Impact of indole-3-butyric acid (IBA) on the root induction of Strawberry tree *(Arbutus pavarii*) culture in vitro

ABSTRACT

The main objective of this study was to clarify the best concentration of the indole-3-butyric acid (IBA) in order to induce the formation of strong roots of the ***Arbutus pavarii*** plant, it is an endangered plant in El-Jabel El-Akhdar- Libya, this study was created to find a protocol for the propagation of ***Arbutus pavarii*** in vitro. We investigated the effect of different concentrations of IBA plant growth regulator on the rooting. *In vitro* germination three weeks old seedlings were transferred to MS roots induction medium which supplemented with different concentrations of IBA (0, 1, 1.5 and 2 mg L^{-1}). the highest response was obtained when transfer the seedlings to the MS medium half strength and supplemented with IBA (0.1 mg L^{-1}) . All the growth indicators [rooting percentage (77%) number, length (7.4 cm) and dry weight (0.0449 g/jar)] that were measured significantly enhanced when using concentration 0.1 mg L^{-1} of IBA.

Key words: Sterilization and germination; *Arbutus pavarii*; root length ; roots dry weight and IBA.

INTRODUCTION

Arbutus pavarii spreads naturally in El-Jabel El-Akhdar region in the north-eastern part of Libya, between latitudes 25° 20° east and latitude 33° 30° north Its presence is concentrated in the northern and central regions, in the valleys, slopes, mountain slopes and lands with shallow or rocky calcareous soils, Arbutus has about 20 species of trees and has different local names in its spread areas [1]. Endemic species from around 4% from the total species of Libyan flora, *A. pavarii* Pamp. (Ericaceae) locally known as ‘‘Shmeri’’ is one of the endemic species in El-Jabel El-Akhdar it is shrubs grows in the El-Jabel El-Akhdar region, mixed with many trees and shrubs such as *Pistacia lentiscus*, *Ceratonia siliqua* L, *Juniperus phoenicea* L, *Quercus coccifera*, *Rhus tripartita* (Ucria)Grande, *Phillyea media* L., *Ziziphus lotus*(L)Desf,[2 and 3]. These trees and shrubs are the main species in the Maquis formation in -Jabel El-Akhdar region, in the form of shrubs dense of branches which is a small tree or a large evergreen shrub with a smooth reddish brown bark and a multi-patterned leaves with Serrate or Entire edge. Flowering occurs in late spring and fruits mature in late summer. The fruit is globose, a many-seeded berry, yellow to orange in colour, turning red when fully ripehence [4]. This plant suffers from increasing degradation due to negative human activities in many of the El-Jabel El-Akhdar areas and agricultural expansion, urbanization, overgrazing, charcoal making In addition to the climatic factors of the fluctuation of rainfall and temperature

fluctuations which are accompanied by dry southern winds caused by physiological diseases due to drought, and whereas *A. pavarii* has never been widely cultivated by afforestation of other species.

The need for the continuous improvement of traits in crop species remains an ongoing effort for crop scientists and farmers. Different plant species have their own set of phenotypes that need to be improved in order to both add nutritional values and enhance economic gains for humankind. The increase in food demand worldwide, associated with unequal distribution, and the disequilibrium in the distribution of wealth has caused an increasingly important pressure on food producers who, in parallel, have increased their requirements for new technologies that allow greater yields and better quality of the products that they offer [4].While at the same time, there has been an increasing consumer led demand for lower environmental damage and greater sustainability in the food production chain. Strawberry is propagated by runners; therefore the health of daughter plant depends on their mother plants. Strawberry is affected by numerous viruses that greatly reduce the yield [5]. The viruses caused the smaller leaves, decreasing the photosynthesis rate and eventually reducing fresh and dry weight. In complex infections (more than one virus), the rate of photosynthesis per unit area also was profoundly reduced [6].

Plant tissue culture (PTC) is a set of techniques for the aseptic culture of cells, tissues, organs and their components like genes and enzymes under defined physical and chemical conditions *in vitro* and controlled environment. PTC technology also explores conditions that promote cell division and genetic re-programming in *in vitro* conditions and it is considered an important tool in both basic and applied studies, as well as in commercial application [7]. PTC techniques have become of major industrial importance in the area of plant propagation, disease elimination, plant improvement, and production of secondary metabolites.

