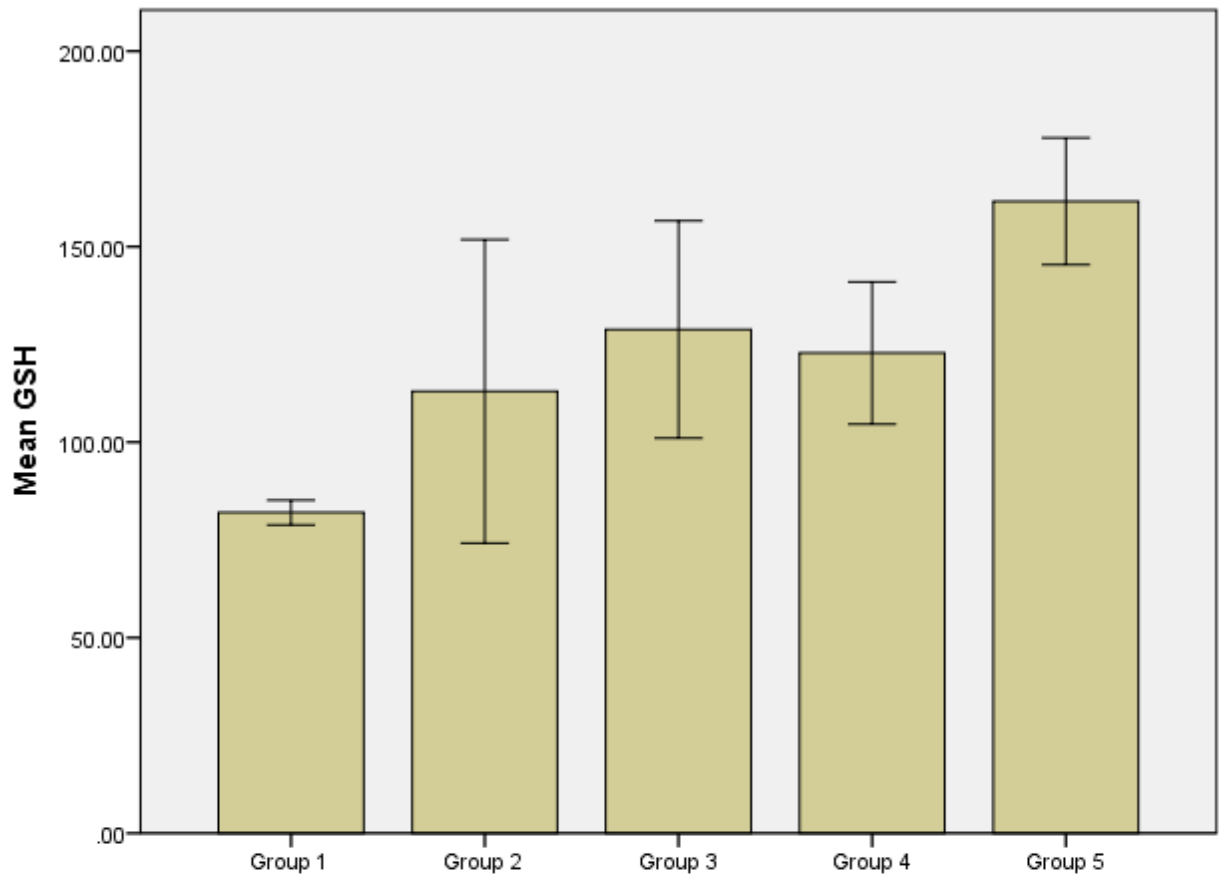


Fig. 4 Mean comparison of the control group (Group 1) and the test groups (Group 2, 3, 4 and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract for reduced glutathione (GSH).

The result of the mean comparison of the control group (group 1) and the test groups (Group 2, 3, 4 and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract showed that there is a significant increase ( $p < 0.05$ ) between the control group and the test groups.