Growth regulators constitute one of the key and more expensive elements used for *in vitro* propagation. That is why; they must be optimized or substituted for more efficient and cheaper bio regulators [8]. Indole-3-butyric acid (IBA) is a plant hormone in the auxin family and is an ingredient in many commercial horticultural plant rooting products.. In plant tissue culture IBA is used to initiate root formation in vitro in a procedure called micropropagation. Micropropagation of plants. the effect of three different auxins, IBA, IAA and NAA were examined to

determine the relative effect of each auxin on root formation. According to the result for the species, IBA was shown to produce a higher yield of roots compared to the other auxins.[9] The effect of IBA is in concurrence with other studies where IBA is the most commonly used auxin for root formation.[10]

Therefore, the aim of this study was to try to Propagation the plant by seed germination using different concentrations of IBA to obtain seedlings to enable them to re-plant them in their natural environment.

MATERIAL AND METHODS

MATERIAL

Fresh **Seed** *A. pavarii* were collected in december month from the outskirts of the city of Al Bayda – Libya. Taxonomist at the Department of Botany Herbarium, Faculty of Science, and Omar Al-Mukhtar University further identified the samples.

METHODS

Preparation of culture media

Half basal of Murashige and Skoog 1962 (MS) [11] salts nutrient medium with vitamins, glycine and supplemented with 30 g L^{-1} sucrose, 0.1 g L^{-1} myo- inositol (Table 1) was used for *in vitro* seedlings germination. While full MS nutrient salt medium was used for rooting media . After supplementation of full MS media with different concentrations (0,1,1.5 and 2 mg L^{-1}) of **indole-3-butyric acid** (IBA) plant growth regulators [for rooting], all cultures pH were adjusted to 5.8 with 1N KOH or 1N HCl, then with 7 g L^{-1} agar prior to autoclaving at 121ºC and 1.2 kg cm^{-2} for 20 minutes. Culture medium was dispensed as 50 ml per jar (350 ml) for *in vitro* seedlings germination, and rooting. All types of culture media were kept for three days under completely darkness for test of contamination.

Tissue culture chemicals **MS** medium and growth regulator **indole-3-butyric acid** were purchased from Sigma company.

Sterilization and germination

Seeds of the *A. pavarii* were washed with running tap water for 30 min. Then they were taken to the laminar air flow cabinet in which they surface sterilized by dipping in 70% (v/v) ethanol for 2 min and rinsed with sterilized distilled water. Further, disinfected with 20 % (v/v) of commercial Clorox (5.25% Cl_2) solution for 15min

(Rabha [12])and rinsed three times with sterilized distilled water. In complete aseptic conditions equal number from sterilized seeds represents were inoculated in culture medium aseptically as six seeds per each. Cultures were maintained under normal condition (16/8 hours light/dark) at 1500 lux using cool white fluorescent lamps and incubated in a controlled growth chamber at 26±1°C.

performed for studying effect of indole-3-butyric acid capacity to enhance rooting on seedling derived *in vitro. In vitro* germination three weeks old seedlings (reached about 5 - 6 cm in height were subjected as a plant materials) which resulted from MS free growth regulators were transferred to MS roots induction medium (R) which supplemented with different concentrations of IBA(0, 1, 1.5 and 2 mg L^{-1}) as follow:

R_0= control (MS free growth regulators)
R_1 = MS + 1 mg L^{-1} IBA
R_2= MS + 1.5 mg L^{-1} IBA
R_3= MS + 2 mg L^{-1} IBA

Each treatment was consisted of 6 replicates (jar) and each replicate contained three seedling. Cultures were incubated in a controlled growth chamber in complete darkness for 3 days at 26±1°C then transferred to normal condition. After 4 weeks from incubation, the number, length (cm) and dry weight (g/jar) for root were recorded.

Table 1: Culture media composition (Murashige and Skoog 1962).

Ingredients	Amount (mg/L)
Macronutrients	
NH_4NO_3	1650.00
KNO_3	1900.00
$CaCl_2.2H_2O$	440.00
$MgSO_4.7H_2O$	370.00
KH_2PO_4	170.00
Micronutrients	
KI	0.83
H_3BO_3	6.20
$MnSO_4.4H_2O$	22.30
$ZnSO_4.7H_2O$	8.60
$Na_2MoO_4.2H_2O$	0.25
$CuSO_4.5H_2O$	0.025
$CoCl_2$	0.025
Iron stock	
$FeSO_4.7H_2O$	27.80
$Na_2.EDTA.2H_2O$	37.30
Vitamins	
Myo-inositol	100.00
Nicotinic acid	1.00
Pyridoxine HCl	1.00
Thiamine HCl	10.00
Glycine	2.00
Sucrose (g)	30.00
Agar (g)	8.00

RESULT AND DISCUSSIONS

This experiment was conducted to provide the *in vitro* growing seedlings needed for present study as explants source. Indole-3-butyric acid in different concentrations was used to study its effect on the roots**.** The effect of MS medium supplemented with different concentrations of IBA (1, 1.5 and 2 mg L^{-1}) on rooting frequency (%), number of roots, root length (cm) and roots dry weight (g/jar) of Arbutus seedlings was investigated. Data tabulated in Table 2 clearly showed that MS medium supplemented with 1 mg L^{-1} IBA (R_1) gave the maximum value for rooting percentage (70%). In addition, there were non-significant differences among R_0, R_2, and R_3 and they recorded the minimum value for rooting percentage (55%).

About number of roots and concerning to Arbutus seedlings , There were clear differences between the treatments used in the study. It was found that MS medium fortified with 1.5 mg L^{-1} IBA(R_2) in the best number of roots as the number of five is the highest compared to the control. On the other hand, the data are also shown in the table 2 the longest root (7.4 cm) recorded with R_1 followed by R_2 medium, which recorded 5.4 cm length. Furthermore, there were non-significant differences among

R_0, and R_3 media they were shorter in length. The highest dry (0.09 g/jar) weight increment was scored on R_1 (0.0449) and R_2 media compared to other used media.

There have been many studies devoted to the cultivation of strawberry trees and the exact propagation has been reported about 30 years ago **[13].** In recent years, many research groups have been involved in establishing reliable regeneration protocols for agronomical important *A. pavarii*, because it would be a primary step to facilitate gene introduction and improvement of the crop. Our aim in the study was to investigate the effect of IBA hormone on root induction in vitro. Through our study of the effect of IBA on the root induction we found that it significantly enhanced the number and length and dry weight in strawberry seedling. In vitro plant regeneration of Strawberry from different parts, has been reported by seeds, leaves, petioles [14], stem [15],stipules [16], and roots[17]. The results in Table 2 showed that the growth and formation of roots were very low in the treatment of control (MS free growth regulators) compared to all other treatments. These findings are somewhat similar to those previously reported by **Ashraf [18]** .Regarding the effect of IBA on the root response, the results indicated that the IBA with (1 and 1.5 mg/L) showed the highest roots response compared to all other treatments. These results do not exactly match what he concluded **[19].** Our results agreed with **Gautam [20]** indicated that the highest root induction frequency obtained was 95.23% on MS medium with IBA at 1.0 mg/l . **Mereti [21]** found that the highest percentages of rooting were achieved in MS medium contained 10 µM IBA (92%) and 10 µM IAA (82%). Additionally, by increasing the concentration of IBA the height of root was decreased. **Haddadi et al [22]** reported that the presence of NAA strength the rooting percentage and root number but the medium without any Auxin had the lower number of root. However, the highest root development was observed in the control treatment. Here it was concluded that the root phenotype (number and length) was diverse as influenced by different Auxin treatments. All different concentrations of IBA (0, 1, 1.5 and 2 mg L^{-1}) induced the root induction in strawberry and significantly differences were observed among treatments in number and length of regenerated seedling.

CONCLUSION

In vitro regeneration of **Strawberry tree *(Arbutus pavarii*)** is a requirement for genetic transformation, which involves induction and development to the whole plant. Several studies have shown that IBA the effects of root induction in plants. This research demonstrated that IBA (0.1mg L^{-1}) was an effective concentration among different other concentrations on root induction.

Table 2: Effect of culture media composition with different concentrations of IBA on rooting percentage number of roots, root length and root dry weight of A. *pavarii* after 4 weeks of culturing and incubation at normal condition.

Parameters / Treatments	rooting percentage (%)	No. of roots	Root length (cm)	Dry weight (g/jar)
R_0= control (MS free growth regulators)	55^b	2^c	3.2^c	0.008^d
R_1 = MS + 1 mg L^{-1} IBA	70^a	3^b	7.4^a	0.0449^a
R_2= MS + 1.5 mg L^{-1} IBA	55^b	5^a	5.4^b	0.0303^b
R_3= MS + 2 mg L^{-1} IBA	55^b	3^b	3.2^c	0.0183^c

Means having the same letters in a column were not significantly different at p<0.05

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