#### Effects of Coconut water extract on Vitamin C



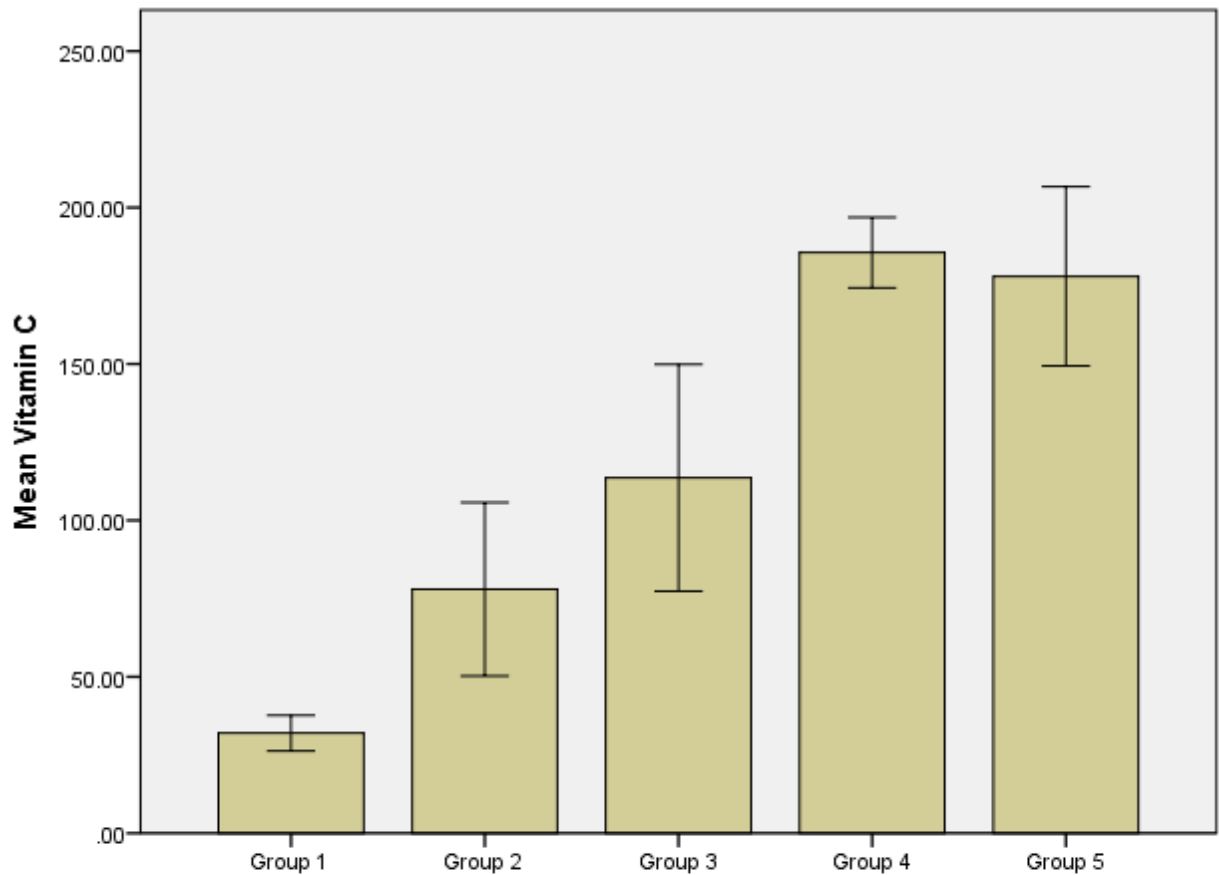


Fig. 5 Mean comparison of the control group (Group 1) and the test groups (Group 2, 3, 4 and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract for Vitamin C.

The result of the mean comparison of the control group (group 1) and the test groups (Group 2, 3, 4 and 5) that received 10ml, 20ml, 30ml and 40ml of the coconut water extract showed that there is a significant increase ( $p < 0.05$ ) between the control group and the test groups.

Antioxidant enzymes (made in the body) and antioxidant nutrients (found in foods) can scavenge and deactivate this reactive free radicals turning them to harmless particles [16]. Improving body antioxidant status is a way to fight against degenerative diseases. This could be achieved by higher consumption of vegetables and fruits [17]. The positive effect attributable to antioxidant is due to the presence of carotenoids, flavonoids, lycopene, phenolics, vitamin C and B-carotene [18]. The effective of the antioxidants usually increases with their concentrations [19].

The effect of the antioxidants usually increases with their concentrations [19]. The effect of the coconut water on some parameters were examined, for the test group that received 10ml, 20ml,

30ml, and 40ml of the coconut water extract in comparison with the mean difference of the control group, there was a significant increase ( $p < 0.05$ ) between the control group and the test groups signifying that the level of these free radical scavengers (MDA, SOD, GSH Catalase, and Vitamin C) increased in the test groups in comparison with the control group, which indicates that the coconut water extract enhances antioxidant activities in wistar albino rats.

Malondialdehyde (MDA) which is the organic compound with formula  $\text{CH}_2(\text{CHO})_2$ . MDA mainly exist in the enol form. MDA results from the lipid peroxidation of polyunsaturated extract)) fatty acids [20]. MDA is the end product of lipid peroxidation and measures free radical generation. Also, there is no significant difference ( $p < 0.05$ ) between the control group and the test group (group 4), for the MDA and SOD, but there is a significant increase ( $p < 0.05$ ) between the control group and the test group for Catalase. SOD which are enzyme that alternately Catalase the dismutation of the superoxide ( $\text{O}_2^-$ ) radical into either ordinary molecular oxygen ( $\text{O}_2$ ) of hydrogen peroxide ( $\text{H}_2\text{O}_2$ ), hydrogen peroxide is also damaging, but less so, and is degraded by other enzymes such as Catalase. The ligands of copper and zinc which are the active sites of SOD or proteins are histidine and one aspartate side chain, one histidine is bound between the two metals. This shows that the coconut water extract has some antioxidant properties that help to detoxifier the effect of some harmful substances such as nitrogen oxides or drugs [21]. Glutathione (GSH) is a tripeptide found in most cells and reacts with the free radicals to protect cells against hydroxyl radical, singlet oxygen and superoxide radical [5]. The activity of GSH increased in the test groups. This shows that the coconut water extract possesses antioxidant properties that helps to stabilize the integrity of cell membrane and also prevent hepatic damage mediated by free radicals [22].

Vitamin C (ascorbic acid) activity in the study showed a dose-dependent increase in the test groups that received 10 ml, 20 ml, 30 ml and 40 ml. Again, indicating the ability of the coconut water extract to act as an antioxidant supplement.

#### **4. CONCLUSION**

The present investigation showed that the coconut water extract increased antioxidant properties in wistar albino rats.

#### **ETHICAL APPROVAL**

All authors hereby declare that “Principles of laboratory animal care” (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee.

## COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

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